Performance improvement in a supercontinuum

fiber-coupled system for near infrared absorption spectroscopy

SILJE SKEIDE FUGLERUD,^{1,2,*} JONG WOOK NOH,¹ ASTRID AKSNES,¹
 AND DAG ROAR HJELME¹

⁶ ¹Department of Electronic Systems, Norwegian University of Science and Technology, Trondheim, Norway

⁷ ²Department of Endocrinology, St. Olavs University Hospital, Trondheim, Norway

8 *silje.fuglerud@ntnu.no

Abstract: High fidelity near infrared spectroscopy (NIRS) requires high signal to noise 9 ratio (SNR) achieved with costly and often bulky components in benchtop setups. Portable 10 spectrometers are available but lack the accuracy to measure analytes in small quantities in, for example, aqueous samples. Towards high accuracy portable NIRS, we present a fully fiber optic 12 spectroscopy setup with a supercontinuum source in the long-pulse regime (2 ns). The noise 13 sources of the system are studied theoretically and experimentally. The relative intensity noise 14 (RIN) was reduced from typical values up to 6% to less than 0.1% by deploying a balanced 15 detector and averaging. At well-balanced wavelengths, the system without transmission cells 16 achieved an SNR above 70 dB, approaching the shot noise limit. With transmission cells and 17 long-term measurements, the overall SNR was 55 dB. Glucose in physiological concentrations 18 was measured as a model system, yielding a root mean square error of 4.8 mM, approaching the 19 needed accuracy for physiological glucose monitoring. 20

21 © 2022 Optical Society of America

22 1. Introduction

Ouantitative near infrared spectroscopy (NIRS) for measurement of low concentrations of trace 23 components in fluids requires careful configurations and optimization of the instrumentation, 24 including calibration and system stability [1]. Therefore, such applications normally require the 25 use of expensive benchtop systems limiting applications of NIRS outside controlled environments. 26 One way to bring higher accuracy NIRS to field applications could be to use optical fiber-27 coupled broad bandwidth sources with high spectral power density and compact and rugged 28 spectrometers [2] in a fully optical fiber-coupled system. Optical fibers in the near infrared (NIR) 29 range come at a low cost due to their use in telecommunications, and can improve the robustness 30 of a setup by replacing bulk optics and simplifying packaging and assembly. 31

Spatially coherent broad bandwidth sources with high spectral power density are now readily 32 available. Robust and relatively inexpensive supercontinuum (SC) sources are replacing more 33 traditional light sources in a number of applications, like optical coherence tomography (OCT) 34 [3–5] and stimulated Raman scattering (SRS) microscopy [6,7]. Within NIRS, special applications 35 like ultra-high repetition rate absorption spectroscopy have been successful [8–10]. However, 36 utilizing the broad spectrum from SC sources in conventional NIR absorption spectroscopy 37 is challenging due to the high relative intensity noise (RIN) [4, 11]. A low-noise SC source 38 was only recently demonstrated [5] and might be expected to come at a higher cost if reaching 39 commercialization in the near future. Close to shot noise limited detection using commercially 40 available high RIN SC sources might be possible using a balanced detection scheme, i.e. by 41 using a reference arm for an independent measurement of the source noise. However, in practice 42 the difficulty to maintain precise balance of the two arms limits the achievable noise suppression. 43 In the area of Raman spectroscopy, various balancing schemes to account for variations in the 44

sensing arm relative to the reference arm have been developed [7, 12, 13]. This is made possible
by the non-linear interaction and the pump-probe configuration of the systems. Unfortunately,
these methods are not readily available for linear absorption spectroscopy set-ups. The achievable
performance of fully fiber-coupled NIR absorption based sensor systems using SC sources
remains unknown.

Implementing balanced detection schemes over a broad spectral range for optical fiber coupled 50 NIR absorption spectroscopy systems is challenging for a number of reasons including: i) 51 wavelength dependent splitting ratio of beam splitters and couplers, ii) modal interference, or 52 modal noise, in multi mode (MM) optical fiber systems, and iii) path length difference in the 53 reference and sensing absorption cell in cases when the solvent (background medium) has 54 wavelength dependent absorption. The last point is particularly challenging for NIR absorption 55 measurements in aqueous samples. Modal noise of MM fused couplers is an issue if the 56 wavelength resolution required is lower than a few tens of nanometers [14, 15]. MM systems 57 therefore usually rely on bulk optics with low insertion loss to avoid modal noise limitations. 58 Any coupling noise in the absorption cells will also result in modal noise [16–18]. 59

In this work, we will show how a NIRS system based on an SC source, balanced detection and fiber optics can be designed and optimized to meet the required accuracy for many field applications. To our knowledge, this is the first study exploring the various design considerations outlined above for a fully fiber-coupled NIR absorption spectroscopy based sensor system utilizing an SC source. We systematically characterize the noise sources and how they can be reduced.

To quantify the results of design choices, we have performed a case study using a commercial 65 SC source for fiber-coupled NIR absorption spectroscopy in the wavelength range from 1400 nm 66 to 1700 nm, to sense physiological concentrations of aqueous glucose. Combining bright SC 67 sources and fiber optics can be a possible solution towards measuring glucose by NIR spectroscopy 68 in vivo. Despite 40 years of research on non-invasive glucose sensing using NIRS and a trail 69 of startups, a device capable of commercialization has not been obtained, partly due to the low 70 glucose signal hidden within confounding factors [19], and difficulties translating the calibration 71 across patients and different conditions [20]. A solution might be to apply NIRS to measure in 72 a body fluid, such as peritoneal fluid, which is less influenced by confounding factors [21,22] 73 Benchtop spectrometer solutions have obtained the required accuracy [23], but are not useful for 74 in vivo measurements. 75

76 2. Theoretical background

77 2.1. Noise sources and formalism

The noise variance in a laser-system can be defined as

$$\sigma_{\text{tot}}^2 = \sigma_{\text{r+d}}^2 + \sigma_{\text{tec}}^2 + \sigma_{\text{shot}}^2 + \sigma_{\text{ex}}^2.$$
(1)

The first and second term, the receiver and detector noise σ_{r+d}^2 and technical noise σ_{tec}^2 , are independent of the laser power. In publications describing similar applications [4,5], Eq. 1 is 78 79 defined without the term σ_{tec}^2 . The receiver and detector noise, σ_{r+d} , is caused by the electronics 80 in the photodetector and receiver, and quantization error in digitization. The technical noise, σ_{tec} , 81 is caused by slow changes within the system due to temperature and mechanical instabilities and 82 drift. The third term, σ_{shot}^2 , represents the shot noise, and is linearly related to the incoming 83 optical power [24, Ch. 12.2], $\sigma_{\text{shot}}^2 \propto S$, where S is the quantity that corresponds to the light 84 intensity (e.g. measured counts, power, voltage). The shot noise is a fundamental physical limit 85 and can only be reduced by limiting the measurement bandwidth. The fourth term, σ_{ex}^2 , is the 86 excess laser noise, which reflects the source relative intensity noise. The RIN is the normalized 87 $\sigma_{\rm ex}$ expressed by $\sigma_{\rm ex}/\bar{S}$, where \bar{S} is the average of S. The excess laser noise variance is squarely 88 proportional to the power [4]. The magnitude of the RIN is related to the source in use, and is 89

⁹⁰ known to be a large contributor in SC sources. To highlight the dependence on source power, we

91 express the system noise as

$$\sigma_{\rm tot}^2 = a_0 + a_1 S + a_2 S^2, \tag{2}$$

where a_0, a_1, a_2 are coefficients determined by the instrumentation and S is the measured quantity 92 related to the light intensity. The actual observed noise variance depends on the integration 93 time and the number of samples averaged. If the power spectral density is flat, the variance 94 scales linearly with bandwidth. Similarly, if N samples are integrated, the variance scales with 95 1/N [24]. The total relative noise can be expressed as the coefficient of variation (CV = σ_{tot}/S). 96 The constant noise term a_0 encompasses all source power independent contributions, including 97 the readout noise and technical noise sources such as modal noise, mechanical instability and 98 quantization noise. Modal noise in MM fibers is caused by small perturbations of the fiber 99 combined with spatial aperturing of the mode-dependent speckle pattern [17,18]. External factors 100 such as temperature, mechanical movement of the fiber, or air flow conditions can influence the 101 relative phase between excited modes [15]. Using single mode (SM) fibers eliminates modal 102 noise, but with a disadvantage of higher coupling losses due to a smaller core and numerical 103 aperture. The effect of modal noise was investigated experimentally for different conditions 104 by Oliva et al. [25], and the signal to noise ratio (SNR) was decreased approximately 10dB 105 when a mechanical disturbance to the fiber was introduced but depended on the illumination 106 conditions. By applying a mechanical agitator (fiber shaker), the SNR could be improved 5 dB 107 due to averaging across the phase differences between the modes, yet this is not a viable solution 108 for all applications. 109

To utilize absorption spectroscopy sample interaction must take place, in which the light passes through some sort of transmission cell for aqueous samples. Another noise source adding to the constant noise term can be caused by physical misalignment if the parts to the transmission cell are not completely fixed. In a glued holder, mechanical strain in the system can cause small movement of the fibers leading to a change in transmission. For example, if the path length changes with 1 µm, the water absorption will change by 0.04 % (see S1A, Supplement 1).

The quantization error of digitizing a signal can be approximated to $1/2^n$ [26], where n is the 116 number of bits. When measuring a small absorber with a large variation in background such 117 as water, the quantization error is applied to the full water absorption, and the small absorption 118 signal "riding" on this large signal can be lost in the quantization. Using a balanced detector will 119 decrease the background considerably, allowing an order of magnitude higher difference signal 120 gain. By introducing a balanced detector and increasing the signal gain by 10 dB or more, the bit 121 resolution requirement can be reduced. A higher bit resolution increases the analog-to-digital 122 converter (ADC) cost considerably and also constrains the bandwidth resolution. 123

124 2.2. Beer Lamberts law using a balanced detector

Beer-Lambert's law states that there is a linear relation between the absorbance A at a given wavelength and the concentration c of n absorbing analytes in a sample. It can be formulated as

$$A \equiv -\ln\left(\frac{I}{I_0}\right) = \sum_{i=1}^n \alpha_i l = \sum_{i=1}^n \epsilon_i c_i l,$$
(3)

where *I* and *I*₀ are the signal intensity and the reference intensity, α_i is the absorption coefficient for the given analyte, ϵ_i is the molar absorptivity, and *l* is the path length. A balanced detector consists of two inputs with balanced optical paths for sample and reference and outputs a voltage linearly related to the optical power in the two arms, $U_{\text{bal}} = (P_{\text{samp}} - P_{\text{ref}})\Re G$, where \Re , *G* are the responsivity and transimpedance gain for the detector, respectively. The balanced scheme enables low-noise detection of an analyte with orders of magnitude lower absorption coefficient than the solvent, such as glucose. For an ideal balanced detector, the resulting difference signal for an analyte in water becomes (see Supplement 1 S1D for the full calculation for glucose):

$$U_{\text{bal,analyte}} = (\epsilon_{H_2O} f c_A - \epsilon_A c_A) \, l U_{\text{unbal}} \tag{4}$$

for small (mM) concentrations. Here, f is the displacement factor for the reduction in water concentration by the analyte (glucose: [27]). U_{unbal} is the signal measured with optical power only in the reference arm (with water in the transmission cell) and can therefore be written as $U_{unbal} = \exp(-\alpha_{H_2O}l)I_0A_{det}\Re G/2$, A_{det} is the illuminated area on the detector and a 50:50 split is assumed. Thus, the analyte concentration is directly proportional and linear in relation to the measured difference signal, given that the terms stay constant.

141 2.3. Shot noise estimation

The shot noise limit is a useful estimate to obtain a lower bound on the achievable stability. An estimate of the coefficient of variation (CV = σ_{tot}/\bar{S}) originating from the shot noise is

$$CV_{shot} = \sqrt{\frac{4eMGB}{NU_{unbal}}},$$
(5)

where B is the bandwidth, G is the transimpedance gain of the detector, M is the number of 142 sample points acquired per pulse, and N is the number of pulses averaged (details in S1D in 143 Supplement 1). The shot noise limit for a balanced detector is 3 dB higher than the shot noise for 144 a single detector [28] since the balance is the difference between two separate detectors. The 145 gain factor is given as 1×10^3 V/A [29] for the balanced detector in use, but was also estimated 146 to 0.4×10^3 V/A from the optical power and voltage response, which shows that the estimate is 147 somewhat uncertain. We keep the gain estimate at $G = 1 \times 10^3$ V/A to set an upper bound for the 148 shot noise. The estimates assume dependent samples, due to the oversampling of the detector. 149

150 2.4. Reduction in RIN in balanced detector

The common mode rejection ratio (CMRR) of a balanced detector reflects how much of the common noise can be removed. The output signal expressed as a function of CMRR reads [30]

$$U_{\text{bal}} = G(P_{\text{samp}} - P_{\text{ref}}) + \frac{G}{\text{CMRR}} \frac{1}{2} (P_{\text{samp}} + P_{\text{ref}}).$$
(6)

We can express $P_{\text{samp}} + P_{\text{ref}} = P_0$ and $P_{\text{samp,ref}} = C_{\text{samp,ref}}P_0$ representing the splitting ratios. The

ratio between the balanced and unbalanced signal (with power in the sample arm and reference arm blocked) can then be described as

$$\frac{U_{\text{bal}}}{U_{\text{unbal}}} = 2(C_{\text{samp}} - C_{\text{ref}}) + \frac{1}{\text{CMRR}},\tag{7}$$

where we have assumed that the CMRR is large and that the split ratio is close to 1/2. For CMRR= 30 dB [29] and perfect split ($C_{samp} = C_{ref}$), only 3% of the original RIN remains ($\sigma_{bal,50:50} = 0.03\sigma_{unbal}$). For the case to be presented later, some wavelengths have a poorer balance with $C_{samp} = 0.44$, $C_{ref} = 0.56$. With such a balance, the remaining RIN is 31% ($\sigma_{bal,44:56} = 0.31\sigma_{unbal}$). Thus, obtaining a good balance on the full broadband spectrum is important to achieve low noise levels on all wavelengths.

162 2.5. Case study: Stability requirements for glucose measurements

As previously reported in [11], the low absorption coefficient of glucose in the NIR range requires high SNR of a system measuring glucose at physiological levels. Within the first overtone band (1500 nm to 1850 nm) that is the focus of this work, the glucose absorption coefficient is

 $\alpha_g < 1 \times 10^{-4} \text{ mm}^{-1} \text{ mm}^{-1}$. The water absorption coefficient in the same wavelength region is $\alpha_w > 0.2 \text{ mm}^{-1}$. The molar absorptivities for water and glucose from [27] are shown graphically in Figure 1(a). Water dominates as a solvent because the concentration is > 1000× higher than the glucose concentration. In the first overtone band, a change in glucose concentration of 1 mM is a change of less than on average 7.5×10^{-3} % (electrical SNR ~ 82 dB) of the full unbalanced signal, as shown in Figure 1(b). The noise levels should be lower than the signal ($\sigma_0/\bar{S} < 7.5 \times 10^{-3}$ %). The electrical SNR of the photoelectric current or voltage S_{el} is defined as [24, Ch. 18.6]

$$SNR = \frac{\bar{S_{el}}^2}{\sigma^2} \quad or \tag{8}$$

$$SNR_{dB} = 20 \log\left(\frac{\bar{S}_{el}}{\sigma}\right).$$
 (9)

The accuracy required at each wavelength can be relaxed because the measurements are a 163 scan across wavelengths (and time) that can result in correlated measurements. Trend analysis 164 can therefore be utilized. By using dimensionality reduction, a set of spectra for different 165 concentrations can be reduced to N_{ν} orthogonal vectors (latent variables) comprised of p 166 wavelengths [31]. Since the wavelengths p are correlated, a crude estimate of an increase in the 167 accepted variance is p/N_{ν} . The standard deviation should then be lower than $\sigma_{\text{meas}} < \sigma_0 \sqrt{p/N_{\nu}}$. 168 We require a coefficient of variation (CV) less than 0.03 % (SNR ~ 70 dB) applying a number 169 of 60 wavelengths and $N_{\nu} = 3$ which is realistic for the measurement setup to be presented, i.e. 170 giving a 12 dB reduction. 171

Other wavelength regions of interest are the combination band and short-wave infrared 172 (SWIR) band. The SWIR (900 nm to 1450 nm) has been investigated for non-invasive glucose 173 measurements due to favorable tissue light absorption in this wavelength range (therapeutic 174 window), and absorption from glucose has been shown between 1000 nm to 1400 nm [32, 33], 175 but without stating molar absorptivities of glucose for the range. Studies investigating the 176 most informative NIR wavelengths for glucose spectroscopy emphasize the first overtone and 177 combination band [34,35]. The combination band (2050 nm to 2300 nm) has a higher absorption 178 cross section, but will not be included here due to instrumental limitations. This would require 179 use of a photodetector with orders of magnitude higher dark currents [36, 37]. 180



Fig. 1. (a) From literature [27]: The molar absorptivity of water and glucose at 37 °C. The concentration of pure water is more than $1000 \times$ higher than physiological glucose levels. (b) The estimated difference in the signal of the balanced detector for a 1 mm change in glucose concentration for path lengths l = 1 and 2 mm from Eq. 4 with data in (a) as input.



Fig. 2. Schematic of the experimental setup. The components are labeled for easier reference throughout the text.

181 3. Experimental section

182 3.1. Methods

Figure 2 shows the system setup, consisting of a O-switched pumped SuperK Compact SC 183 laser (1) with approximately 2 ns pulse length set to 18.4 kHz repetition rate, paired with an 184 acousto-optic tunable filter (AOTF) (2) with wavelength selection in the NIR range covering 185 1175 nm to 2000 nm and with a spectral bandwidth of 6.4 nm to 19.8 nm. The AOTF can also 186 regulate the laser power amplitude in increments of 1 % of the original power, which is tabulated 187 to 200 mW across the full spectral range. Using the AOTF (set to 100 % transmitted laser 188 power amplitude), we measured 0.09 mW output power for $\lambda = 1315$ nm, with a corresponding 189 linewidth of > 6.4 nm. Four 50:50 splitters (3) were investigated: a MM fused splitter, a SM 190 fused splitter, a MM mirror splitter, and a SM mirror splitter. 191

Two transmission cells (4-1,4-2) were manufactured by gluing two connector ends into bronze sleeves with a machined window that ensured alignment with 2 mm path length (details in S2A, Supplement 1). The splitter and the transmission cells were connected with MM fibers for the MM splitters, and SM fibers for the SM splitters. 1 m long MM fibers were used to collect the light from the transmission cells and were coupled directly to the balanced detector (5).

The signals were received by a balanced detector (5) set to 150 MHz bandwidth and 1×10^3 V/A 197 transimpedance gain. The wavelength range investigated (1175 nm to 1700 nm) was based on 198 the AOTF limitation on the lower bound and the detector responsivity on the upper bound. The signal was digitized using a 500 MS/s 14 bit ADC digitizer card (6) set to measure with 125 MHz 200 bandwidth to match the difference channel from the detector. The combination of the detector 201 and digitizer was based on a holistic evaluation of cost and optimal bandwidth. The optimal SNR 202 of a 2 ns pulse has been estimated to 50 MHz to 100 MHz [38]. The ADC card was connected 203 directly to the pulse trigger from the laser with a BNC cable, and 10 sample points were integrated 204 and recorded from every pulse, as shown in Figure 3. 205

A set of characterization experiments was performed to determine the optimal scanning parameters and to characterize the system, outlined in Table 1. The details about the measurement procedures can be found in Figure 3 and in Section S2A in Supplement 1. U_{unbal} refers to the measurement with one input blocked into the balanced detector (only measure 5-1 or 5-2). The first measurements characterized the system without the transmission cell (component 4).

The case study consisting of two glucose experiments was conducted afterwards (further details in Section S2A, Supplement 1):

Discrete measurements of glucose solutions in phosphate buffered saline (PBS) buffer in sample arm and PBS buffer in reference arm.

Continuous measurements of glucose solutions in PBS buffer (sample arm, PBS buffer in reference arm) pumped in steps as a function of time.



Fig. 3. Acquisition and signal processing. A narrow wavelength range was selected from pulses generated by the SC laser by the AOTF. The pulse was split in two by a fiber splitter and passed through two transmission cells. The pulses were received by a balanced detector and the difference signal was sampled by an ADC card. One value per pulse was obtained by integrating across ten measurement points. One spectral scan was obtained by averaging across 200 pulses, and iterating across 1175 nm to 1700 nm. A final spectrum was generated from the median across six scans to reduce the effect of outliers. On the bottom line, the unbalanced traces after the splitter and transmission cell with water are shown (power adjusted to avoid detector saturation) along with a balanced trace. Details and equipment numbers are shown in Table S4 (Supplement 1).

Component	Purpose	Measurement type	# pulses,	Results
Component	i u pose i i i cusu cinemi (y pe		(# scans)	Results
3. Optical fiber splitters	Critical component,	PMS	200ava x 3	Fig. 4
	select the most stable	ICIVI5	200avg x 5	
1. System w/o balance	Quantify lacer PIN	Std	1000	Fig. 5
w/o transmission cell	Qualitity laser KIN	510	1000	
	Quantify stability improvement,	Std	1000	Fig. 5
5. System w/balanced detector	optimize averaging, Std quantify CV w/avg, Std		700 000	Fig. 6
w/o transmission cell			200avg x 11	Fig. 6
	long-term stability	Allan variance	200avg x 70, 209, 211	Fig. 9
1.,4.,5. System w/balance	Quantify variance full system,	Std	1000	Fig. 7
and transmission cells	including averaging,	Std	200avg x 11	Fig. 7
w/water	estimate noise contributions	Var vs mean	1000	Fig. 8

Table 1. Overview of characterization and measurements. Number of scans is mentioned when average across pulses were employed. If not, the standard deviation was computed across the pulses.



Fig. 4. Example of noise characteristics for the investigated fiber optic splitters (component 3). The optical fibers were moved between three scans obtained for each splitter. The traces show one of the scans, with the mean of the three scans with a three-point moving average subtracted for visibility.

The first experiment aimed at determining precision, whereas the second experiment mimics 217 a physiological situation with fluid constantly changing, without the possibility to take blank 218 samples. A blank spectrum of the buffer solution was subtracted from the discrete glucose 219 measurement to account for experimental drift. For the time series, a blank spectrum of pure 220 buffer was obtained in the beginning and at the end of the series, and a time dependent correction 221 factor assuming linear drift was subtracted for each wavelength from the time series. Prediction 222 models for the two experiments were built using partial least squares regression (PLSR) and 223 cross-validation to obtain the optimal number of latent variables. These chemometric analysis 224 techniques are standard in analysis of NIR spectra and provided in more detail elsewhere [31,39]. 225

226 3.2. Noise characterization

227 Effect of optical splitter and fiber components

The direct signal from the splitters was measured (averaged across 200 pulses) and the fibers 228 going in and out of the splitters were moved between measurements (repeated three times). One 229 of the traces obtained is shown in Figure 4 for all four splitters. The three-point moving mean 230 average of the traces per splitter was subtracted for visibility. The mean rms across the spectra of 231 the three traces in reference to a smoothed mean is shown in Table 2. The modal noise was a large 232 noise contributor for the MM components. The output of the MM fused splitter was extremely 233 unstable, and the difference signal changed with mechanical movements and temperature changes. 234 Although the MM fused splitter transmitted seven times as much light as the other splitters, 235 the instability and the sensitivity to mechanical movement are too high. The challenge using 236 SC sources for this application is not low power (the source was attenuated to avoid detector 237 saturation), but rather high noise levels. 238

	MM fused	MM mirror	SM fused	SM mirror
$\operatorname{rms}(rac{U_{ ext{bal}}-ar{U}_{ ext{bal}}}{U_{ ext{unbal}}})$	1.69 %	1.07 %	0.85 %	0.16 %

Table 2. Root mean square error difference from the mean from three traces where the optical fibers were moved between for each splitter, averaged across all wavelengths.

The fused splitters were also found to be more sensitive to changes in the room temperature and the air condition system flow rate setting than the mirror splitters in long-term experiments. The MM splitters have in common that the trace fluctuates across neighboring wavelengths, whereas both the SM traces are smooth in comparison. The SM fused splitter can be seen to have a baseline shift, whereas the SM mirror splitter is the most stable splitter that was investigated. It is therefore used in the rest of the measurements and characterizations, along with SM illumination fibers between the splitter and transmission cells. An example of a balanced and unbalanced (reference intensity) trace are shown in the bottom line of Figure 3 for the SM mirror splitter coupled directly to the detector, and through the transmission cell with water.

248 Noise suppression by balanced detector

The CV of the pure laser noise measured in one of the arms of the balanced detector was found 249 to be from 2.8 % to 6 % across 1000 pulses as shown in Figure 5(a). This is the pulse-to-pulse 250 variation that is dominated by the RIN and has been reported within similar levels for this laser 251 type [3]. Note that this value is subject to spectral averaging due to the AOTF broadness (up 252 to 19.8 nm), and a narrower filter would give a larger variance per wavelength [4]. With the 253 only measure being the balanced detector, the CV was reduced to between 0.19% and 1.5%, a 254 24 dB reduction for 1550 nm. The measured balanced CV corresponds reasonably well with the 255 predicted CV based on Eq. 7 with the split ratio in Figure 5(b) as input. The predicted CV for 256 the balanced detector does not take the shot noise limit into account, which is a lower bound on 257 the achievable CV. 258



Fig. 5. (a) The measured laser RIN, the predicted standard deviation for the balanced case, the measured balanced standard deviation, and the predicted shot noise limit (Eq. 5). The predicted standard deviation was found using Eq. 7, based on measured split ratios shown in (b) and the measured laser RIN. The fibers were directly coupled from the SM mirror splitter and measured across 1000 pulses.

259 Noise reduction by averaging

If the noise in each sampled signal is uncorrelated, the SNR should increase by \sqrt{N} across N 260 averages. The effect of averaging was investigated across a measurement series including 700 000 261 consecutive pulses recorded per wavelength. The results are shown in Figure 6(a), for 1300 nm, 262 1500 nm and 1600 nm together with a guiding line indicating the \sqrt{N} improvement. The well 263 balanced wavelength 1600 nm deviates from the \sqrt{N} -line around 50 averages, whereas the less 264 balanced 1300 nm starts out with a lower SNR and follows the \sqrt{N} -line almost up to 200 averages. 265 The scan procedure was therefore set to 200 pulses per wavelength per scan. The difference 266 between the measurement and ideal \sqrt{N} line is smaller for 1300 nm than for the more balanced 267 wavelengths because there is more random instability at 1300 nm, which can be improved by 268 averaging. Low frequency noise and technical noise with long correlation times become more 269 important on a longer time-scale. 270



Fig. 6. (a) The improvement by averaging for different wavelengths compared to the \sqrt{N} predicted improvement. (b) CV for the setup directly coupled from the split for the measured laser RIN, the measured CV with a balanced detector, the measured CV acquired with 200 averages, and the predicted shot noise limit (Eq. 5).

The total improvement from the original laser RIN-dominated standard variation in the directly coupled system can be seen in Figure 6(b) including averaging, with the measured CV between 0.018 % and 0.2 %. We obtained 19 dB lower noise at 1550 nm when averaging across 200 pulses, which matches well with a theoretical improvement of 23 dB corresponding to $1/\sqrt{200}$. The mean SNR with averaging was 66 dB.

276 Full system stability

With the introduction of the transmission cell, the CV increased to between 0.02 % and 1 %, as 277 can be seen in Figure 7(a). Two $20\,\mu$ L water drops were placed in the two transmission cells, 278 and measured continuously. The high CV around 1400 nm is due to the high water absorption. 279 The lower CV around 1650 nm can be attributed to a more favorable balance achieved with the 280 water cells in combination with less water absorption than at other wavelengths. The splitting 281 ratios for the transmission cells are shown in Figure 7(b). The split ratio for wavelengths between 282 1400 nm to 1420 nm is inaccurate due to almost zero transmission. The measured standard 283 deviation deviates more from the estimated shot noise limit than the measurements without a 284 water transmission cell. The short-term full system SNR was 58 dB, an 8 dB SNR reduction 285 accompanied by the introduction of the transmission cells and the MM collection fibers compared 286 to the stability for the directly coupled system shown in Figure 6. 28

288 Short-term noise contributions

The variance between consecutive measurements across 1000 pulses at one wavelength was 289 recorded as a function of the input light level, which could be tuned by the AOTF. A polynomial 290 fit was applied to the resulting relation in Figure 8 in order to determine the constant terms (in 291 Eq. 2) for the shot noise and the excess laser noise. Within short time periods, the technical 292 noise does not have a significant contribution, and it has therefore been omitted. The system 293 is dominated by excess laser noise (not shot noise limited), but as the balance improves from 294 1550 nm to 1610 nm (see Figure 7(b)), the system variance approaches the shot noise limit. The 295 SNR of the balanced pulses (no averaging) was up to 49 dB, which is 14 dB lower than that of 296 the estimated shot noise limit from regression. For less balanced wavelengths, the difference was 297 around 18 dB. The estimated shot noise from regression was a few dB higher than the predicted 298



Fig. 7. (a) The CV for the system through the water cell, without balance showing added noise from the transmission cell in addition to the laser RIN, balanced, with 200 averages, and the predicted shot noise limit (Eq. 5). The discontinuity at 1420 nm is due to a change in the laser amplitude settings to avoid detector saturation. (b) The splitting ratios from the full setup with the water transmission cells.



Fig. 8. The variance of the balanced signal through water as a function of the input light level at 1550 nm and 1610 nm with coefficients for the noise terms found by regression.

shot noise level from Eq. 5. The experimentally determined shot noise limit is perceived as more
 accurate, since the estimated limit is based on typical values for the gain and responsivity, which
 might differ in this exact setup.

302 Long-term stability

The long-term stability was investigated by Allan variance analysis. The SM mirror splitter was 303 coupled directly to the balanced detector and scans were continuously recorded with 200 pulses 304 per wavelength and 5 nm increments in three measurement series (one 74 min, two 230 min). The 305 resulting average Allan variance across the three measurement series is shown in Figure 9(a) for 306 four selected wavelengths. For all the wavelengths, there is an improvement in averaging across 307 subsequent scans, and a high degree of stability up to approximately 15 min for most wavelengths. 308 At longer time scales, the Allan variance increases at some wavelengths. The Allan variance 309 for time constant $\tau = 32$ min is shown in Figure 9(b). The long-term stability declines for the 310 wavelengths below 1400 nm and above 1600 nm. The splitter and fibers are optimized at 1550 nm, 311

and although the specified supported wavelengths span down to below 1300 nm, the balance is not 312 as good at these wavelengths as around 1550 nm (see split ratio in Figure 5(b)). The slow change 313 can be due to external factors such as temperature or mechanical changes affecting the splitter 314 characteristics or laser characteristics which have more impact at wavelengths with less optimal 315 balance. The changes in the temperature or laser characteristics over time could also affect 316 polarization and guided modes in the system, which could give small changes in transmission 317 in the splitter or aperture of the detector. On the basis of the poor long-term stability of the 318 lower wavelengths and to reduce acquisiton time, only wavelengths > 1400 nm were included 319 in the following case study. We chose to record the higher wavelengths due to a better balance 320 through the transmission cells (Figure 7(b)). Based on these and similar stability recordings for 321 this system, we included 6 scans with a reduced scan measurement time (40 s) and created the 322 spectrum by taking the median of the 6 consecutive scans on every wavelength. We obtained a 323 more stable signal less influenced by outliers due to technical noise that was not suppressed by 324 averaging consecutive pulses by taking the median instead of averaging. 325



Fig. 9. (a) The mean Allan variance for a selection of wavelengths, together with the raw measurements. (b) The mean Allan variance for all the recorded wavelengths for $\tau = 32$ min, with the three raw traces in the background.

326 3.3. Case study: Glucose measurements

327 Discrete measurements

A prediction model was obtained by using cross-validation and a PLSR model with 3 latent 328 variables. The result of the analysis of the discrete glucose measurements are shown in Figure 329 10(a). A root mean square error of cross validation (RMSECV) of 4.8 mm was obtained. The 330 spectra after median filtering in the spectral domain (three points) and subtracting the previous 331 PBS reference is shown in Figure 10(b). The obtained mean absolute relative difference (MARD) 332 was 24.3 %. The glucose absorption has a similar shape to what was predicted in Figure 1(b). 333 although the main absorption peak is somewhat blue-shifted (~ $1580 \text{ nm} \rightarrow 1610 \text{ nm}$). This 334 discrepancy could be due to the temperature not being identical (20 °C vs. 37 °C in Figure 1(b)) 335 or that assumptions of ideal conditions, such as imperfect split ratios, were not fulfilled for the 336 experimental setup. The ripples across the spectrum are caused by features of the experimental 337 setup and are not glucose features. The SNR of the system was determined to be 55.4 dB. The 338 decrease in SNR compared to the short-term system is likely due to uncompensated drift and 339 mechanical changes due to the sample procedure, but is an in-use bound for the system. 340



Fig. 10. (a) The prediction from a multivariate regression on the measurement series. (b) Examples of difference spectra with glucose concentrations that are used as the input to the model.



Fig. 11. Predicted glucose concentration with solution continuously pumped across the transmission cell to simulate the change in concentration in a body fluid.

341 Continuous measurements

To simulate a glucose measurement series in the human body, a pump was continuously pumping 342 glucose solution to the sample arm of the transmission cell. The result of a prediction model 343 built on the stabilized measurements (4 latent variables) is shown in Figure 11. There is a delay 344 of approximately 10–15 min between setting the glucose concentration on the pump until the 345 solution reaches the transmission cell. An oscillation was observed in the predicted response 346 on several measurement series, which was attributed to imperfect mixing in the tubing and less 347 accurate control with a large plunger and low flow-rates. The perceived signal stability was 348 improved by taking the median across six scans. A three point moving mean was added to 349 the measurement series to remove the oscillations caused by the pump plunger. The deviation 350 from the pumped concentration around 225 min can be caused by the instability of the pumped 351 concentration. Although this measurement series can not quantify the error due to pump and 352 mixing instability, it shows that it is possible to obtain a prediction across a time series of several 353 hours with liquid constantly flowing through the measurement system. 354

355 4. Discussion and conclusion

We have experimentally explored the use of a commercial SC laser source for fully fiber coupled broadband absorption spectroscopy sensing in the NIR range. Introducing a reference arm, balanced detector, and signal averaging, noise suppression of 2 orders of magnitude was achieved, along with a SNR of 66 dB without the transmission cells and a long-term SNR of 55 dB including
 transmission cells and sample exchange. We present a thorough characterization of the system
 and can point to specific limitations and improvements for future use of SC sources in low-noise
 absorption spectroscopy measurement systems:

- A balanced detector is required to reduce the noise levels, especially the effects of RIN
 noise and quantization error on the small signal.
- The stability and reproduciblity of the fiber splitter are of high importance and could be improved further.
- 367 3. The wavelength range is limited to the SM operation of the fibers.
- 4. The transmission cell must be engineered with high precision and extremely stable components.

We use glucose sensing as a model system requiring very high accuracy, and we are able to obtain an accuracy approaching a benchtop free space beam spectrometer, shown in Figure 12.

372 Long-term instability

Although the short-term stability was shown to be acceptable for glucose sensing with averaging, 373 the long-term stability limits the achievable accuracy. The balance was shown to be an important 374 factor, but spectra were also not completely stable on the well-balanced wavelengths. Neither 375 the temperature measured nor low bandwidth reference measurements from one of the arms 376 (to detect larger changes in split ratio) could properly explain the changes, that were slightly 377 different from measurement series to measurement series and looked like an oscillation across the 378 spectrum. It was possible to partially correct for the drift by taking a blank reference spectrum 379 within 15 min. The observed changes with time are likely due to a combination of change in 380 temperature and strain as the experiment progressed and looked similar to previously reported 381 spectral modal noise [18] for a system with few modes interfering. The use of a MM fiber to 382 collect the light may be the cause of this. The ripples across the spectra seen in Figure 10(b) 383 are not due to glucose, but rather noise likely caused by modal instability. In Figure 10(b), the 384 advantage of scanning across several wavelengths is clear, it is easier to mitigate noise using a 385 prediction model that takes into account the broad spectral features. 386

387 System variance and glucose measurements accuracy

With the development of the system, several series of glucose measurements have been obtained 388 on different systems within our group. The root mean square error (RMSE) of these measurements 389 is plotted against the standard deviation of the systems measured in Figure 12. As expected, 390 there is a clear relationship between the noise level and the obtained glucose measurement error. 391 Using an SC source without any reference arm gave a very low accuracy, and RMSE above 392 60 mм. A substantial improvement was obtained by implementing a reference arm. In the first 393 iteration, measurements of the input pulse powers without passing through water as reference 394 were conducted, and the correction was performed by dividing the output pulse power by the 395 input pulse power [11]. In that setup, a relatively slow oscilloscope with 8 bit resolution was 396 used. The system presented here gave a substantial performance improvement compared to that, 397 with RMSE below 5 mм. To compare to the state-of-the-art NIR benchtop spectrometers, we 398 also include the results of a measurement series on a Metrohm benchtop spectrometer with 1 mm 399 cuvettes measured [39]. The error in repetability measurements using the benchtop spectrometer 400 was calculated to 0.003 % based on measurements and noise levels [39, 40]. This is the level 401 that a NIR system should reach in order to have acceptable measurement uncertainty for in vivo 402



Fig. 12. Case study glucose monitoring: Plot of the RMSECV versus the measured standard deviation using different system configurations. The points correspond to: no reference arm for an SC source, a reference arm but unequal split between the arms (details in [11]) and two single detectors, the system presented here, and a benchtop spectrometer (details in [39]).

use. We obtained a SNR of 55 dB for the glucose measurements, which could not resolve 1 mm glucose changes which we estimated would need 70 dB.

The state of the art continuous glucose sensors are mostly electrochemical, one sensor on the market is based on optical readout [23]. The error is most often stated as MARD or points within a consensus error grid. For these sensors, the MARD is commonly around 11 % [41] but has also been reported up to 18 % [42]. With the experimentally obtained MARD of 24.3 % that was obtained in vitro in this study, small improvements are needed before it would be possible to make use of the system for continuous physiological glucose measurements.

411 Comparison to similar work

To the best of our knowledge, this is the first time that an SC source is used in a fully fiber-coupled absorption spectroscopy setup without the more common use of bulk optics [8,9,43–45]. In transitioning to a fully fiber-optic setup, issues arise with the stability that we have characterized and improved upon.

Ultra-high repetition rate absorption spectroscopy applied to gas sensing often labeled 416 supercontinuum broadband absorption spectroscopy (SCLAS) use a dispersive fiber to temporally 417 separate the wavelengths in a pulse [8-10, 44], which allows for rapid measurements as well as 418 improved correction algorithms in post-processing. For narrow linewidth absorbers, a fit to the 419 absorption trace can provide a reference signal and give a stable measurement without a reference 420 arm setup. However, the temporally resolved pulse requires an extremely fast detector and ADC. 421 which increases the cost of such a system. Furthermore, a dispersion compensation module might 422 require several km of dispersion compensating fiber. These systems are not entirely comparable 423 to ours. As Kaminski et al. note [44], the requirement for a broadly absorbing species and narrow 424 absorbing lines also differ, and a balanced cell must be implemented to cope with the high RIN of 425 SC sources for broadly absorbing species such as aqueous samples instead of using a correction 426 algorithm. 427

Guo et al. [46] have proposed a similar system as the one presented, albeit using what appears 428 to be bulk optics coupled transmission cells. They measured glucose concentrations down to 429 5 mg/dL (0.28 mM) with a tailored system centered around an SC source in the combination 430 band, which has been favored alone or in combination with the first overtone band for glucose 431 sensing [34,35]. It is possible to use extended InGaAs sources to reach this wavelength region, but 432 they also have higher noise floors, costs, and require cooling. Unfortunately, this aspect was not 433 discussed in [46]. In comparison, our results were not as accurate (RMSECV 4.8 mM), although 434 the short-term noise levels were comparable to ours and they report an SNR of 47 dB (compared 435 to 55 dB here). The improvement in their results is likely due to the choice of wavelength region. 436

We aimed at using commercially available equipment and had a cost perspective in mind. Guo et al. [46] used a scanning monochromator that was relatively slow, giving a measurement time of 10 min. Using an AOTF in our setup is advantageous because it decreases the acquisition time so that a full scan can be performed within 4 min, including averaging. Their results along with the study presented here, point towards the applicability of SC sources in applications that require extremely low noise such as glucose sensing.

443 Outlook and possibilities

To achieve high stability for accurate measurements, every component must be optimized for the 444 wavelength range: use of SM optical fibers, a stable beam splitter, as stable as possible source, 445 and as accurate as possible balance between the detectors. As was outlined in the theory, a good 446 balance with a CMRR of 30 dB can reduce the noise levels down to 3 % of the original values. It 447 is difficult to achieve a perfect balance including transmission through the sample and reference 448 absorption cells across a wide wavelength range. A balanced detector with a programmable 449 wavelength dependent gain for the two detectors could be a solution to compensate the difference 450 in split values and the differences in the reference and sample cells. The end stability can be 451 further increased with a source with lower RIN. Recent advances in low-noise supercontinuum 452 source development [5] show that the technology could be optimized beyond the instrument used 453 here. With the cost perspective in mind, long pulsed SC lasers are advantageous (a low-noise 454 ps source is 5-10 times the cost of our ns source [3]). The measurement speed can also be 455 increased, as shown by applications of SC lasers applied to gas sensing (SCLAS), where the 456 pulse is dispersed in time [10, 44] and not filtered so that the acquisition times can be on the 457 order of one wavelength in our setup. The viability and cost of such an approach depends on the 458 analyte requirements on wavelength resolution and wavelength range to select an appropriate 459 dispersive element, detector, and ADC. 460

When laser RIN was suppressed, the dominant noise source contributing on a longer time scale 461 was modal noise from the transmission cell and the MM collecting fibers. SM fibers have cores 462 with diameter around 8 µm, which can be extended up to 25 µm for some photonic crystal SM 463 fibers, which make them more difficult to couple into without using free space optics. Strategies 464 using lensed fibers can increase the transmission somewhat [47]. For an even more stable system, 465 replacing the splitter and interaction region by integrated optics could be a potential solution. SM 466 fibers could be used to connect to the integrated optical platform, utilizing a stable splitter [48] 467 and an exposed waveguide to perform evanescent field sensing [49]. 468

⁴⁶⁹ **Funding.** Content in the funding section will be generated entirely from details submitted to Prism.

Acknowledgments. The Central Norway Regional Health Authority is acknowledged for the support
to Silje S. Fuglerud, project number 46055510. The Research Council of Norway is acknowledged for
supporting the Double Intraperitoneal Artificial Pancreas project, project number 248872, and the support
of Jong W. Noh through Lab-on-a-chip Biophotonic Sensor Platform for diagnostics, project number
248869/O70. We thank Jana Jagerska for helpful tips on experimental aspects, detailed discussions, and
research insights.

⁴⁷⁶ **Disclosures.** The authors declare no conflicts of interest.

477 Data Availability Statement. Data underlying the results presented in this paper are available in Ref. [50].

478 Supplemental document. See Supplement 1 for supporting content.

479 **References**

C. Pasquini, "Near infrared spectroscopy: A mature analytical technique with new perspectives – a review," Anal.
 Chimica Acta 1026, 8–36 (2018).

- 482 2. K. B. Beć, J. Grabska, and C. W. Huck, "Principles and applications of miniaturized near-infrared (NIR) spectrometers,"
- 483 Chem. A Eur. J. 27, 1514–1532 (2021).

- M. Maria, I. B. Gonzalo, T. Feuchter, M. Denninger, P. M. Moselund, L. Leick, O. Bang, and A. Podoleanu,
 "Q-switch-pumped supercontinuum for ultra-high resolution optical coherence tomography," Opt. Lett. 42, 4744–4747 (2017).
- M. Jensen, I. B. Gonzalo, R. D. Engelsholm, M. Maria, N. M. Israelsen, A. Podoleanu, and O. Bang, "Noise of supercontinuum sources in spectral domain optical coherence tomography," JOSA B 36, A154–A160 (2019).
- 5. S. Rao D. S., M. Jensen, L. Grüner-Nielsen, J. T. Olsen, P. Heiduschka, B. Kemper, J. Schnekenburger, M. Glud,
 M. Mogensen, N. M. Israelsen, and O. Bang, "Shot-noise limited, supercontinuum-based optical coherence
 tomography," Light. Sci Appl 10, 133 (2021).
- C. W. Freudiger, W. Yang, G. R. Holtom, N. Peyghambarian, X. S. Xie, and K. Q. Kieu, "Stimulated raman scattering microscopy with a robust fibre laser source," Nat. Photonics 8, 153–159 (2014).
- F. Crisafi, V. Kumar, T. Scopigno, M. Marangoni, G. Cerullo, and D. Polli, "In-line balanced detection stimulated raman scattering microscopy," Sci. Rep. 7, 1–8 (2017).
- S. Dupont, Z. Qu, S. Kiwanuka, L. Hooper, J. Knight, S. Keiding, and C. Kaminski, "Ultra-high repetition rate absorption spectroscopy with low noise supercontinuum radiation generated in an all-normal dispersion fibre," Laser Phys. Lett. 11, 075601 (2014).
- 499
 9. N. G. Blume and S. Wagner, "Broadband supercontinuum laser absorption spectrometer for multiparameter gas phase
 500 combustion diagnostics," Opt. Lett. 40, 3141–3144 (2015).
- J. Emmert, N. G. Blume, A. Dreizler, and S. Wagner, "Data analysis and uncertainty estimation in supercontinuum
 laser absorption spectroscopy," Sci. Rep. 8, 1–16 (2018).
- 503 11. S. S. Fuglerud, K. Milenko, R. Ellingsen, A. Aksnes, and D. R. Hjelme, "Feasibility of supercontinuum sources
 504 for use in glucose sensing by absorption spectroscopy," in *Clinical and Preclinical Optical Diagnostics II*, (Optical
 505 Society of America, 2019), pp. 11073–13.
- K. Seto, T. Tsukada, Y. Okuda, E. Tokunaga, and T. Kobayashi, "Development of a balanced detector with biased
 synchronous detection and application to near shot noise limited noise cancelling of supercontinuum pulse light,"
 Revi Sci Instrum 85, 023702 (2014).
- 13. K. Nose, Y. Ozeki, T. Kishi, K. Sumimura, N. Nishizawa, K. Fukui, Y. Kanematsu, and K. Itoh, "Sensitivity
 enhancement of fiber-laser-based stimulated raman scattering microscopy by collinear balanced detection technique,"
 Opt. Express 20, 13958–13965 (2012).
- 14. B. S. Kawasaki, K. O. Hill, and Y. Tremblay, "Modal-noise generation in biconical-taper couplers," Opt. Lett. 6,
 499–501 (1981).
- 15. P. J. Severin and W. H. Bardoel, "Bandwidth and modal noise effects in fused-head-end multimode fiber passive components," J. Light. Technol. 7, 1932–1940 (1989).
- 516 I. S. Das, C. G. Englefield, and P. A. Goud, "Modal noise and distortion caused by a longitudinal gap between two
 517 multimode fibers," Appl. Opt. 23, 1110–1115 (1984).
- I7. Grupp, F., "The nature of the fiber noise with the FOCES spectrograph nature, modeling and a way to achieve S/N
 400," A & A 412, 897–902 (2003).
- 18. C.-H. Chen, R. O. Reynolds, and A. Kost, "Origin of spectral modal noise in fiber-coupled spectrographs," Appl. Opt.
 45, 519–527 (2006).
- 19. S. Delbeck, T. Vahlsing, S. Leonhardt, G. Steiner, and H. M. Heise, "Non-invasive monitoring of blood glucose
 using optical methods for skin spectroscopy—opportunities and recent advances," Anal. Bioanal. Chem. 411, 63–77
 (2019).
- 20. J. L. Smith, "The pursuit of noninvasive glucose: "Hunting the deceitful turkey", seventh edition," (2020).
 Available online: https://www.researchgate.net/publication/343695948_The_Pursuit_
- 527 of_Noninvasive_Glucose_Hunting_the_Deceitful_Turkey_Seventh_Edition (accessed on 528 August 2nd 2021).
- 21. M. K. Åm, K. Kölle, A. L. Fougner, I. Dirnena-Fusini, P. C. Bösch, R. Ellingsen, D. R. Hjelme, Øyvind Stavdahl,
 S. M. Carlsen, and S. C. Christiansen, "Effect of sensor location on continuous intraperitoneal glucose sensing in an
 animal model," PLoS ONE 13 (2018).
- 22. M. K. Åm, A. L. Fougner, R. Ellingsen, D. R. Hjelme, P. C. Bösch, Øyvind Stavdahl, S. M. Carlsen, and S. C.
 Christiansen, "Why intraperitoneal glucose sensing is sometimes surprisingly rapid and sometimes slow: A
 hypothesis," Med. Hypotheses 132, 109318 (2019).
- 23. I. L. Jernelv, K. Milenko, S. S. Fuglerud, D. R. Hjelme, R. Ellingsen, and A. Aksnes, "A review of optical methods
 for continuous glucose monitoring," Appl. Spectrosc. Rev. 54, 543–572 (2019).
- 537 24. B. E. Saleh and M. C. Teich, Fundamentals of photonics; 2nd ed. (Wiley, 2007).
- 25. E. Oliva, M. Rainer, A. Tozzi, N. Sanna, M. Iuzzolino, and A. Brucalassi, "Experimental characterization of modal noise in multimode fibers for astronomical spectrometers," A & A 632, A21 (2019).
- 26. K. Sayood, "Scalar quantization," in *Introduction to Data Compression (Fourth Edition)*, K. Sayood, ed. (Morgan Kaufmann, 2012), The Morgan Kaufmann Series in Multimedia Information and Systems, pp. 251 294, 4th ed.
- 27. A. K. Amerov, J. Chen, and M. A. Arnold, "Molar absorptivities of glucose and other biological molecules in aqueous solutions over the first overtone and combination regions of the near-infrared spectrum," Appl. Spectrosc.
 58 1195–1204 (2004)
- 28. P. C. D. Hobbs, "Ultrasensitive laser measurements without tears," Appl. Opt. 36, 903–920 (1997).
- 546 29. Thorlabs, Balanced Amplified Photodetectors, PDB440A, PDB440A-AC, PDB440C, PDB440C-AC, PDB450A,

- 547 PDB450A-AC, PDB450C, PDB450C-AC, Operation Manual.
- 30. R. Pallás-Areny and J. G. Webster, "Common mode rejection ratio in differential amplifiers," IEEE Trans Instrum
 Meas 40, 669–676 (1991).
- M. Goodarzi, S. Sharma, H. Ramon, and W. Saeys, "Multivariate calibration of NIR spectroscopic sensors for continuous glucose monitoring," TrAC Trends Anal. Chem 67, 147 – 158 (2015).
- 552 32. K. J. Jeon, I. D. Hwang, S. J. Hahn, and G. Yoon, "Comparison between transmittance and reflectance measurements
 553 in glucose determination using near infrared spectroscopy," J. Biomed. Opt. 11, 014022 (2006).
- 33. W. Yang, N. Liao, H. Cheng, Y. Li, X. Bai, and C. Deng, "Determination of NIR informative wavebands for transmission non-invasive blood glucose measurement using a fourier transform spectrometer," AIP Adv. 8, 035216 (2018).
- J. Chen, M. A. Arnold, and G. W. Small, "Comparison of combination and first overtone spectral regions for nearinfrared calibration models for glucose and other biomolecules in aqueous solutions," Anal. Chem. 76, 5405–5413
 (2004), PMID: 15362899.
- 35. M. Goodarzi and W. Saeys, "Selection of the most informative near infrared spectroscopy wavebands for continuous
 glucose monitoring in human serum," Talanta 146, 155–165 (2016).
- 562 36. K. Linga, G. Olsen, V. Ban, A. Joshi, and W. Kosonocky, "Dark current analysis and characterization of 563 $\ln_x Ga_{1-x} As/InAs_y P_{1-y}$ graded photodiodes with x > 0.53 for response to longer wavelengths (> 1.7 µm)," 564 J. Light. Technol. **10**, 1050–1055 (1992).
- 37. C. Besikci, "Extended short wavelength infrared FPA technology: status and trends," in *Quantum Sensing and Nano Electronics and Photonics XV*, vol. 10540 M. Razeghi, G. J. Brown, J. S. Lewis, and G. Leo, eds., International
 Society for Optics and Photonics (SPIE, 2018), pp. 110 125.
- 38. R. M. Kurtz and T. C. Forrester, "Optimizing reception bandwidth of a pulsed signal," in *Laser Radar Technology and Applications XXV*, vol. 11410 M. D. Turner and G. W. Kamerman, eds., International Society for Optics and
 Photonics (SPIE, 2020), pp. 77 95.
- 39. S. S. Fuglerud, R. Ellingsen, A. Aksnes, and D. R. Hjelme, "Investigation of the effect of clinically relevant interferents on glucose monitoring using near-infrared spectroscopy," J. Biophotonics 14, e202000450 (2021).
- 40. Metrohm, *NIRS XDS RapidContent Analyzer, Manual.*
- 41. G. Freckmann, M. Link, U. Kamecke, C. Haug, B. Baumgartner, and R. Weitgasser, "Performance and usability of three systems for continuous glucose monitoring in direct comparison," J. Diabetes Sci. Technol 13, 890–898 (2019).
- 42. R. Z. Jafri, C. A. Balliro, F. El-Khatib, M. M. Maheno, M. A. Hillard, A. O'Donovan, R. Selagamsetty, H. Zheng,
 E. R. Damiano, and S. J. Russell, "A three-way accuracy comparison of the Dexcom G5, Abbott Freestyle Libre
 Pro, and Senseonics Eversense continuous glucose monitoring devices in a home-use study of subjects with type 1
 diabetes," Diabetes Technol. & Ther. 22, 846–852 (2020).
- 43. R. Watt, C. Kaminski, and J. Hult, "Generation of supercontinuum radiation in conventional single-mode fibre and its application to broadband absorption spectroscopy," Appl. Phys. B **90**, 47–53 (2008).
- 44. C. Kaminski, R. Watt, A. Elder, J. Frank, and J. Hult, "Supercontinuum radiation for applications in chemical sensing and microscopy," Appl. Phys. B 92, 367–378 (2008).
- 45. M. P. Halloran, J. Choi, T. Lee, and J. Yoo, "High-speed supercontinuum laser absorption spectroscopy of light
 hydrocarbons in elevated pressures and temperatures," Meas. Sci Technol 32, 095502 (2021).
- 46. K. Guo, R. A. Martinez, M. Freeman, H. S. Gurm, and M. N. Islam, "High SNR glucose monitoring using a SWIR
 super-continuum light source," in *Conference on Lasers and Electro-Optics*, (Optical Society of America, 2016), p.
 AM4J.5.
- 47. S. S. Fuglerud, K. B. Milenko, R. Ellingsen, A. Aksnes, and D. R. Hjelme, "Glucose sensing by absorption
 spectroscopy using lensed optical fibers," Appl. Opt. 58, 2456–2462 (2019).
- 48. D. González-Andrade, C. Lafforgue, E. Durán-Valdeiglesias, X. Le Roux, M. Berciano, E. Cassan, D. Marris-Morini,
 A. V. Velasco, P. Cheben, L. Vivien, and C. Alonso-Ramos, "Polarization-and wavelength-agnostic nanophotonic
 beam splitter." Sci. Rep. 9, 1–9 (2019).
- 49. E. Ryckeboer, R. Bockstaele, M. Vanslembrouck, and R. Baets, "Glucose sensing by waveguide-based absorption
 spectroscopy on a silicon chip," Biomed. Opt. Express 5, 1636–1648 (2014).
- 50. S. S. Fuglerud, "Performance improvement in a supercontinuum fiber-coupled system for near infrared absorption
 spectroscopy," https://doi.org/10.18710/IZ63JH (2021).