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Modelling temperature-driven changes in species associations across freshwater communities

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Abstract

Due to global climate change-induced shifts in species distributions, estimating changes in community composition through the use of Species Distribution Models has become a key management tool. Being able to determine how species associations change along environmental gradients is likely to be pivotal in exploring the magnitude of future changes in species' distributions. This is particularly important in connectivity-limited ecosystems, such as freshwater ecosystems, where increased human translocation is creating species associations over previously unseen environmental gradients. Here, we use a large-scale presence-absence dataset of freshwater fish from lakes across the Fennoscandian region in a Joint Species Distribution Model, to measure the effect of temperature on species associations. We identified a trend of negative associations between species tolerant of cold waters and those tolerant of warmer waters, as well as positive associations between several more warm-tolerant species, with these associations often shifting depending on local temperatures. Our results confirm that freshwater ecosystems can expect to see a large-scale shift towards communities dominated by more warm-tolerant species. While there remains much work to be done to predict exactly where and when local extinctions may take place, the model implemented provides a starting-point for the exploration of climatedriven community trends. This approach is especially informative in regards to determining which species associations are most central in shaping future community composition, and which areas are most vulnerable to local extinctions.

KEYWORDS

climate change, co-occurrence, fish, JSDMs

1 | INTRODUCTION

Due to global trends in species distribution range shifts and biodiversity loss, ecosystems worldwide are likely to undergo considerable changes in community composition (Jennings & Harris, 2017; Seebens et al., 2020). Along with the increased spread of nonnative species as a result of globalisation, increasing average temperatures due to climate change will enable new species to make their way into environments previously too cold for them to either enter or establish in (Rahel & Olden, 2008; Sharma et al., 2007;

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Walther et al., 2005). Many of these species are capable of causing extirpations of native species or even the restructuring of entire food webs once established (Nackley et al., 2017; Rockwell-Postel et al., 2020; Walther et al., 2009). Many species are vulnerable to local population declines and extinctions as increasing temperatures and extreme weather events compound threats posed by other anthropogenic factors, among them biological invasions (Dawson et al., 2011).

A changing climate is also capable of changing associations between species. While many species may be capable of co-occurring at certain temperatures, as annual temperatures increase, one species may gain a competitive advantage. For instance, at moderate temperatures a beech forest may contain three or four species, yet as temperatures tend to either extreme, negative associations may occur in the form of a single species beginning to outcompete the others and dominate (Leathwick, 2002). Associations becoming more negative with temperature increases could lead to declines in some species' populations, and local extinctions occurring, well before these species' upper thermal tolerances are reached. A deeper understanding of changes in species associations over a temperature gradient would contribute greatly to our understanding of the likely effects of climate change on community composition (Early & Keith, 2019; Freeman et al., 2018).

Climate change is likely to affect freshwater ecosystems particularly harshly, with a rise in temperature likely to lead to a population increase in species with higher thermal tolerance, with species of lower tolerance shifting further upstream (Comte et al., 2013; Daufresne & Boët, 2007). Increases in human translocations over recent decades are leading to novel species associations (Carpio et al., 2019), making understanding the impacts of these associations particularly important to predict future ecosystem effects. Research in sub-Arctic regions - which are likely to warm substantially in the coming decades - has already demonstrated a shift towards more warm-adapted species both within individual lakes and across catchments. (Hayden et al., 2017; Sharma et al., 2007; Van Zuiden et al., 2015; Winfield et al., 2008). Northern pike (Esox Lucius; Linnaeus, 1758) and brown trout (Salmo trutta; Linnaeus, 1758) may co-occur at lower temperatures in the sub-Arctic, but as average annual temperatures increase, a negative association

results in a drop in the brown trout population as pike begin to predate brown trout at higher rates, eventually leading to local brown trout extinctions (Hein et al., 2013). The accelerated nature of climate change in the Arctic and sub-Arctic means that these regions are particularly important in giving an insight into potential community shifts, which are likely to take place in the coming decades in warmer regions.

Although much of the current research on shifts in associations over a temperature range concerns only a few species, or at a relatively small scale, attempts to increase focus on broader community models in predicting the impacts of climate change at a larger scale are ongoing (Comte et al., 2013; Radinger et al., 2019; Silknetter et al., 2020). Here, we construct a Joint Species Distribution Model (JSDM) as proposed by Tikhonov et al. (2017) to predict trends in species associations across a temperature gradient on a multi-national scale. JSDMs arose as a combination of habitat modelling and community ecology, in response to the need to account for associations which are not explained by the effects of environmental covariates (Ovaskainen et al., 2017: Pollock et al., 2014; Warton et al., 2015). Although they are capable of estimating correlations between species, these correlations may be a product of shared habitat specialisation, not interactions (Hargreaves et al., 2020; Hayden et al., 2013). As such, the output of JSDMs are often referred to as representing species associations, as opposed to interactions (Blanchet et al., 2020).

Here we use a dataset containing presence-absence data of fish species across more than 3000 freshwater lakes in the region of Fennoscandia, covering large climate gradients across mainland Norway, Sweden and Finland (Tammi et al., 2003). Using this modelling framework we aim to assess (a) whether we are able to identify the changes in associations between species of different thermal tolerances over a temperature gradient at large scales, (b) the reliability of our predictions based on whether these associations corroborate the results of previous research into pairwise associations and (c) how accurately these models can be used to predict future community shifts in the face of climate change. We aim to provide ecological modellers with a promising framework to build upon when exploring shifts in species distributions and abundances and provide managers with key information regarding potential extinction and invasion hotspots going forward.

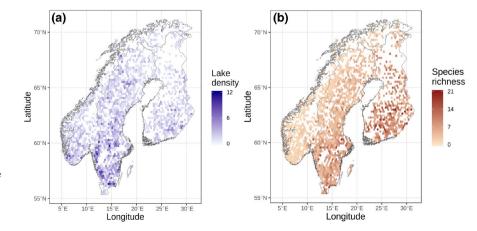


FIGURE 1 (a) Density of the 3308 lakes surveyed in the 1995 Nordic Fish Status Survey, represented by the number of lake centrepoints contained within hexagons. (b) Distribution of freshwater fish species richness across the same lakes, with species richness equal to average species richness of lakes found within hexagons

2 | METHODS

2.1 | Study system

Our study system was a series of 3308 lakes throughout the Fennoscandian region (Norway, Sweden and Finland). The lakes were located between 55.4 degrees and 71.1 degrees in latitude, 4.6 and 31.4 degrees longitude (WGS84), and at an altitudinal range of 0 to 1540 m. Surveyed lake density is highest throughout Sweden (Figure 1a).

The region itself is particularly species poor, given relatively recent deglaciation (Huitfeldt-Kaas, 1918). Species richness increases eastwards, with Finland and Sweden showing higher species richness than western Norway (Figure 1b). This is a product of mountainous regions dividing Norway, which have provided fewer immigration pathways for freshwater fish. Studies in the region have predicted that native species belonging to warmer guilds ('cool-water species'), like the Northern pike, the European perch (Perca fluviatilis; Linnaeus, 1758), and cyprinids like the common roach (Rutilus rutilus; Linnaeus, 1758) (Elliott, 2010; Hayden et al., 2014; Hokanson, 1977; Wehrly et al., 2003) will expand their range (Comte et al., 2013; Hayden et al., 2017). This could result in the local extirpation of species like the brown trout, whitefish (Coregonus lavaretus; Valenciennes, 1848) and Arctic charr (Salvelinus alpinus; Linnaeus, 1758), which are more tolerant of cold waters ('cool-cold/cold-water species'; Elliott, 2010; Hayden et al., 2014; Parkinson et al., 2016; Wehrly et al., 2003). The co-occurrence of species from various thermal guilds, within Fennoscandia, makes the region ideal for studying the impacts of climate change on freshwater communities (Comte et al., 2013).

2.2 | Occurrence data

The occurrence data were collected from a Fennoscandian survey of freshwater fish, originally conducted between 1995 and 1997 (Tammi et al., 2003). The resulting dataset consists of presence-absence information on fish species in 3821 lakes across Fennoscandia (Table 1). Henceforth, all species will be referred to by their common name (Table 1). Species that appeared in less than 1% of total lakes were excluded, as it is unlikely species-specific parameters can be accurately estimated given such low levels of occurrence. None were species thought likely to have a large-scale negative impact on Fennoscandian freshwater communities, either through competition or direct predation (for the full list of species, see Appendix S1).

2.3 | Environmental data

Temperature data were derived from the EuroLST data set for the centre of each lake (Metz et al., 2014). The temperature covariate used was the average air temperature of the warmest quarter, which has shown to have a strong correlation with water lake temperature

(Livingstone & Lotter, 1998). This was the available data most likely to be strongly correlated with ice-off dates and spring warming temperatures, both of which have been shown to have a strong effect on both the life histories of aquatic species and interactions between different species (Mehner et al., 2011; Munsch et al., 2019).

For each lake, we obtained six additional covariates describing either environmental properties or human impact with the potential to influence establishment risk. Our study focuses on the effect of temperature; however, other environmental covariates are likely to have a strong effect on community composition, and as such are included as covariates. Environmental properties included lake surface area, shoreline complexity, total area of lakes situated upstream of focal lake, water pH, total organic carbon and human impact at the site. Area, shoreline complexity, and total upstream area were all derived from GIS analyses. Shoreline complexity was then calculated as:

$$SC = \frac{P}{2000\sqrt{\pi A}}$$

where P is the lake perimeter and A is the lake surface area (Wetzel, 2001). Water chemistry covariates were taken from the 1995 Nordic Lake Survey (Henriksen et al., 1998). To approximate human impact on each site, we used the Human Footprint Index as compiled by Venter at al. (2016), henceforth referred to as HFI. HFI is a point score which combines eight human impact covariates to approximate the level of human pressure on nature, assigned to cells one kilometre squared in size (Venter et al., 2016). These impact covariates include presence of built environment, crop lands and roads, and local human population density. HFI was taken for the cell in which the centre point of the lake lay, with previous research suggesting that human activity in the immediate vicinity of freshwater sites is more likely to affect species occurrences than activity upstream or downstream (Chapman et al., 2019). Lakes for which any of the environmental data was incomplete were disregarded (n = 119, 3.5% of total lakes). Area, total upstream area and total organic carbon were heavily right skewed and were thus log-transformed to assist with model convergence. All covariates were standardised by scaling to a mean of zero and a standard deviation of one to assist in model convergence. Further information on all covariates, included expected effects on populations, are summarised in Table 2.

The immigration history of freshwater fish in the area and steep topography that makes up much of Norway's west coast mean that many species have historically been unable to naturally colonise this region (Figure 1b; Huitfeldt-Kaas, 1918; Sandlund & Hesthagen, 2011). As such, environmental conditions which would normally result in the presence of species across other parts of Fennoscandia may have little to no effect on the likelihood of their presence in this region. This spatial divide could potentially make for a source of spatial autocorrelation, so to account for this we used the historical distribution range of species which were unable to colonise Norway's west coast as an additional covariate (see Appendix S2). PERRIN ET AL.

TABLE 1 Freshwater fish species surveyed in the 1995 Nordic Fish Status Survey

Common name	Scientific name	Family	Naming authority	Frequency of occurrence (%)
Perch	Perca fluviatilis	Percidae	Linnaeus, 1754	72.1
Pike	Esox lucius	Esocidae	Linnaeus, 1754	65.8
Roach	Rutilus rutilus	Cyprinidae	Linnaeus, 1754	52.9
Brown trout	Salmo trutta	Salmonidae	Linnaeus, 1754	46.7
Burbot	Lota lota	Lotidae	Linnaeus, 1754	37.8
Bream	Abramis brama	Cyprinidae	Linnaeus, 1754	24.5
Whitefish	Coregonus lavaretus	Salmonidae	Valenciennes, 1844	23.5
Ruffe	Gymnocephalus cernuus	Percidae	Linnaeus, 1754	21.0
Arctic charr	Salvelinus alpinus	Salmonidae	Linnaeus, 1754	14.9
Bleak	Alburnus alburnus	Cyprinidae	Linnaeus, 1754	13.7
Tench	Tinca tinca	Cyprinidae	Linnaeus, 1754	13.5
Vendace	Coregonus albula	Salmonidae	Linnaeus, 1754	12.1
Zander	Stizostedion lucioperca	Percidae	Linnaeus, 1754	11.7
Crucian carp	Carassius carassius	Cyprinidae	Linnaeus, 1754	11.0
Rudd	Scardinius erythrophthalmus	Cyprinidae	Linnaeus, 1754	10.1
Minnow	Phoxinus phoxinus	Cyprinidae	Linnaeus, 1754	9.2
Smelt	Osmerus eperlanus	Osmeridae	Linnaeus, 1754	8.1
White bream	Blicca bjoerkna	Cyprinidae	Linnaeus, 1754	6.5
Grayling	Thymallus thymallus	Salmonidae	Linnaeus, 1754	6.1
Ide	Leuciscus idus	Cyprinidae	Linnaeus, 1754	6.0
Rainbow trout	Oncorhynchus mykiss	Salmonidae	Walbaum, 1792	4.1
Threespine stickleback	Gasterosteus aculeatus	Gasterosteidae	Linnaeus, 1754	1.6
Brook trout	Salvelinus fontinalis	Salmonidae	Mitchill, 1814	1.4
Ninespine stickleback	Pungitius pungitius	Gasterosteidae	Linnaeus, 1754	1.0

Note: Table shows species taxonomy, as well as percentage of 3308 lakes that the species were found in. Species in bold occurred in more than 10% of lakes and were, thus, considered high-occurrence species.

2.4 Statistical modelling

We constructed a JSDM which predicts changes in species associations over a temperature gradient. For our matrix of recorded observations, we assume that the presence-absences of species j = 1...p at lakes i = 1...n are independent observations, conditional on a vector of $h = 1...n_{f}$ latent factors per lake, modelled as:

$$y_{ij} \sim \text{Bernoulli}(p_{ij}),$$

with
$$p_{ij} = \varphi^{-1}(\eta_{ij})$$

where p_{ij} denotes the probability of species *j* being present at site *i*, and φ^{-1} denotes the inverse of a probit link-function. We denote η_{ij} as:

$$\eta_{ij} = \alpha_j + \sum_{k=1}^{n_c} x_{ik} \beta_{jk} + \varepsilon_{ij}$$

where x_{ik} denotes the value of environmental covariate $k = 1...n_c$ at site *i*, α_i denotes the intercept for species *j*, and β_{ik} denotes the effect

of environmental covariate k on species j. The species-by-site random effects $\epsilon_{ii} \sim N(0, R(x_i))$ are defined by a latent factor model:

$$\varepsilon_{ij} = \sum_{h=1}^{n_{\rm f}} z_{ih} \lambda_{jh} \left(x_i^* \right),$$

where z_{ih} denotes our $h = 1...n_f$ latent factors for lake *i*, where $\lambda_{ih}(\mathbf{x}_i^*)$ denotes the responses (loading) of species j = 1...p to each of the latent factors. We model the loadings per species *j* and for each factor *h* as a function of temperature:

$$\lambda_{jh}\left(x_{i}^{*}\right) = \sum_{i=1}^{n_{l}} \tau_{jh} + x_{i}u_{jh},$$

where τ_{ih} denotes an additional intercept for species j and latent factor h, u_{ih} denotes the response of latent factor h for species j to temperature, x_i denotes the temperature in degrees at site *i*, and n_i denotes the total number of sites. Here, we assume $n_f = 3$, as our Deviance information criteria did not improve significantly with the addition of more latent factors, and a few major gradients usually account for

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TABLE 2 Environmental covariates, description, environmental effects, units and mean (±standard deviation) used in Joint Species Distribution Models of freshwater fish across European freshwater lakes

Environmental covariate	Description	Expected biological effect	Unit	Mean (±SD)
Area	Surface area of lake	Larger area increases potential habitat and niche breadth	Square kilometres	6163 (±52149)
Shoreline complexity	Calculated using area and perimeter	Increased shoreline complexity creates variation in habitat type (Verdiell-Cubedo et al., 2012)	Unitless	0.20, 0.14
Temperature	Average surface air temperature during maximum quarter	Temperature may alter various life history aspects of species (Magnuson et al., 1979)	Degrees	12.75, 1.74
Human Footprint Index	Index comprising 10 different variables, which represents impact of human activity (Venter at al., 2016)	Higher HFI increases chances of local human introductions (Chapman et al., 2019)	Unitless scale from 1 to 50	6.79, 7.03
Total upstream area	Aggregated area of lakes occurring directly upstream from focal lake	Higher upstream area increases chance of species' persistence	Square kilometres	388.68, 6405.17
рН	Taken from Nordic Lake Survey (Henriksen et al., 1998)	Acid sensitivity can limit local species' distributions (Ohman et al., 2006)	Unitless scale from 1 to 14	6.62, 0.66
Total organic carbon	Taken from Nordic Lake Survey (Henriksen et al., 1998)	Higher levels can cause anoxia and limit species' distributions (Ohman et al., 2006)	Mg per litre	7.09, 4.95
Biogeographic zone	Whether or not lake was found in a drainage basin cut-off from the rest of the region by the natural dispersal barrier running through central Norway	Presence of dispersal barrier provides fewer immigration pathways into western Norway for species, which did not colonise area via Norwegian Sea (Sandlund & Hesthagen, 2011)	Binary covariate	NA

most inter-species variation (Halvorsen, 2012). We define the matrices $\Lambda(\mathbf{x}_i^*)$ with elements $\lambda_{jh}(\mathbf{x}_i^*)$ and use these to construct a temperaturedependent, inter-species residual covariance matrix:

$$\Omega\left(\mathbf{x}_{i}^{*}\right) = \Lambda\left(\mathbf{x}_{i}^{*}\right)\Lambda\left(\mathbf{x}_{i}^{*}\right)^{T} + \mathbf{I}.$$

We then scale this covariance matrix to an inter-species correlation matrix *R* representing temperature-dependent associations between species that are not explained by fixed species-specific effects of environmental covariates:

$$R_{j_1j_2} = \Omega_{j_1j_2} / \sqrt{\Omega_{j_1j_1}\Omega_{j_2j_2}}$$

These resulted in values between -1 and 1, with positive values indicative of positive associations between species, implying that species are likely to co-occur, and negative values implying the opposite. To compare species associations to the similarity in species

responses to fixed effects in the models, we calculated an additional correlation matrix (Hui, 2017).

Although temperature was included in the random-effect, it was also included as a fixed-effect with quadratic function, to account for potential non-linear responses of species to temperature (Boddy & McIntosh, 2017; Veen et al., 2021).

We fit the model in a Bayesian framework using the greta R-package (Golding, 2019). All parameters were specified noninformative normally-distributed priors with a mean of zero and a standard deviation of 10, with the exception of the latent factors z_{ih} (mean = 0, SD = 1) and alpha parameters α_j (mean = -2, SD = 1). Alpha parameters had a lower mean to assist with convergence, as most species were prevalent at a low number of locations. Further exceptions were u and τ matrices, for which the diagonals had positively truncated non-informative normal priors, and all values in the upper triangle, which were set to zero to enforce identifiability (Hui et al., 2015). Markov Chain Monte-Carlo (MCMC) sampling was done using 4000 samples on one chain, with a burn-in of 2000 samples. Although most Bayesian analyses would use multiple chains, latent variable models are often invariant to sign-switching (Hui, 2017), so here we choose to only run one chain. We used a Hamiltonian Monte Carlo sampler, sampling the number of leapfrog steps at each iteration uniformly between 40 and 60 (these numbers were manually tuned to achieve efficient sampling). The leapfrog integrator step sizes for each parameter were automatically tuned during the burn-in phase, then fixed for sampling. Parameters were considered to have adequately converged if their Geweke *Z*-score was below 1.96 (Geweke, 1992). 96 of our 105 species association parameters converged adequately. Trace plots for all association parameters can be found in Appendix S3.

We estimated changes in species associations over a continuous gradient from 6.68 to 16.80 degrees Celsius, which represented the minimum and maximum temperature observed in the data. Three models were constructed. Model 1 included data from all 24 species ("all species" model). Model 2 included data from species which only occurred in more than 10% of lakes (henceforth referred to as high-occurrence species, which are indicated as bold names in Table 1), as we wanted to test whether the inclusion of low-occurrence or low-detectability species produced a better model fit for commonly occurring species. Of the 24 species included in this study, 15 were classified as high-occurrence, and were thus used in model 2 (the "reduced species" model). To test whether accounting for species associations over a temperature gradient improved model fit, model 3 accounted for species associations, but not over a temperature gradient ("base JSDM" model). In this model, ε_{ij} is defined as:

$$\varepsilon_{ij} = \sum_{h=1}^{n_{\rm f}} z_{ih} \lambda_{jh}.$$

Model fit was quantified using the Bernoulli deviance D_i, where

$$D_{j} = -2 \times \sum_{i} \left(y_{ij} \log \left(\mu_{ij} \right) + \left(1 - y_{ij} \right) \log \left(1 - \mu_{ij} \right) \right),$$

which was calculated for each high-occurrence species *j* of each model using the posterior medians of p_{ij} . To ensure that our models were an improvement over single species distribution models (SDMs), we created a stacked species distribution model (SSDM) consisting of single species distribution models for the fifteen species with occurrences in over 10% of lakes. These were also probit models with a Bernoulli distribution and used the same environmental covariates as our three previous models, but they did not include latent factors to account for associations between species. Model fit between the three models was compared using the improvement in deviance D_j from the SSDM for each of the high-occurrence species.

All statistical analyses were completed using R version 3.4.4 (R Core Team, 2017) and RStudio (RStudio Team, 2020). Additionally, the following R-packages were used for analysis and visualisation; dplyr (Wickham et al., 2019), rgeos (Bivand & Rundel, 2019), spdep (Bivand & Wong, 2018), postGIStools (Marchand & Ellison, 2019), tensorflow (Allaire & Tang, 2019), corrplot (Wei & Simko, 2017), ggplot2 (Wickham, 2016), magrittr (Bache & Wickham, 2014) and gridExtra (Auguie, 2017). A comprehensive definition of the model, the code, and its analysis can be found at Perrin (2021) (https://doi. org/10.5281/zenodo.4665778).

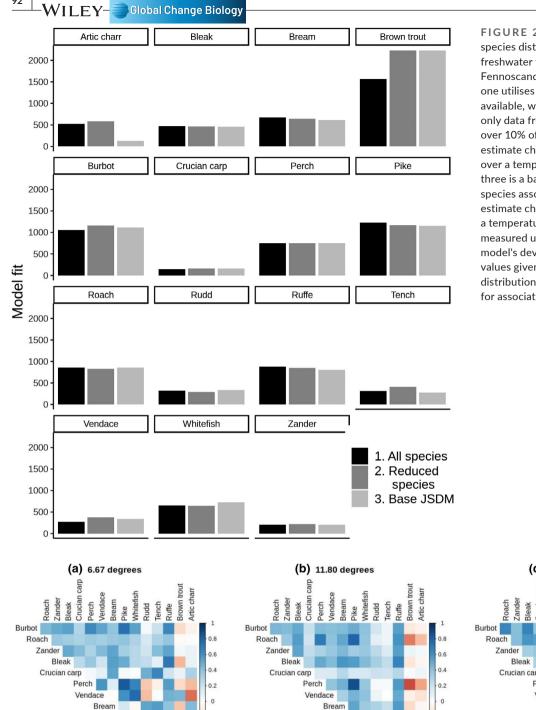
3 | RESULTS

Our results captured variations in species associations across a temperature gradient, with many negative associations between coldwater species and those tolerant of warmer temperatures.

Model fit was relatively similar across the all species model, reduced species model and base JSDM for each species (Figure 2). The only notable exceptions were brown trout, tench and arctic charr, for which the reduced species model (model two) had better model fit than either one of or both the other models. As such, the reduced species model was used for further analysis, as computing time was considerably lower. Biogeographic zone did not have a significant effect on any species and did not affect species associations, and as such the covariate was removed from all models.

Although several species associations at lower temperatures were negative - indicating a low likelihood of co-occurrence - at the mean and higher temperatures most associations between species were positive or close to neutral (Figure 3). The majority of negative associations between species at mean or high temperatures occurred between the cold-water species (brown trout or Arctic charr) and other species, with the most negative associations occurring between these species and those classified as belonging to a higher thermal guild (roach, perch and pike). Correlations in response to aggregated environmental variables were positive between the majority of species, with the exception of Arctic charr, for which many correlations were negative (Figure 4a). Correlations between species in responses to temperature were stronger than responses to all environmental variables, with a more even mix of positive and negative correlations (Figure 4b). Correlations in responses to each environmental covariate can be found in Appendix S4, as can species individual responses to environmental covariates.

As previous research has suggested potential associations between commonly occurring cool-water species (pike, perch, roach, whitefish) and cold-water species (brown trout and Arctic charr), these associations are shown in more detail in Figure 5, with thermal guild classifications found in Table 3. The association between brown trout and Arctic charr was negative at lower temperatures, becoming gradually positive before peaking at the mean temperature and decreasing as temperatures further increased. Arctic charr associations with perch, pike and roach were all negative at the lowest temperature, remained such until the mean temperatures. Arctic charr associations with whitefish increased consistently from being negative at the lowest temperature, before levelling out and remaining neutral at higher temperatures. Brown trout associations with perch, pike and roach all followed similar patterns, with



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FIGURE 2 Model fit for three joint species distribution models mapping freshwater fish associations across 3308 Fennoscandian freshwater lakes. Model one utilises data from every species available, whereas model two utilises only data from species which occurred in over 10% of lakes. Models one and two estimate changes in species associations over a temperature gradient. Model three is a basic JSDM which monitors species associations, but does not estimate changes in associations over a temperature gradient. Model fit was measured using the improvement of each model's deviance values over the deviance values given by a stacked species distribution model which did not account for associations between species

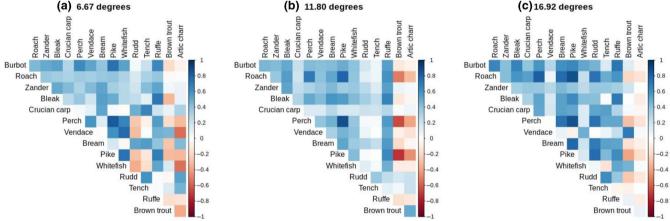
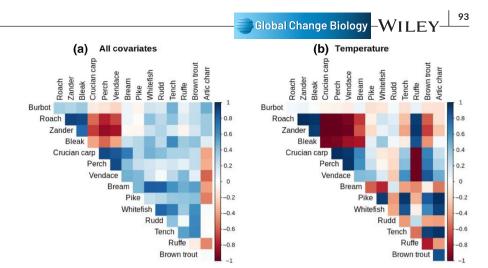


FIGURE 3 Estimates of associations between 15 different freshwater fish species across freshwater lakes in 3308 lakes across the Fennoscandia region at different temperatures. Associations are shown for the region's (a) minimum (6.67 degrees), (b) mean (11.80 degrees) and (c) maximum (16.92 degrees) temperatures. Temperatures used represent average surface temperature during the warmest quarter of the year. Associations vary between 1 (most positive) and -1 (most negative)

associations close to neutral at the lowest temperature, becoming negative at the mean temperature and re-ascending towards zero as temperatures further increased. Associations between whitefish and brown trout were weakly positive at low temperatures, decreasing

to weakly negative at the mean temperature before becoming more positive at higher temperatures. Whitefish associations with pike, perch and roach were positive at lower temperatures, and became weaker (although still positive) at higher temperatures. Associations

FIGURE 4 Correlation due to shared environmental response of 15 different fish species across 3308 freshwater lakes in Fennoscandia. Figures show correlation due to response to (a) all environmental covariates used in the Joint Species Distribution Model and (b) average surface air temperature of the warmest quarter of the year. Associations vary between 1 (most positive) and -1 (most negative)



between perch, pike and roach were all strongly positive, with little variation across temperature.

Maps visualising modelled predictions of likely changes in freshwater species distributions can be seen in Appendix S5.

ily equate to a demonstrable negative impact in this study unless one species were driven to local extinction, unlike when fitting a latent variable model to abundance data.

significant impacts on habitat use or life-history would not necessar-

4 | DISCUSSION

The ability to predict changes in species associations over environmental gradients will be crucial to incorporate into species distribution modelling as climate change modifies temperatures (Freeman et al., 2018). Here, we quantify changes in species associations over a climate gradient on a multi-national scale, using a presenceabsence data set comprising 3308 lakes to fit a series of JSDMs of different freshwater fish species.

In accordance with Tikhonov et al. (2017) we demonstrate that large-scale presence-absence data are capable of shedding light on species associations over environmental gradients. Species belonging to cold-water guilds are generally negatively associated (and thus have a low likelihood of co-occurrence) with species from warmer thermal guilds, and these associations may change as temperatures warm.

Our confidence in these results is boosted by the fact that many of the associations predicted by our model corroborate previous research performed on pairwise associations. The negative associations between the two cold-water species and the cool-water species included here have been observed on smaller scales in this and similar study regions (Byström et al., 2007; Hayden et al., 2017; Hein et al., 2013; Winfield et al., 2008). Likewise, some of the positive associations shown among cool-water species here also have historical precedence (Eklöv & Hamrin, 1989; Mills & Hurley, 1990; Sharma & Borgstrøm, 2008).

A positive association between two species does not imply the lack of a negative impact of one species on another. Our results indicate a positive association between whitefish and perch, despite past evidence suggesting that whitefish are negatively impacted by the presence of perch (Hayden et al., 2013). However, it is possible for the two species to co-occur, e.g. through niche segregation (Hayden et al., 2014). However, since our response variable is binary, It is important to note that predictions of species associations may become uninformative at certain temperatures. For example, when temperatures reach levels that preclude a species occurring in that region at all, any effect of species associations in an environmental context becomes void (Tikhonov et al., 2017). This is reflected in the associations between some species of different thermal guilds, which are predicted to increase towards zero as temperatures reach the higher ends of the spectrum. Summer temperatures in the region's warmer lakes are higher than the temperature range of lakes typically occupied by cold-water species (Mandeville et al., 2019). As such our predictions of associations between species should only be considered reliable at temperature ranges where both species are capable of persisting independently.

Although such models are capable of estimating future shifts in community composition in response to climate change, we recommend instead treating estimates provided by such models as indications of potential larger trends and – similar to Wagner at al. (2020) – as a basis for generating hypotheses and focussing future research (Zurell et al., 2020). Although some of the associations here match previous research, others point to new potential threats to native cold-water species like the Arctic charr and brown trout, which should be studied more thoroughly. Although it was beyond the scope of this paper, further research could also take into account possible interactions between temperature and other environmental covariates, for instance habitat area, as research has suggested that often colder-tolerant species can withstand potential competition if there is enough available habitat and niche segregation within a given habitat patch (Hein et al., 2013).

These results show that on a broad, multi-national scale, shifts towards communities dominated by species which have higher thermal tolerance are likely to occur as climate change drives average temperatures higher. Although many lakes may not reach the thermal maximum of native species, our model confirms that local extinctions are likely to occur earlier, driven by changing associations between native species and either invasive non-native species,

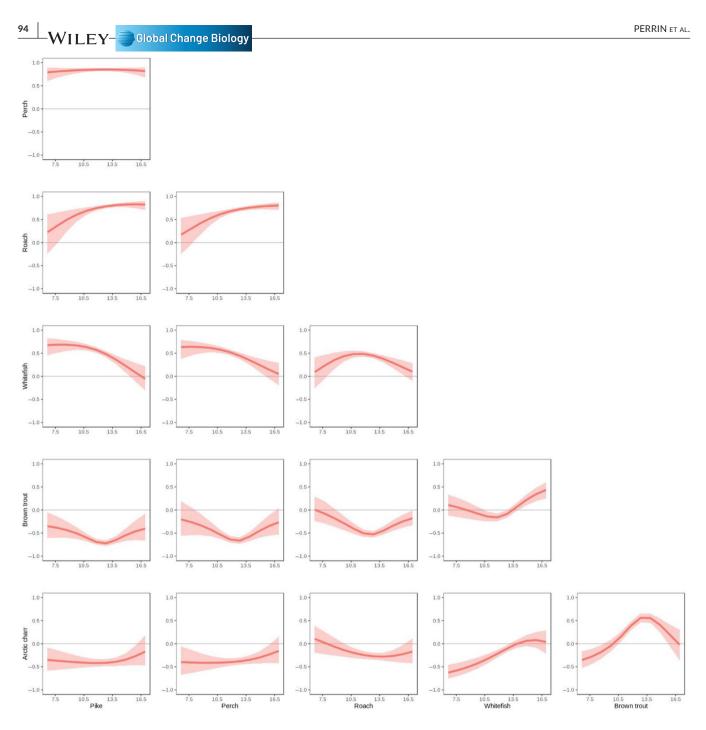


FIGURE 5 Associations between six freshwater fish species over a temperature gradient inferred from residual correlations from a joint species distribution model across 3308 freshwater lakes in the Fennoscandian region. Temperatures displayed on *x*-axes represent average surface temperature during the warmest quarter of the year. Associations are displayed on *y*-axes on a scale from -1 to 1, with ribbons representing 95% credible intervals. Negative values represent negative associations between species

Common name	Thermal guild	References
Perch	Cool	Hayden et al. (2014); Hokanson (1977)
Pike	Cool	Wehrly et al. (2003)
Roach	Cool	Elliott (2010)
Whitefish	Cool-cold	Hayden et al. (2014)
Brown trout	Cold	Elliott (2010); Wehrly et al. (2003)
Arctic charr	Cold	Elliott (2010)

TABLE 3 Thermal guild classification of six freshwater fish species surveyed in 1995 Nordic Fish Status Survey, with references citing precedent for classification range shifting species or species with which native fish had previously co-occurred. The tendency of many species towards positive associations with an increase in temperature suggests a trend towards homogenisation of freshwater communities, though a variation in species individual responses to increased temperature could affect this.

Previous research indicates that more cold-tolerant species are capable of persisting in larger, deeper lakes due to the possibility of spatial segregation (Hein et al., 2013). Areas with strong topographical variation could provide dispersal barriers for novel species, and subsequently provide refugia for species likely to be outcompeted (Perrin et al., 2020). However such refugia are only likely to be tenable if human translocation is sufficiently regulated so as to prevent the introduction of novel species (Hesthagen & Sandlund, 2004; Perrin et al., 2021). As such, models like the one constructed here could enable researchers not only to identify environmental covariates which may drive changes in species are vulnerable to local extinctions, but also to identify areas where such species are likely to persist.

The ability of large-scale SDMs to predict changes in species associations while corroborating smaller-scale pairwise research over a temperature gradient is encouraging. It implies that largescale presence-absence data may be capable of predicting changes in community composition as temperatures increase in the coming decades. Although much work remains to ensure the accuracy and reliable management application of such models, our results here indicate that JSDMs can be used to identify the potential impacts of climate change and range-shifting species on global ecosystems.

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CONFLICT OF INTEREST

No conflict of interest for the article.

DATA AVAILABILITY STATEMENT

Data and code used for the species distribution modelling are archived in Zenodo at http://doi.org/10.5281/zenodo.4665778.

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REFERENCES

- Allaire, J. J., & Tang, Y. (2019). tensorflow: R interface to "TensorFlow". https://github.com/rstudio/tensorflow
- Auguie, B. (2017). gridExtra: Miscellaneous functions for "grid" graphics. https://CRAN.R-project.org/package=gridExtra
- Bache, S. M. & Wickham, H. (2014). magrittr: A forward-pipe operator for R. https://CRAN.R-project.org/package=magrittr
- Bivand, R. & Rundel, C. (2019). rgeos: Interface to Geometry Engine -Open Source ('GEOS'). https://CRAN.R-project.org/package=rgeos
- Bivand, R. S., & Wong, D. W. S. (2018). Comparing implementations of global and local indicators of spatial association. TEST, 27, 716–748. https://doi.org/10.1007/s11749-018-0599-x
- Blanchet, F. G., Cazelles, K., & Gravel, D. (2020). Co-occurrence is not evidence of ecological interactions. *Ecology Letters*, 23, 1050–1063. https://doi.org/10.1111/ele.13525
- Boddy, N. C., & McIntosh, A. R. (2017). Temperature, invaders and patchy habitat interact to limit the distribution of a vulnerable freshwater fish. *Austral Ecology*, 42(4), 456–467. https://doi.org/10.1111/aec.12463
- Byström, P., Karlsson, J., Nilsson, P., Van Kooten, T., Ask, J., & Olofsson, F. (2007). Substitution of top predators: Effects of pike invasion in a subarctic lake. *Freshwater Biology*, 52(7), 1271–1280. https://doi. org/10.1111/j.1365-2427.2007.01763.x
- Carpio, A. J., De Miguel, R. J., Oteros, J., Hillström, L., & Tortosa, F. S. (2019). Angling as a source of non-native freshwater fish: A European review. *Biological Invasions*, *5*, 3233–3248. https://doi. org/10.1007/s10530-019-02042-5
- Chapman, D. S., Gunn, I. D. M., Pringle, H. E. K., Siriwardena, G. M., Taylor, P., Thackeray, S. J., Willby, N. J., & Carvalho, L. (2019). Invasion of freshwater ecosystems is promoted by network connectivity to hotspots of human activity. *Global Ecology and Biogeography: A Journal of Macroecology*, 88, 528.
- Comte, L., Buisson, L., Daufresne, M., & Grenouillet, G. (2013). Climateinduced changes in the distribution of freshwater fish: Observed and predicted trends. *Freshwater Biology*, 58(4), 625–639. https:// doi.org/10.1111/fwb.12081
- Daufresne, M., & Boët, P. (2007). Climate change impacts on structure and diversity of fish communities in rivers. *Global Change Biology*, 13(12), 2467–2478. https://doi.org/10.1111/j.1365-2486.2007.01449.x
- Dawson, T. P., Jackson, S. T., House, J. I., Prentice, I. C., & Mace, G. M. (2011). Beyond predictions: Biodiversity conservation in a changing climate. *Science*, 332(6025), 53–58.
- Early, R., & Keith, S. A. (2019). Geographically variable biotic interactions and implications for species ranges. Global Ecology and Biogeography: A Journal of Macroecology, 28(1), 42–53. https://doi. org/10.1111/geb.12861
- Eklöv, P., Hamrin, S. F., & Eklov, P. (1989). Predatory efficiency and prey selection: Interactions between pike *Esox lucius*, perch *Perca fluviatilis* and rudd *Scardinus erythrophthalmus*. *Oikos*, 56(2), 149–156. https://doi.org/10.2307/3565330
- Elliott, A. (2010). A comparison of thermal polygons for British freshwater teleosts. *Freshwater Forum*, 5. https://core.ac.uk/download/ pdf/228601267.pdf
- Freeman, B. G., Lee-Yaw, J. A., Sunday, J. M., & Hargreaves, A. L. (2018). Expanding, shifting and shrinking: The impact of global warming on species' elevational distributions. *Global Ecology and Biogeography:* A Journal of Macroecology, 27(11), 1268–1276.
- Geweke, J. (1992). Evaluating the accuracy of sampling-based approaches to the calculations of posterior moments. *Bayesian Statistics*, *4*, 641-649.
- Golding, N. (2019). greta: Simple and scalable statistical modelling in R. Journal of Open Source Software, 4(40), 1601. https://doi. org/10.21105/joss.01601
- Halvorsen, R. (2012). A gradient analytic perspective on distribution modelling. *Sommerfeltia*, 35(1), 1–165. https://doi.org/10.2478/ v10208-011-0015-3

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- Hargreaves, A. L., Germain, R. M., Bontrager, M., Persi, J., & Angert, A. L. (2020). Local adaptation to biotic interactions: A meta-analysis across latitudes. *The American Naturalist*, 195(3), 395–411. https:// doi.org/10.1086/707323
- Hayden, B., Harrod, C., & Kahilainen, K. K. (2014). Lake morphometry and resource polymorphism determine niche segregation between cool-and cold-water-adapted fish. *Ecology*, 95(2), 538–552. https:// doi.org/10.1890/13-0264.1
- Hayden, B., Holopainen, T., Amundsen, P. A., Eloranta, A. P., Knudsen, R., Præbel, K., & Kahilainen, K. K. (2013). Interactions between invading benthivorous fish and native whitefish in subarctic lakes. *Freshwater Biology*, 58(6), 1234–1250. https://doi.org/10.1111/ fwb.12123
- Hayden, B., Myllykangas, J. P., Rolls, R. J., & Kahilainen, K. K. (2017). Climate and productivity shape fish and invertebrate community structure in subarctic lakes. *Freshwater Biology*, 62(6), 990–1003. https://doi.org/10.1111/fwb.12919
- Hein, C. L., Ohlund, G., & Englund, G. (2013). Fish introductions reveal the temperature dependence of species interactions. *Proceedings of* the Royal Society B: Biological Sciences, 281(1775), 20132641.
- Henriksen, A., Skjelvåle, B. L., Mannio, J., Wilander, A., Harriman, R., Curtis, C., Jensen, J. P., Fjeld, E., & Moiseenko, T. (1998). Northern European Lake Survey, 1995: Finland, Norway, Sweden, Denmark, Russian Kola, Russian Karelia, Scotland and Wales. *Ambio*, 27(2), 80–91.
- Hesthagen, T., & Sandlund, O. T. (2004). Fish distribution in a mountain area in south-eastern Norway: Human introductions overrule natural immigration. *Hydrobiologia*, 521, 49–59. https://doi. org/10.1023/B:HYDR.0000026350.93171.ba
- Hokanson, K. E. F. (1977). Temperature requirements of some percids and adaptations to the seasonal temperature cycle. *Journal of the Fisheries Research Board of Canada*, 34(10), 1524–1550. https://doi. org/10.1139/f77-217
- Hui, F. K. C. (2017). Boral: Bayesian ordination and regression analysis. R Package Version, 1.
- Hui, F. K. C., Taskinen, S., Pledger, S., Foster, S. D., & Warton, D. I. (2015). Model-based approaches to unconstrained ordination. *Methods in Ecology and Evolution*, 6(4), 399–411. https://doi. org/10.1111/2041-210X.12236
- Huitfeldt-Kaas, H. (1918). Ferskvandsfiskenes utbredelse og indvandring i Norge, med et tillæg om krebsen. Centraltrykkeriet.
- Jennings, M. D., & Harris, G. M. (2017). Climate change and ecosystem composition across large landscapes. *Landscape Ecology*, 32(1), 195– 207. https://doi.org/10.1007/s10980-016-0435-1
- Leathwick, J. R. (2002). Intra-generic competition among Nothofagus in New Zealand's primary indigenous forests. *Biodiversity & Conservation*, 11(12), 2177–2187.
- Livingstone, D. M., & Lotter, A. F. (1998). The relationship between air and water temperatures in the lakes of the Swiss Plateau: A case study with paleolimnological implications. *Journal of Paleolimnology*, 19(2), 181–198.
- Magnuson, J. J., Crowder, L. B., & Medvick, P. A. (1979). Temperature as an ecological resource. *American Zoologist*, 19(1), 331–343. https:// doi.org/10.1093/icb/19.1.331
- Mandeville, C. P., Rahel, F. J., Patterson, L. S., & Walters, A. W. (2019). Integrating fish assemblage data, modeled stream temperatures, and thermal tolerance metrics to develop thermal guilds for water temperature regulation: Wyoming case study. *Transactions* of the American Fisheries Society, 148(4), 739–754. https://doi. org/10.1002/tafs.10169
- Marchand, P. & Ellison, R. (2019). postGIStools: Tools for interacting with "PostgreSQL" / "PostGIS" databases. https://CRAN.R-project.org/ package=postGIStools
- Mehner, T., Emmrich, M., & Kasprzak, P. (2011). Discrete thermal windows cause opposite response of sympatric cold-water fish species to annual temperature variability. *Ecosphere*, 2(9), 1–16. https://doi. org/10.1890/ES11-00109.1

- Metz, M., Rocchini, D., & Neteler, M. (2014). Surface temperatures at the continental scale: Tracking changes with remote sensing at unprecedented detail. *Remote Sensing*, 6(5), 3822–3840; https://doi. org/10.3390/rs6053822
- Mills, C. A., & Hurley, M. A. (1990). Long-term studies on the Windermere populations of perch (*Perca fluviatilis*), pike (*Esox lucius*) and Arctic charr (*Salvelinus alpinus*). *Freshwater Biology*, 23(1), 119–136. https:// doi.org/10.1111/j.1365-2427.1990.tb00257.x
- Munsch, S. H., Greene, C. M., Johnson, R. C., Satterthwaite, W. H., Imaki, H., & Brandes, P. L. (2019). Warm, dry winters truncate timing and size distribution of seaward-migrating salmon across a large, regulated watershed. *Ecological Applications*, 29(4), e01880. https://doi. org/10.1002/eap.1880
- Nackley, L. L., West, A. G., Skowno, A. L., & Bond, W. J. (2017). The nebulous ecology of native invasions. *Trends in Ecology & Evolution*, 32(11), 814–824. https://doi.org/10.1016/j.tree.2017.08.003
- Ohman, J., Buffam, I., Englund, G., Blom, A., Lindgren, E., & Laudon, H. (2006). Associations between water chemistry and fish community composition: A comparison between isolated and connected lakes in northern Sweden. *Freshwater Biology*, *51*(3), 510–522. https://doi. org/10.1111/j.1365-2427.2006.01514.x
- Ovaskainen, O., Tikhonov, G., Norberg, A., Guillaume Blanchet, F., Duan, L., Dunson, D., Roslin, T., & Abrego, N. (2017). How to make more out of community data? A conceptual framework and its implementation as models and software. *Ecology Letters*, 20(5), 561–576. https://doi.org/10.1111/ele.12757
- Parkinson, E. A., Lea, E. V., Nelitz, M. A., Knudson, J. M., & Moore, R. D. (2016). Identifying temperature thresholds associated with fish community changes in British Columbia, Canada, to support identification of temperature sensitive streams. *River Research and Applications*, 32(3), 330–347. https://doi.org/10.1002/rra.2867
- Perrin, S. W. (2021). samaperrin/ChangingAssociations (Version v2) [Computer software]. https://doi.org/10.5281/zenodo.4665778
- Perrin, S. W., Bærum, K. M., Helland, I. P., & Finstad, A. G. (2021). Forecasting the future establishment of invasive alien freshwater fish species. *Journal of Applied Ecology*. https://doi. org/10.1111/1365-2664.13993
- Perrin, S. W., Englund, G., Blumentrath, S., O'Hara, R. B., Amundsen, P.-A., & Finstad, A. G. (2020). Integrating dispersal along freshwater ecosystems into species distribution models. *Diversity and Distributions*, 26(11), 1598–1611. https://doi.org/10.1111/ddi.13112
- Pollock, L. J., Tingley, R., Morris, W. K., Golding, N., O'Hara, R. B., Parris, K. M., Vesk, P. A., & McCarthy, M. A. (2014). Understanding cooccurrence by modelling species simultaneously with a Joint Species Distribution Model (JSDM). *Methods in Ecology and Evolution / British Ecological Society*, 5(5), 397–406.
- R Core Team. (2017). R: A language and environment for statistical computing. R Foundation for Statistical Computing. http://www.R-Project.org/
- Radinger, J., Alcaraz-Hernández, J. D., & García-Berthou, E. (2019). Environmental filtering governs the spatial distribution of alien fishes in a large, human-impacted Mediterranean river. *Diversity* & Distributions, 25(5), 701–714. https://doi.org/10.1111/ddi.12895
- Rahel, F. J., & Olden, J. D. (2008). Assessing the effects of climate change on aquatic invasive species source. *Conservation Biology*, 22(3), 521– 533. https://doi.org/10.1111/j.1523-1739.2008.00950.x
- Rockwell-Postel, M., Laginhas, B. B., & Bradley, B. A. (2020). Supporting proactive management in the context of climate change: Prioritizing range-shifting invasive plants based on impact. *Biological Invasions*, 22(7), 2371–2383. https://doi.org/10.1007/s10530-020-02261-1
- RStudio Team. (2020). RStudio: Integrated development environment for R. RStudio, Inc. http://www.rstudio.com/
- Sandlund, O. T., & Hesthagen, T. (2011). Fish diversity in Norwegian lakes: Conserving species poor systems. In M. Jankun, G. Furghala-Selezniow, M. Wozniak, & A. M. Wisniewska (Eds.), Water biodiversity assessment and protection (pp. 7–20). University of Warmia and Mazury.

Global Change Biology

- Seebens, H., Bacher, S., Blackburn, T. M., Capinha, C., Dawson, W., Dullinger, S., Genovesi, P., Hulme, P. E., van Kleunen, M., Kühn, I., Jeschke, J. M., Lenzner, B., Liebhold, A. M., Pattison, Z., Pergl, J., Pyšek, P., Winter, M., & Essl, F. (2020). Projecting the continental accumulation of alien species through to 2050. *Global Change Biology*, 27(5), 970–982. https://doi.org/10.1111/ gcb.15333
- Sharma, C. M., & Borgstrøm, R. (2008). Shift in density, habitat use, and diet of perch and roach: An effect of changed predation pressure after manipulation of pike. *Fisheries Research*, 91(1), 98–106. https://doi.org/10.1016/j.fishres.2007.11.011
- Sharma, S., Jackson, D. A., Minns, C. K., & Shuter, B. J. (2007). Will northern fish populations be in hot water because of climate change? *Global Change Biology*, 13, 2052–2064. https://doi. org/10.1111/j.1365-2486.2007.01426.x
- Silknetter, S., Creed, R. P., Brown, B. L., Frimpong, E. A., Skelton, J., & Peoples, B. K. (2020). Positive biotic interactions in freshwaters: A review and research directive. *Freshwater Biology*, 65(4), 811–832. https://doi.org/10.1111/fwb.13476
- Tammi, J., Appelberg, M., Beier, U., Hesthagen, T., Lappalainen, A., & Rask, M. (2003). Fish status survey of Nordic lakes: Effects of acidification, eutrophication and stocking activity on present fish species composition. *Ambio*, 32(2), 98-105. https://doi. org/10.1579/0044-7447-32.2.98
- Tikhonov, G., Abrego, N., Dunson, D., & Ovaskainen, O. (2017). Using joint species distribution models for evaluating how speciesto-species associations depend on the environmental context. *Methods in Ecology and Evolution*, 8(4), 443–452. https://doi. org/10.1111/2041-210X.12723
- Van der Veen, B., Hui, F. K. C., Hovstad, K. A., Solbu, E. B., & O'Hara, R. B. (2021). Model-based ordination for species with unequal niche widths. *Methods in Ecology and Evolution*, 12(7), 1288–1300. https:// doi.org/10.1111/2041-210X.13595
- Van Zuiden, T. M., Chen, M. M., Stefanoff, S., Lopez, L., & Sharma, S. (2015). Projected impacts of climate change on three freshwater fishes and potential novel competitive interactions. *Diversity and Distributions*, 22(5), 603–614. https://doi.org/10.1111/ddi.12422
- Venter, O., Sanderson, E. W., Magrach, A., Allan, J. R., Beher, J., Jones, K. R., Possingham, H. P., Laurance, W. F., Wood, P., Fekete, B. M., Levy, M. A., & Watson, J. E. M. (2016). Global terrestrial Human Footprint maps for 1993 and 2009. *Scientific Data*, *3*, 160067. https://doi. org/10.1038/sdata.2016.67
- Verdiell-Cubedo, D., Torralva, M., Andreu-Soler, A., & Oliva-Paterna, F. J. (2012). Effects of shoreline urban modification on habitat structure and fish community in littoral areas of a Mediterranean Coastal Lagoon (Mar Menor, Spain). Wetlands, 32(4), 631–641. https://doi. org/10.1007/s13157-012-0296-6
- Wagner, T., Hansen, G. J. A., Schliep, E. M., Bethke, B. J., Honsey, A. E., Jacobson, P. C., Kline, B. C., & White, S. L. (2020). Improved understanding and prediction of freshwater fish communities through

the use of joint species distribution models. *Canadian Journal* of Fisheries and Aquatic Sciences, 77(9), 1540–1551. https://doi.org/10.1139/cjfas-2019-0348

- Walther, G.-R., Beisner, S., & Burga, C. A. (2005). Trends in the upward shift of alpine plants. *Journal of Vegetation Science*, *16*(1998), 541–548.
- Walther, G.-R., Roques, A., Hulme, P. E., Sykes, M. T., Pysek, P., Kühn, I., Zobel, M., Bacher, S., Botta-Dukát, Z., Bugmann, H., Czúcz, B., Dauber, J., Hickler, T., Jarosík, V., Kenis, M., Klotz, S., Minchin, D., Moora, M., Nentwig, W., ... Settele, J. (2009). Alien species in a warmer world: Risks and opportunities. *Trends in Ecology & Evolution*, 24(12), 686–693. https://doi.org/10.1016/j.tree.2009.06.008
- Warton, D. I., Blanchet, F. G., O'Hara, R. B., Ovaskainen, O., Taskinen, S., Walker, S. C., & Hui, F. K. C. (2015). So many variables: Joint modeling in community ecology. *Trends in Ecology & Evolution*, 30(12), 766–779. https://doi.org/10.1016/j.tree.2015.09.007
- Wehrly, K. E., Wiley, M. J., & Seelbach, P. W. (2003). Classifying regional variation in thermal regime based on stream fish community patterns. *Transactions of the American Fisheries Society*, 132(1), 18–38.
- Wei, T., & Simko, V. (2017). R package "corrplot": Visualization of a correlation matrix. https://github.com/taiyun/corrplot
- Wetzel, R. G. (2001). Limnology: Lake and river ecosystems. Gulf Professional Publishing.
- Wickham, H. (2016). ggplot2: Elegant graphics for data analysis. Springer.
- Wickham, H., François, R., Henry, L., & Müller, K. (2019). dplyr: A grammar of data manipulation. https://CRAN.R-project.org/package=dplyr
- Winfield, I. J., Fletcher, J. M., & James, J. B. (2008). The Arctic charr (Salvelinus alpinus) populations of Windermere, UK: Population trends associated with eutrophication, climate change and increased abundance of roach (Rutilus rutilus). Environmental Biology of Fishes, 83(1), 25–35. https://doi.org/10.1007/s10641-007-9235-4
- Zurell, D., Zimmermann, N. E., Gross, H., Baltensweiler, A., Sattler, T., & Wüest, R. O. (2020). Testing species assemblage predictions from stacked and joint species distribution models. *Journal of Biogeography*, 47(1), 101–113. https://doi.org/10.1111/jbi.13608

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