```
1
```

1	Title
2	Environmental influences of life history strategies in partial anadromous brown trout (Salmo
3	trutta, Salmonidae)
4	
5	Running title
6	Life history of brown trout
7	
8	Authors
9	Marie Nevoux ^{1,2} , Bengt Finstad ³ , Jan Grimsrud Davidsen ⁴ , Ross Finlay ⁵ , Quentin Josset ^{6,2} ,
10	Russell Poole ⁵ , Johan Höjesjö ⁷ , Kim Aarestrup ⁸ , Lo Persson ⁹ , Oula Tolvanen ¹⁰ & Bror
11	Jonsson ¹¹
12	
13	Affiliations
14	¹ UMR ESE, Ecology and Ecosystem Health, Agrocampus Ouest, INRA, Rennes, France
15	² Management of Diadromous Fish in their Environment, AFB, INRA, Agrocampus Ouest
16	Université de Pau et des Pays de l'Adour UPPA, Rennes, France
17	³ Norwegian Institute for Nature Research, P.O. Box 5685 Torgarden, N-7485 Trondheim,
18	Norway
19	⁴ NTNU University Museum, Department of Natural History, N-7491 Trondheim, Norway
20	⁵ Marine Institute, Furnace, Newport, Co. Mayo, Ireland
21	⁶ DRED, Direction de la Recherche, de l'Expertise et des données, AFB, Observatoire Long
22	Terme de la Bresle, rue des Fontaines 76260 Eu, France
23	⁷ Department of Biological and Environmental Sciences, University of Gothenburg,
24	Gothenburg, Sweden
25	⁸ Technical University of Denmark, National Institute of Aquatic Resources

- 2
- 26 Vejlsoevej 39, DK-8600 Silkeborg, Denmark
- ⁹Department of Wildlife, Fish and Environmental Studies, Swedish University of Agricultural
- 28 Sciences, Umeå, Sweden
- ¹⁰ The Water Protection Association of the River Vantaa and Helsinki Region,
- 30 Ratamestarinkatu 7 b (3. krs), 00520 Helsinki, Finland
- 31 ¹¹Norwegian Institute for Nature Research, Department of Landscape Ecology, Gaustadalléen
- 32 21, 0349 Oslo, Norway
- 33

34 Abstract

35 This paper reviews the life history of brown trout (Salmo trutta, Salmonidae) and factors 36 influencing decisions to migrate. Decisions that maximize fitness appear dependent on size at 37 age. In partly anadromous populations, individuals that attain maturity at the part stage 38 typically become freshwater resident. For individual fish, the life history is not genetically fixed, and can be modified by the previous growth history and energetic state in early life. 39 40 This phenotypic plasticity may be influenced by epigenetic modifications of the genome. 41 Thus, factors influencing survival and growth, determine life history decisions. These are 42 intra- and interspecific competition, feeding and shelter opportunities in fresh and salt water, temperature in alternative habitats and flow-conditions in running water. Male trout exhibit 43 44 alternative mating strategies and can spawn as a subordinate sneaker or a dominant 45 competitor. Females do not exhibit alternative mating behaviour. The relationship between 46 growth, size and reproductive success differs between sexes in that females exhibit a higher 47 tendency to migrate than males. Southern populations are sensitive to global warming. In addition, fisheries, aquaculture with increased spreading of salmon lice, introduction of new 48 49 species, weirs and river regulation, poor water quality and coastal developments all threaten 50 trout populations. The paper summarizes life history data from six populations across Europe 51 and ends by presenting new research questions and directions for future research.

52

53 Keywords:

54 freshwater resident; habitat; migration; partial anadromy, Salmonidae, sex ratio

3

55 **Table of contents**

- 56 Abstract
- 57 **Table of contents**
- 58 Introduction
- 59 Resident versus anadromous brown trout
- 60 *Historical overview*
- 61 *Two phenotypes and a single species*
- 62 *Contrast in gene expression*
- 63 *Phenotypic plasticity*
- 64 *Epigenetics*

65 Trade-offs between growth and survival

- 66 *Habitat selection theory*
- 67 *Migration to improve growth*
- 68 *Migration can improve survival*
- 69 *Improved fecundity*
- 70 The energy surplus hypothesis
- 71 Sex-specific response
- 72 Bias in sex ratio
- 73 Strong size dependence in females
- 74 An alternative strategy in males
- 75 Environmental drivers of anadromy in juveniles
- 76 *Food availability*
- 77 Thermal condition
- 78 Flow condition
- 79 *Density-dependence*

- 5
- 80 Inter-specific competition
- 81 *Predation*
- 82 Influence of the marine environment on migration outcome
- 83 *Growth potential*
- 84 Seascape
- 85 Sea lice
- 86 Influence of the spawning habitat on reproduction outcome
- 87 *Habitat suitability*
- 88 Flow condition and scouring risk
- 89 Thermal condition and spawning phenology
- 90 Oxygen and siltation
- 91 Male mating strategies: frequency dependence, density dependence and shelter
- 92 Human impacts on sea trout
- 93 *Climate change*
- 94 Fisheries
- 95 Aquaculture
- 96 Degradation of water quality
- 97 *Threat to connectivity*
- 98 *Coastal development*
- 99 Interactions among impact factors
- 100 **Research Questions and future directions**
- 101 Assessment of partial anadromy
- 102 *Responses to climate change*
- 103 Epigenetics
- 104 Management strategies

- 105 Acknowledgements
- **References**
- **Table**
- 108 Figure legends

111 Introduction

7

112 Brown trout (Salmo trutta, Salmonidae) is a facultative (i.e. partly) anadromous species 113 (Lobón-Cerviá, Rasmussen, & Mortensen, 2017). It typically spawns in freshwater, but may reproduce successfully in estuaries at salinities below 4 ppt, such as on the coast of Gotland 114 115 in the Baltic Sea (Landergren & Vallin, 1998; Limburg, Landergren, Westin, Elfman, & Kristiansson, 2001). The species is partly migratory as some individuals within a population 116 117 may reside in or near the spawning area all year round, whereas other individuals move out of 118 this area for feeding. Migratory trout can be anadromous, feeding in the marine habitat. 119 Migrants generally return to breed with high precision to their area of origin for spawning, 120 but exceptions occur (B. Jonsson, Jonsson, & Jonsson, 2018). 121 122 Brown trout are phenotypically variable. Adult body length varies from approximately 10 to 123 100 cm (Evangelista, Boiche, Lecerf, & Cucherousset, 2014; B. Jonsson & Jonsson, 2011; 124 Sánchez-Hernández, Eloranta, Finstad, & Amundsen, 2017). The species exploit habitats 125 ranging from small brooks to rivers, lakes, estuaries and coastal sea, but are seldom found in 126 the open ocean, though recent literature indicate that some brown trout may live a more 127 pelagic life while at sea (B. Jonsson & Jonsson, 2011; Kristensen, Righton, del Villar-Guerra, 128 Baktoft, & Aarestrup, 2018). Populations adapt trophically to, and vary ecologically, 129 morphologically, behaviourally and genetically with local conditions over the distribution 130 area. This diversity complicates the systematics of the species, and makes some scientists 131 term Salmo trutta a species complex rather than a single species (Keller, Taverna, & 132 Seehausen, 2011; Patarnello, Bargelloni, Caldara, & Colombo, 1994; Sanz, 2017). Although 133 the systematics of brown trout have still to be resolved, the phylogeographic and the genetic 134 structure of the species were recently clarified (Sanz, 2017).

135

136 Freshwater resident populations are well described (Baglinière & Maisse, 2002; Frost & Brown, 1967; Gosset, Rives, & Labonne, 2006; B. Jonsson, 1989; Maisse & Baglinière, 137 138 1990). More complex is the ecology of anadromous trout (Harris, 2017; Harris & Milner, 139 2006), which occur naturally along the length of the Atlantic coast of Europe from northern 140 Russia to Portugal, Iceland included, and occur as an introduced fish in North and South 141 America, Australia, New Zealand and Kerguelen Islands (Baglinière, 1999; Elliott, 1994; B. Jonsson & Jonsson, 2011; Lecomte, Beall, Chat, Davaine, & Gaudin, 2013) (Figure 1). The 142 143 species is present along the coast of the Black and Caspian-Aral Seas in central Europe 144 (Baglinière, 1999; Elliott, 1994; B. Jonsson & Jonsson, 2011). Among all introduced 145 Salmonidae species, brown trout is the species with the highest success rate of naturalisation 146 and the largest distribution out of its original range. This is likely the result of a high adaptive 147 capacity and tolerance for habitat change (Baglinière, 1999).

148

149 Brown trout have been the focus of several books such as those by Fahy (1985), Elliott 150 (1994), Baglinière and Maisse (1999), Harris and Milner (2006), Harris (2017), Jonsson and 151 Jonsson (2011), and Lobón-Cerviá and Sanz (2017). However, some recent knowledge on migration, its complexity and plasticity is not included in these books (Figure 2). This work is 152 153 also motivated by the desire to complement the literature review with a compilation of data 154 from six populations across Europe. Partly anadromous means that individual populations can 155 consist of both freshwater resident and anadromous individuals. In this review, we present 156 information on both these life history components, and discuss drivers of anadromy, the influence of the marine environment on the migration, and effects of spawning habitat on 157 158 body size and sexual size dimorphism. Furthermore, we summarize knowledge on effects of 159 interspecific competitors and predators on abundance and behaviour of brown trout and 160 impacts of the parasitic sea lice on local sea trout, which constitutes one of the main threats to

wild populations in some areas (Thorstad et al., 2015), and which has led to significant
population collapses (e.g. Gargan, Poole, & Forde, 2006). Environmental constraints in
freshwater may stimulate migration of brown trout, including constraints from competition,
poor feeding and low growth opportunities. We review the effects of these, as well as
negative environmental impacts at sea that may select against the propensity to migrate
(Poole et al., 2006). Last, we present important questions for further research.

167

168 Resident versus anadromous brown trout

169 Historical overview

170 Carolus Linnaeus, in his 10th edition of Systema Naturae (1758), classified anadromous (sea)

171 trout (*Salmo eriox*) as a different species from river trout (*Salmo trutta*). The classification

172 was based on colouration and body form, a taxonomic classification maintained into the 19th

173 century (B. Jonsson & Jonsson, 2011). Among others, Dahl (1904) questioned the

174 classification as he observed that river trout could move downstream to sea. Furthermore,

175 Regan (1911) proposed that anadromous and non-anadromous trout could be freely

176 interbreeding fractions of a single species. Nevertheless, until the development of modern

177 genetic techniques in the 1980's, much uncertainty remained about whether or not sea trout

178 and river trout were the same or separate species (Frost & Brown, 1967).

179

180 Two phenotypes and a single species

181 Brown trout may have split from Atlantic salmon Salmo salar between 10 and 14 million

182 years ago (Crête-Lafrenière, Weir, & Bernatchez, 2012) and the five major evolutionary

183 lineages of brown trout evolved in its native Eurasian and North African range of distribution

184 with geographic isolation occurring during the Pleistocene Ice Ages and have largely

185 remained allopatric since then (Bernatchez, 2001). Their evolutionary histories have been

shaped by glaciations, habitat loss and varying potential for dispersal. They survived in icefree refuges during the periods of glaciation and colonized rivers as the ice cover retreated
(McKeown, Hynes, Duguid, Ferguson, & Prodöhl, 2010). After the last glaciation period
some 14000 years ago they entered rivers in the former glaciated northern areas and gradually
acquired their natural area we see today (Ferguson, 2006). The anadromous behaviour
probably existed before speciation of the salmonid family and the anadromous types, mainly
of the genus *Salmo*, evolved from the freshwater forms (Balon, 1980).

193

194 By origin, brown trout is chiefly a European species, but populations have been introduced to 195 areas outside their natural range they were unable to reach naturally (B. Jonsson & Jonsson, 196 2011). For instance, offspring of anadromous trout were released in some North American 197 rivers, and from these progenitors both anadromous and non-anadromous trout developed 198 (Rounsefell, 1958), proving that the two trout forms could develop from single gene pools. 199 Similar evidence was obtained from releases in the Kerguelen Islands (Davaine & Beall, 200 1997). The close connection between the two phenotypes was further highlighted by the 201 observation that offspring of a population of resident mountain living brown trout feed and 202 grow well at sea when transferred to a coastal river with free access to and from the sea (N. 203 Jonsson, Jonsson, & Hansen, 1994). They also survive and grow when they are released 204 directly into sea water as unsmoltified parr or immature brown trout (N. Jonsson, Jonsson, 205 Hansen, & Aass, 1994), although osmoregulatory performance is favoured by a progressive 206 transfer to seawater (Boeuf & Harache, 1982). Experimentally, Skrochowska (1969) and 207 Ombredane et al. (1996) demonstrated that anadromous as well as non-anadromous parents 208 produced both freshwater resident and sea-run migratory offspring. However, the proportion 209 of anadromous offspring was higher for anadromous than non-anadromous parents, 210 indicating a difference in gene expression between the two forms. Also, within single river

211 systems, anadromous and non-anadromous trout spawn together as has been observed in the 212 field (Charles, Guyomard, Hoyheim, Ombredane, & Baglinière, 2005; Cucherousset, 213 Ombredane, Charles, Marchand, & Baglinière, 2005; B. Jonsson, 1985; Vøllestad, 2017), but 214 the proportions of anadromous vs resident maternal origin parr will probably differ in different sections of the stream (Rohtla et al., 2017). Population diversification through 215 216 anadromous and non-anadromous individuals is not unique to brown trout, but is also found 217 in other salmonids (e.g. rainbow trout: Oncorhynchus mykiss, Arctic charr Salvelinus alpinus) 218 and some non-salmonid species (e.g. American shad: Alosa sapidissima) (B. Jonsson & 219 Jonsson, 1993).

220

11

221 Contrast in gene expression

222 Although anadromous and non-anadromous trout are genetically similar when in sympatry (Charles et al., 2005; Cross, Mills, & Williams, 1992; Hindar, Jonsson, Ryman, & Ståhl, 223 1991; Pettersson, Hansen, & Bohlin, 2001), around 50% of the variability in migration v. 224 225 residency, among individuals within a population, may be due to genetic variance (Ferguson, 226 Reed, Cross, McGinnity, & Prodöhl, 2019). Recent evidence suggests differences in the gene expression influencing the life history of the two trout phenotypes (Giger et al., 2006; 227 228 Lemopoulos et al., 2018, 2017). For instance, Giger et al. (2006) demonstrated that the gene 229 expression was primarily related to the migratory trait and not to genetic relatedness, whether 230 the fish migrate to the sea or a lake. They found that migrant and freshwater resident brown 231 trout from the same area exhibited different gene expression profiles, whereas evolutionarily 232 or geographically distant populations sharing the same life histories showed similar gene 233 expression, i.e. similar levels of mRNA transcripts. For example, a resident population 234 belonging to a Mediterranean lineage that diverged more than 500 000 years ago from the 235 Atlantic lineage, exhibited a gene expression profile like that of resident Atlantic populations. 236 By contrast, migratory and resident trout from the same river had very different profiles (sets 237 of mRNAs). Migration destination (mainstream river, lake, or sea) also appears to be 238 genetically programmed (Ferguson et al., 2019). Giger et al. (2006) suggested that the genetic 239 difference between life history types of brown trout is the result of a few controlling genes that determine the expression of many other genes defining the life history pattern. They 240 241 found that life history differences explained 45% of the total variability in gene expression levels, three times more than the variability explained by genetic diversity between 242 243 populations. Thus, gene expression appears to be influenced by the environment and 244 interactions between genes and environment that drive life history and migration decisions.

245

246 Phenotypic plasticity

247 There is considerable variation in life history strategies among individuals and populations, and in the timing and duration of marine migrations of brown trout (Aldvén & Davidsen, 248 2017; B. Jonsson, 1989; Thorstad et al., 2016). Figure 2 illustrates this diversity: after 1 to 7 249 250 years in freshwater, some individuals make a physiological transformation from parr to smolt 251 and migrate to sea. Young parr may also make excursions into brackish water. Downstream migration usually takes place in spring and autumn (Aarestrup, Birnie Gauvin, & Larsen, 252 253 2017; Poole et al., 2006; Winter, Tummers, Aarestrup, Baktoft, & Lucas, 2016). Anadromous 254 trout spend from 1 up to 36 months at sea on their maiden sea sojourn. In the marine habitat, 255 they feed on polychaetes, crustaceans and small fish (Knutsen, Knutsen, Gjøsæter, & 256 Jonsson, 2001), and grow more than in freshwater. Sometimes, anadromous trout perform an early (premature) return, characterised by a brief incursion in brackish or freshwater, before 257 258 heading back to the sea, this phenomenon may be exacerbated by sea louse infestation 259 (Birkeland, 1996). In summer, autumn and even winter, mature anadromous trout return to 260 their natal river to breed. While immature anadromous trout from northern Europe may spend

261	the winter in brackish or freshwater (often not in their natal river) before moving back to the
262	sea in the following spring (Thomsen, Koed, Nielsen, & Madsen, 2007). Mature trout spawn
263	in freshwater in the autumn/winter and return to sea immediately after spawning (B. Jonsson
264	& Jonsson, 2009b; N. Jonsson & Jonsson, 2002) or in the subsequent spring (B. Jonsson,
265	1985). The former is more dominating for trout spawning in small water courses with poor
266	shelter for wintering trout (B. Jonsson, Jonsson, Brodtkorb, & Ingebrigtsen, 2001; B. Jonsson
267	et al., 2018). Survivors typically undertake a shorter sea sojourn before heading back to
268	freshwater as repeat spawners (B. Jonsson & Jonsson, 2009b). Some of the parr do not smolt
269	and remain in freshwater during their entire life. Resident and anadromous trout can
270	reproduce on sympatric spawning grounds, or spawn in separate areas of the same river
271	(Hindar et al., 1991; Rohtla et al., 2017).
272	
273	Brown trout exhibit a large range of body sizes across their endemic distribution range, with
274	resident trout being on average smaller that anadromous trout (Figure 3). Within a given
275	catchment, resident trout rarely become as large as their anadromous counterparts of similar
276	age. However, older resident trout may become larger than young anadromous trout and the
277	largest resident trout can be larger than the smallest anadromous specimen within age-classes
278	(i.e. total age from birth). Large resident trout feeding on fish can grow equally large or larger
279	than anadromous trout do. Ferox trout from Scottish and Irish Lochs are examples of such
280	large resident trout (R. N. Campbell, 1979; Hughes et al., 2019).
001	
281	

The parameters controlling juvenile growth rate play an important role and various thresholds regulate the individual's decision of life history and migration strategy. Growth rate variation induces two antagonistic phenomena: sexual maturation or migration (Baglinière & Maisse, 1985; B. Jonsson & Jonsson, 1993; Maisse & Baglinière, 1999). The higher the growth rate, 286 the earlier the onset of maturation or migration. Presumably, the developmental threshold for precocious sexual maturation appears earlier in the season than that for migration. In partly 287 288 anadromous populations, mature parr typically become freshwater resident, although a few of 289 them may migrate to sea in a later year (B. Jonsson & Jonsson, 2011). The characteristics of 290 these thresholds, relative to the life stage of the fish, depend on environmental factors, the 291 stage and sex of the trout, and its genotype (Baglinière, Guyomard, Héland, Ombredane, & 292 Prévost, 2001). Across its European distribution area, the age of smolt varies widely, from 1-293 3 years in France, to 5-7 years in northern Norway (Figure 4). Within each population, the 294 size of smolts increases with age. This large-scale pattern suggests that despite phenotypic 295 plasticity and local environmental constraints, a physiological, threshold may underlie smolt 296 migration in brown trout. We may add, however, that smolt size is particularly small among 297 trout spawning in small streams, with some smolts as small as 8 cm being recorded (B. 298 Jonsson et al., 2001).

299

300 Like in the Atlantic salmon, the parr-smolt transformation takes place in the largest juvenile 301 trout of a given population (Tanguy, Ombredane, Baglinière, & Prunet, 1994) and smolting 302 occurs in spring. Silvering encompasses a peak of gill Na⁺/ K⁺-ATPase activity and a 303 simultaneous decrease in plasma prolactin is observed, together with morphological changes 304 (Aarestrup, Nielsen, & Madsen, 2000; Boeuf & Harache, 1982). But, compared to the 305 Atlantic salmon, no surge in growth hormone is documented (Tanguy et al., 1994). Hypo-306 osmoregulatory ability is greatest at the time of peak gill Na⁺/ K⁺-ATPase activity and it 307 increases in spring in all trout, irrespective of smolt status (C. Nielsen, Aarestrup, & Madsen, 308 2006). Smolting appears less stringent in brown trout than in Atlantic salmon, and does not 309 seem to be an obligatory process for seawater adaptation in this species. The level of smolting also depends on stock origin and body size, e.g. fast-growing juveniles can migrate to sea 310

311 without smolting (Tanguy et al., 1994). Usually, juvenile trout migrate from rivers in spring 312 (March-June in Europe) (Byrne, Poole, Dillane, Rogan, & Whelan, 2004; Flaten et al., 2016; 313 A. J. Jensen et al., 2012), but may also migrate at other times of the year, e.g. during autumn 314 (Aarestrup et al., 2017; B. Jonsson & Jonsson, 2009a; B. Jonsson et al., 2018; N. Jonsson & Jonsson, 2002; Poole et al., 2006; Poole, Whelan, Dillane, Cooke, & Matthews, 1996; Taal et 315 316 al., 2014; Winter et al., 2016) indicating that the time of seaward migration is highly plastic. 317 There are indications that smolt run timing may be changing with migrations occurring 318 earlier in recent years (Byrne et al., 2004), a phenomenon also observed in Atlantic salmon 319 (B. Jonsson, Jonsson, & Finstad, 2014; Otero et al., 2014), possibly caused by climate 320 change.

321

322 Post-smolt brown trout feed at sea and some immature fish return to estuarine or freshwater to overwinter, whereas others remain at sea (B. Jonsson et al., 2001, 2018). Evidence of 323 324 unsmoltified parr making brief brackish water excursions to switch streams has also been 325 documented (Taal et al., 2018). In northern Europe, immature trout can also return to 326 freshwater in summer after a short stay at sea. Ionoregulation in sea water at low temperature 327 is arduous, but anadromous trout have been observed at sea during winter and tolerate full 328 salinity seawater at temperatures as low as 1-2°C (Eldøy et al., 2017; J. L. A. Jensen & 329 Rikardsen, 2012; Knudsen et al., 2009; Olsen, Knutsen, Simonsen, Jonsson, & Knutsen, 330 2006). In the brackish Baltic Sea, parr can migrate from the freshwater to the Baltic coastal 331 zone without undergoing smolting. There, they may experience little or no physiological cost in terms of lower survival and growth from this transition (Landergren, 2005). Otoliths 332 333 collected from brown trout in the Baltic Sea sometimes show no evidence of a freshwater 334 history, raising the possibility of a contingent of the coastal population that does not depend 335 on riverine spawning or that the fish move to sea as fry (Limburg et al., 2001). The duration

and timing of marine migrations are likely governed by trade-offs between mortality risk and
growth opportunities in different habitats and the most beneficial strategy may vary among
individuals and populations. Based on life table analysis, Jonsson (1981) found that the
product of survival and fecundity, as a measure of fitness, was similar for freshwater resident
and anadromous brown trout of the same population. Possibly, the fitness of different aged
smolts are also similar as indicated from calculations of survival and fecundity of female
Atlantic salmon (B. Jonsson, Jonsson, & Albretsen, 2016).

343

344 Epigenetics

345 Divergent life history phenotypes may be the result of epigenetic modifications that link 346 environmental factors and the genome to regulate internal cues as reported from studies on 347 steelhead trout Oncorhynchus mykiss (Baerwald et al., 2016). The most studied epigenetic effect is a consequence of DNA methylation. High methylation levels are associated with 348 349 silencing of gene expression, and demethylation is linked to active gene transcription (Bird, 350 2002). Evidence of an epigenetic effect on life history variation was reported by Moran and 351 Pérez-Figueroa (2011). They found a link between DNA-methylation and maturation in Atlantic salmon male parr. Mature male parr exhibit reduced probability of smolting in brown 352 353 trout (B. Jonsson, 1985) and Atlantic salmon (Berglund, 1995). Epigenetic effects can be mediated early, such as at the embryonic stage (B. Jonsson & Jonsson, 2019). For instance, 354 355 thermal conditions during the embryogenesis may influence later growth, as found in Atlantic 356 salmon (A. G. Finstad & Jonsson, 2012) and zebra fish (Scott & Johnston, 2012). Furthermore, there is evidence of multiple differentially methylated genes between 357 358 anadromous and non-anadromous rainbow trout (Baerwald et al., 2016). They reported that 359 smolting of steelhead trout is associated with DNA methylation pattern. Furthermore, it has 360 been shown that salt-induced alterations in DNA methylation patterns play a role in sea water

361 adaptation in fishes (Artemov et al., 2017; Moran, Marco-Rius, Megías, Covelo-Soto, &

362 Pérez-Figueroa, 2013). One may hypothesize that early environmental factors may also

363 influence life history decisions and phenotypic plasticity in brown trout, although this has not

364 yet been investigated.

365

366 Trade-offs between growth and survival

367 Habitat selection theory

368 Mobile organisms are expected to select the most profitable feeding habitat. They should 369 choose the habitat where mortality (μ) over growth (g) (μ/g) is minimized (Werner & Gilliam, 1984). These two components are main determinants of their fitness. The relative 370 371 value of feeding habitats in terms of survival and growth often changes seasonally, or in 372 relation to the developmental stage of the individual. Thus, selection should favour migration 373 from freshwater to the sea when this reduces the value of μ/g , and habitat choices should be 374 influenced by benefits and costs in each habitat. However, organisms only experience the 375 situation where they currently are, and do not know the profitability of moving to distant 376 feeding grounds unless this is innately determined through an epigenetic threshold type 377 response or a genetically predetermined behaviour. Sea trout must therefore rely on 378 additional cues, such as present growth or size, to bias their movements towards the 379 appropriate feeding ground (Dodson, Aubin-Horth, Thériault, & Páez, 2013). Their response 380 is fine-tuned through natural selection, although the response appears phenotypically plastic, allowing the fish to cope with environmental stochasticity and variation. However, the degree 381 382 of plasticity is at least partly inherited and varies among populations (Fusco & Minelli, 2010). 383 There is little knowledge on the extent of epigenetic effects on behavioural decisions 384 (Baerwald et al., 2016), although Jonsson and Jonsson (2018) showed that the temperature

386 years later, when they as adults return from the ocean to spawn in freshwater.

experienced by Atlantic salmon embryos influences the timing of their homing migration

387

385

388 Migration to improve growth

Growth of trout depends largely on food consumption and temperature. Young trout 389 390 experiencing reduced growth because of food restrictions may either move to a more profitable feeding habitat or attain sexual maturation at an early age to make the most of a 391 392 poor environment (B. Jonsson, 1985). Thus, feeding migration is a viable alternative if distant 393 habitats provide improved growth opportunities without a disproportional decrease in 394 survival (B. Jonsson & Jonsson, 1993). Growth is typically higher at sea than in freshwater. 395 For instance, length increase during the second year in freshwater is typically 6 cm in 396 Southern Norway (L'Abee-Lund et al., 1989), which is approximately half the length increase obtained by immature trout spending their second year at sea (B. Jonsson & Jonsson, 397 398 2011; Poole et al., 1996), but growth decreases with age and sexual maturation. In Northern 399 Norway, the difference between freshwater and marine growth may be even larger (Berg & 400 Jonsson, 1990). In addition to better feeding opportunities, growth at sea may be less 401 constrained because of reduced population density and intraspecific competition in northern 402 temperate and sub-Arctic areas. In some systems, lakes may also offer better growing 403 condition than the nearby mainstream river and reduced costs of migration compared to 404 seaward migration (e.g. short migration distance, low predation). A similar pattern may hold 405 true for brown trout moving from tributaries to the mainstream, further downstream, in large river systems. Distinct populations of adfluvial trout have been identified, such as the 406 407 Dollaghan trout in Lough Neagh, Northern Ireland, and the Croneen trout in Lough Derg on 408 the Shannon, Ireland (Ferguson, 2004). Such a trade-off may also be driving lacustrine

409 migration in landlocked trout populations, as observed in lake Geneva (Champigneulle,
410 Buttiker, Durand, & Melhaoui, 1999).

411

412 The aggregation of six datasets from across Europe allowed us to highlight some new 413 patterns (Figure 5). Within our six populations, we observe a general pattern that emerges in 414 the form of a positive correlation in body length between anadromous and resident trout. This 415 means that rivers with large resident trout also produce large anadromous trout. Moreover, 416 the relative growth gained by the sea migration (i.e. the deviation from the 1:1 line in Figure 417 5) is larger in populations of large brown trout. Apparently, for anadromy to be a viable life 418 history tactic, the growth rate of anadromous fish must be higher than that of corresponding 419 freshwater residents.

420

421 Migration can improve survival

422 Mortality is higher during the marine migration than in freshwater, with sea trout facing high 423 predation rates during early sea migration, i.e. when they are small and cross the estuarine 424 zone. For instance, precise estimates from Black Brows Beck, Lake District, England showed that the instantaneous rate of survival of brown trout was about 0.88% day⁻¹ during the 425 426 second year in freshwater while the return rate of post-smolts at sea averages 0.25% day⁻¹ 427 (Elliott, 1993). Aldvén et al. (2015) detected a significantly higher mortality rate in brown 428 trout smolts moving from the river into a shallow estuary (mortality 26 to 51 %) than in 429 smolts entering a deep fjord (17.5-29.2 %). Dieperink et al. (2002; 2001) studied avian predation on emigrating wild and domesticated sea trout post-smolts in fjords of the western 430 431 Baltic Sea and North Sea. In total, 65% of the post-smolts were eaten by fish-eating birds, 432 and during the first two days after entering the sea, both wild and domesticated post-smolts 433 suffered a daily predation rate estimated at 20-34%. Thus, the trout appeared to experience a

434 transient period with elevated risk of predation immediately after exposure to sea water. However, in other places, the early mortality is noticeably lower. Survival of smolts 435 436 migrating through Randers and Mariager Fjords in Denmark showed survivals between 76-80 437 % 30 days after fjord entry and in Poole Harbour in England 88% of the trout smolts entering the estuary made the 12 km transition to the open sea (Aarestrup, Baktoft, Koed, del Villar-438 439 Guerra, & Thorstad, 2014; del Villar-Guerra, Aarestrup, Skov, & Koed, 2013; Lauridsen et al., 2017). Predation rates appear to be influenced by the time and size at migration, and sea 440 441 trout of the River Imsa, Norway exhibit highest survival if migrating in May (ca. 15% until 442 river return) and low survival if migrating to the sea between July and December (ca. 2%) (B. 443 Jonsson & Jonsson, 2009a). However, in Gudsø Stream, Denmark, the return rates of spring 444 and autumn migrants were similar (Birnie Gauvin & Aarestrup, 2018). This suggests 445 difference in autumn mortality between rivers. Also, one would expect that sea migration in 446 the autumn would benefit other fitness components, such as growth, and compensate for this 447 potential high initial migration cost.

448

449 By migrating, sea trout can avoid adverse environmental conditions in the home stream, such 450 as winter icing-up of streams or summer drought. For instance, in small streams regularly 451 experiencing summer drought, sea trout can migrate to sea at a small size early in life, and in 452 this way avoid poor growth and survival conditions (B. Jonsson et al., 2001; Landergren, 453 2004; Titus & Mosegaard, 1992). This pattern resembles the strategy followed by some 454 Pacific salmonids, such as pink salmon Oncorhynchus gorbuscha and chum salmon 455 Oncorhynchus keta, which both start their seaward migration early in life. Trout from streams 456 with low water level during winter may migrate to a neighbouring watercourse for overwintering (Aldvén & Davidsen, 2017), or stay in marine waters (Eldøy et al., 2017; B. 457 458 Jonsson et al., 2018; Olsen et al., 2006).

459

460 Improved fecundity

Fecundity, or number of ova per unit length, changes between stocks (Fahy, 1985; Poole, 461 462 Byrne, Dillane, Whelan, & Gargan, 2002) and between stocks in different regions (N. Jonsson & Jonsson, 1999; Solomon, 1997). But, with larger body size, sea trout increase their 463 464 gamete production and direct competitive ability on the spawning ground, and thus obtain augmented reproductive success (Fleming, 1996; Gross, 1987; Hutchings & Myers, 1985). 465 466 For instance, mean fecundity of sea trout from Vangsvatnet Lake, Norway was 1790 eggs 467 compared to 330 eggs for non-anadromous females of similar age (i.e. < 20%; B. Jonsson, 1981, 1985). Egg size increases with the size of the mother, and with increasing egg size, 468 469 early growth and viability of offspring increases (Bagenal, 1969; Segers & Taborsky, 2011). 470 However, these relationships are dependent on the female life history, with females investing in larger ova as the fish become larger and older. It seems that anadromous females had 471 472 smaller ova compared with freshwater-resident females of similar body size (i.e. older fish) 473 and achieved higher fecundity as they grew bigger (Acolas, Roussel, & Baglinière, 2008). 474 Another advantage for the offspring of anadromous females may come from an earlier 475 hatching date in the season, as reported in anadromous brown trout (N. Jonsson & Jonsson, 476 1999) and rainbow trout relative to sympatric resident trout (Zimmerman & Reeves, 2000). A 477 longer spawning period in resident trout may buffer this phenological difference in some 478 cases, e.g. river Oir, France (Maisse et al., 1991). Less is known about the production of male 479 milt and spermatozoa in brown trout, although differences in sperm count were observed 480 between stocks and the length of the spawning migration (B. Jonsson & Jonsson, 2006b) and 481 a negative trend was linked with size and age of males, possibly indicating a lower fertilising 482 ability of older fish within any one stock (Poole & Dillane, 1998).

483

485 The energy surplus hypothesis

486 Several studies suggest that anadromy in brown trout is triggered by energy limitation in natal 487 rivers (reviewed by Dobson et al. (2013)), and can be terminated if the relative advantage of 488 migration changes (Sandlund & Jonsson, 2016). Some early studies documented an 489 asymptotic size at one year, which is limited by the carrying capacity of the river (Baglinière 490 & Maisse, 1990). Thus, to meet their energy requirements and grow further, individuals 491 should change habitat and move toward a more productive habitat further downstream in the 492 watershed or at sea (Baglinière & Maisse, 2002). Brown trout appear to favour a migration 493 strategy when the energy surplus useable for growth becomes low (Forseth, Nesje, Jonsson, 494 & Hårsaker, 1999; B. Jonsson & Jonsson, 1993).

495

496 Experimentally, Davidsen et al. (2014) induced migration by decreasing ration size during 497 the six months prior to smolting. Similarly, Jones et al. (2015) demonstrated that reduced 498 winter and spring feeding increased the tendency to smoltify for lake feeding brown trout 499 from the River Klarälven, Sweden. In contrast, another experiment reported that trout facing 500 food restriction in late autumn exhibited lower rate of silvering, which is indicative of a delay 501 in smolting (Näslund, Sundström, & Johnsson, 2015). Thus, winter/beginning of spring 502 appears to be a critical period when the decision is made about whether to smolt and migrate 503 to a better feeding area. These experimental findings contrast with Thorpe & Metcalfe's 504 (1998) hypothesis that autumn is the critical time in Atlantic salmon, when decisions of 505 migration versus maturation and residency are taken. This implies that brown trout are more 506 plastic and energy intake during winter and spring may over-rule an initial decision, 507 depending on the energetic state of the fish (cf. B. Jonsson, Jonsson, & Finstad, 2013). In 508 contrast, rearing brown trout in aquaculture with optimum rations reduces the probability of

22

509 smolting, leads to early maturation and lowers the return rates from sea releases (Byrne,

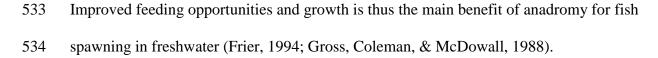
510 Poole, Dillane, & Whelan, 2002; Mills, Piggins, & Cross, 1990).

511

512 Within brown trout populations, fast growers tend to migrate at a younger age, and typically 513 smaller size than slow growers (B. Jonsson, 1985; Økland, Jonsson, Jensen, & Hansen, 1993) 514 and within a given cohort, larger juveniles tend to undertake longer migrations (Ombredane 515 et al., 1996). Individuals with a high metabolic rate may migrate downstream earlier as their 516 energy demands more rapidly exceed those available in their current habitat than in smaller 517 trout (Ferguson, Reed, McGinnity, & Prodöhl, 2017; Forseth et al., 1999; Peiman et al., 518 2017). Sea trout smolts have very low energy density at the time of migration (ca. 350 KJ/100 519 g wet mass), which is similar to that of resident trout after spawning (N. Jonsson & Jonsson, 520 1997a, 1998). This is due to a low lipid density (ca. 1.5 g/100 g wet mass), which may 521 contribute to the compensatory growth exhibited by sea trout during the first weeks at sea 522 (Marco-Rius, Caballero, Moran, & Leaniz, 2012). In brook trout Salvelinus fontinalis, 523 Morinville and Rasmussen (2003) reported that in the year before migration, migrant brook 524 trout have consumption rates 1.4 times higher than those of resident brook trout. However, 525 migrants have lower growth efficiencies (ratio of growth to consumption) than residents, 526 indicating that migrants have higher metabolic costs, and relatively less surplus energy 527 available at the time the migrations commences.

528

Individual growth rate better integrates the mechanisms underlying migration decision
whereas body size is more strongly related to survival in trout (Acolas, Labonne, Baglinière,
& Roussel, 2012). Intrinsic differences between individuals explain why migrants and
residents differ in body size in many partial migratory species (Chapman et al., 2012).



535

536 Size may also interact with the ability of juveniles to establish and hold territories, thus smaller and/ or younger parr may be forced to leave the stream by larger and/ or older 537 538 conspecifics (Landergren, 2004). In Atlantic salmon, some populations exhibit a bimodality in size by the end of the first growing season (Baglinière & Maisse, 1985). Individuals from 539 540 the upper mode, i.e. large juveniles, smolt and migrate to sea the following spring (Baglinière 541 & Maisse, 1985; Thorpe, Talbot, & Villarreal, 1982). Atlantic salmon appear to smolt when 542 the young have reached a critical size, although this pattern may not be detected in slower 543 growing populations of northern Europe (Økland et al., 1993). This is probably because 544 survival at sea is strongly size dependent (Flaten et al., 2016; B. Jonsson et al., 2016). In trout, it is unknown to what degree size influences age at smolting (Figure 4). Juvenile size 545 546 bimodality has not been detected at the end of the growth season (Baglinière, Prévost, & 547 Maisse, 1994). This suggests that the migration decision may be taken later in trout, highlighting the greater flexibility of trout life history compared to Atlantic salmon 548 549 (Baglinière et al., 2001).

550

551 Sex-specific response

552 Skewed sex ratios are commonly observed within the anadromous and resident components 553 of partially migratory brown trout populations. The skewed sex ratios results from differences 554 in physiological constraints and life history trajectories in males and females. The fitness of 555 females appears more closely associated with body size than in males. Although the same 556 environmental conditions interact with threshold limits to determine individual life histories,

560 Bias in sex ratio

561 Although the sex ratio of parr is even in brown trout, there appears to be a female surplus

among emigrating smolts (Cucherousset et al., 2005; B. Jonsson, 1985; Klemetsen et al.,

563 2003). In northern France, the percentage of females among migrating smolts varies between

564 57% and 67% (Euzenat, Fournel, & Richard, 1999; Quéméré, le Gentil, & Launey, 2011),

565 which fits well with what is found elsewhere (Table 1). Also, females typically outnumber

566 males among adults entering streams for spawning (J. S. Campbell, 1977; Euzenat, Fournel,

567 & Fagard, 2006; K. W. Jensen, 1968; B. Jonsson, 1985; Mills et al., 1990; Okumuş,

568 Kurtoglu, & Atasaral, 2006; Poole et al., 2006). On the spawning grounds, however, the

569 operational sex ratio may be skewed in favour of males because they stay longer and if

570 possible, spawn with several females (Aarestrup & Jepsen, 1998).

571

572 The size of the stream used for spawning may influence spawning success and therefore indirectly the sex ratio, with male surplus reported in several small streams (B. Jonsson et al., 573 574 2018; Rubin, Glimsäter, & Jarvi, 2005). The highest proportion of females among the 575 anadromous trout are found in the nutrient rich and productive streams and large rivers of the 576 Baltic and the North Sea areas (Table 1), and this bias may be larger when the high 577 proportion of repeat spawners, as post-spawning survival is greater in females than in males... 578 In small streams, males mature younger and smaller than females, and relatively more males 579 will therefore survive to adulthood (B. Jonsson, 1985; B. Jonsson & Jonsson, 2015). Where 580 there is a female surplus among the smolts, it may be evened out among anadromous adults

because males mature younger than females and more will therefore survive to adulthood (B.Jonsson et al., 2018).

583

584	Anadromous females typically outnumber anadromous males in larger streams, and the same
585	is reported from other species of partly migratory trout and charr, such as rainbow trout
586	(McMillan, Katz, & Pess, 2007; Rundio, Williams, Pearse, & Lindley, 2012; Van Doornik,
587	Berejikian, & Campbell, 2013), Arctic charr (Nordeng, 1983), whitespotted charr Salvelinus
588	leucomaenis (Tamate & Maekawa, 2004) and some Atlantic salmon populations (N. Jonsson,
589	Jonsson, & Hansen, 1998), although exceptions occurs (Bagliniere, personal communication).
590	Dodson et al. (2013) noted an increase in male anadromy of brook trout towards the north.
591	This has not yet been investigated for brown trout.

592

593 Strong size dependence in females

594 Large female size is selected by natural selection as reproductive success is more strongly 595 associated with body size in females than males (B. Jonsson, 1985; Kendall et al., 2014). 596 Furthermore, body size variation is smaller in females than in males. This may be because 597 females do not exhibit alternative spawning tactics (sneaking versus fighting) as males do 598 (McLean, Bentzen, & Quinn, 2004). Furthermore, the gonad development of females requires 599 more energy (Baglinière et al., 2001; Fleming, 1996; Prouzet, Le Bail, & Heydorff, 1984). 600 Thus, females are more likely to migrate to more food rich areas, and stay there for longer 601 periods of time to grow larger before spawning (Bordeleau et al., 2018; Cucherousset et al., 602 2005; Ferguson, 2006; Northcote, 1992). Undertaking a longer and more risky sea migration 603 may reduce survival, resulting in relatively few anadromous female trout returning as adults. 604 Higher marine mortality in females than males has been reported for anadromous whitespotted charr by Tamate & Maekawa (2004), but no empirical evidence is available for 605

sea trout. On the other hand, post-spawning survival is higher in female charr (and salmon)
than in males, which may also hold for brown trout. Thus, sex-ratio of repeat spawners may
be even more skewed in favour of females (Euzenat et al., 1999).

609

610 An alternative strategy in males

611 A large male size may be favoured because of both natural and sexual selection (Young, 2005). Natural selection because large males have larger testes (B. Jonsson & Jonsson, 2005; 612 613 N. Jonsson & Jonsson, 1997b), win spawning contests more often and therefore spawn more 614 times than smaller males (Fleming et al. 1996). They are favoured by sexual selection 615 because females prefer large males as partners (Petersson et al. 1999; Serbezov et al. 2010) 616 and large males gain better access to females due to higher fighting success. On the other 617 hand, male size is not directly associated with reproductive success (Poole & Dillane, 1998) 618 even though large males do enjoy strong competitive advantages over smaller resident males 619 when fighting for mates (Bohlin, Dellefors, & Faremo, 1990). Some males may adopt an 620 alternative spawning behaviour (Dominey, 1984; Gross, 1991). While large males gain access 621 to females through fighting, small resident males effectively employ a sneaking tactic to the 622 same ends (Olsén, Järvi, Mayer, Petersson, & Kroon, 1998), avoiding the risks of migration, 623 and breeding at a younger age (Foote, Brown, & Wood, 1997). However, 'precocious' 624 maturation is less pronounced in brown trout than in Atlantic salmon, where males can 625 mature before the age of 1 (Baglinière & Maisse, 1985). Since maturation and smolting may 626 be considered competing developments in terms of energy allocation and physiology, the 627 energy requirements for smolting after maturing may be too large to allow migration within a 628 reasonable seasonal schedule. Indeed, maturation in brown trout correlates with delayed or 629 reduced rates of smolting (Bohlin et al., 1990; Cucherousset et al., 2005; Dellefors & Faremo, 630 1988; B. Jonsson, 1985), but nevertheless does happen.

632	Intermediate-sized individuals, in contrast to large or small individuals, may be ineffective at
633	both fighting and sneaking mating strategies, putting them at a competitive disadvantage and
634	maintaining the separation of two distinct male phenotypes as observed within many wild
635	salmonid populations (Gross, 1985). It is possible that where intermediate-sized mature males
636	are present on spawning grounds, they utilise female mimicry in order to approach females
637	without attracting the aggression of large dominant males (Esteve, 2005). This behaviour,
638	which is associated with altered male colouration resembling that of mature females, has been
639	reported in Arctic charr, pink salmon, chum salmon and red-spotted masu salmon
640	(Oncorhynchus masou ishikawae) (Kano, Shimizu, & Kondou, 2006; Keenleyside & Dupuis,
641	1988; Sigurjónsdóttir & Gunnarsson, 1989).

642

643 Environmental drivers of anadromy in juveniles

Anadromy and residency in brown trout are considered alternative tactics within a single strategy (Dodson et al., 2013). The decision to migrate to sea results from a trade-off between mortality and growth in fresh and salt waters so that the overall fitness is maximized, and individuals must use proximate cues to adopt the most appropriate life history (Kendall et al., 2014). Environmental factors influencing growth in early life are probably the main cues on which the decision is made (Baglinière et al., 2001; Ferguson et al., 2017; B. Jonsson & Jonsson, 1993).

651

652 Food availability

Both empirical and experimental studies have reported negative correlations between the

654 proportion of migratory trout and food availability in the natal freshwater system (review in

655 Ferguson *et al.* (2017)). For instance, the migration of brown trout between a tributary where

656 they spawned and the main river, ceased when the growth opportunities in the main river decreased because of damming and reduced flow in the main river (B. Jonsson & Sandlund, 657 1979; Sandlund & Jonsson, 2016). Kendall et al. (2014) noted that food quality, i.e. energy 658 659 value, may be as important as food quantity. Hence, resident trout appear to be abundant in the most productive areas, with a high invertebrate biomass. Where lakes are present, a 660 661 lacustrine-adfluvial migration pattern often predominates (Ferguson, 2004; Ferguson et al., 2017; Gresswell, Liss, & Larson, 1994), as some lakes may offer good growth condition with 662 663 a lower predation cost than sea migration. This is supported by the observation that stocking 664 well fed, juvenile hatchery trout often produces freshwater resident fish, but starving the fish before release induces a migratory behaviour (Davidsen et al., 2014; Larsson, Serrano, & 665 666 Eriksson, 2011). Thus, growth opportunities in the local habitat play an important role in the 667 decision whether to migrate (Larsson et al., 2011). Also, environmental stochasticity in the river, such as winter frost or summer drought, can seasonally constrain feeding opportunities 668 669 and initiate migration. In this case, trout may opt for migration to the marine environment, 670 which may appear more stable and protective against harsh physical conditions (Aldvén & 671 Davidsen, 2017; Ferguson et al., 2017).

672

673 Thermal condition

Temperature is a key factor structuring freshwater ecosystems. It may influence migratory
decisions, with both absolute temperature and variation in temperature being important
(Kendall et al., 2014; Morita, Tamate, Kuroki, & Nagasawa, 2014). Optimum temperature for
growth of juvenile 1g brown trout is found to be 13-14°C (Forseth et al., 2009), but it can be
high if energy consumption is very high such as in piscivorous trout (Forseth & Jonsson,
1994), and lower if the fish are particularly large or feeding is reduced (Morita, Fukuwaka,
Tanimata, & Yamamura, 2010). Temperature is clearly linked to food availability, feeding

681 activity, metabolism and lipid storage (Ferguson et al., 2017). High temperature allows higher food consumption but also increases maintenance costs and may accelerate energy shortages. 682 683 Furthermore, standard metabolic rate may be affected by the temperature experienced by the 684 yolk feeding larvae (Álvarez, Cano, & Nicieza, 2006). Thus, unless food production increases by the same order of magnitude, individuals may not be able to meet their energetic 685 686 requirements when temperature increases. However, this pattern may be attenuated by temperature itself, as abnormal temperature may also act as a physiological stressor and 687 688 induce migration (Peiman et al., 2017). However, an increase in temperature may fuel the 689 production of invertebrates and thereby offer improved feeding opportunities for resident 690 trout, as explained above (Morita et al., 2014; Olsson, Greenberg, Bergman, & Wysujack, 691 2006). But further projected increases in river water temperature above the optimal 692 temperature for growth will likely decrease growth, body length and age at smolting, and 693 ultimately reduce the survival of trout (Davidson, Hazlewood, & Cove, 2007; Fealy et al., 694 2010). However, the relative contribution of temperature to trout growth compared to other 695 environmental drivers remains difficult to quantify. In Atlantic salmon, Bal et al. (2011) 696 reported that variation in freshwater temperature had only a small effect on the growth of 697 juveniles compared to variation in salmonid density.

698

699 Flow condition

In the rivers, trout are largely ambush predators, relying on the flow to bring food to them although they are more active hunters than Atlantic salmon. In darkness, they can pick zoobenthos on the bottom. The nursery habitat may differ between resident and anadromous conspecifics as reported by Morinville and Rasmussen (2003). They studied brook trout, and found that the young of migratory trout used faster running water than young resident trout, and that they also fed on different food items. This pattern is expected to come from intrinsic 706 differences in metabolic requirement and is not indicative of an effect of flow condition on 707 migration decision. High flow often correlates with high availability in food resource, which 708 influences the opportunities for growth (Kendall et al., 2014; Morinville & Rasmussen, 709 2003). Furthermore, flow may have an effect on anadromy in rainbow trout, at least in climate zones where summer flows are a limiting factor (Kendall et al., 2014). Flow that 710 711 regularly reaches zero may represent a population bottleneck for non-migratory fishes 712 (Courter, Justice, & Cramer, 2009), and low flow may explain the small size at first smolting 713 of brown trout in some small streams (Borgstrøm and Heggenes 1988; Jonsson et al. 2001). 714 More generally, unsuitable flow condition in streams is likely to drive the propensity to 715 migrate to sea, e.g. due to low flow, or highly variable flow conditions in the home stream, or 716 a reduction in suitable habitat and high keen competition from conspecifics (density 717 dependence effect).

718

719 So far, there is empirical evidences of a positive correlation between flow and the number of 720 migrating smolts in brown trout. In the Danish River Lilleaa the migration speed of wild sea 721 trout smolts was positively correlated with water discharge (Aarestrup, Nielsen, & Koed, 722 2002). But when the flow remains low, trout may wait until the temperature comes over a 723 certain level (Aarestrup et al., 2002). In a Swedish west coast stream, smolts remained in the 724 rivers during years with low precipitation until the temperature reached 10°C, and migration 725 increased thereafter irrespective of the discharge (Aldvén et al., 2015). A shift from nocturnal 726 to diurnal migration appears to occur when the water temperature is 12-13°C and is most pronounced in large smolts (Haraldstad, Kroglund, Kristensen, Jonsson, & Haugen, 2017). 727 728 Single large temperature increases can also initiate a daytime migration (Aarestrup et al., 729 2002). Water temperature and flow influence the phenology of the sea trout smolt migrations. 730 Temperature regulates the timing of the start and the end of the migration period whilst water

- level, change in water flow and water temperature influence the number of smolts migrating
 each day (Byrne et al., 2004; N. Jonsson & Jonsson, 2002).
- 733

734 Density-dependence

735 The food availability interacts with fish density in limiting the resources available per capita.

Higher competition for food and space at high density results in fewer residents, with

migrants maximising growth by moving into the sea (Ferguson et al., 2017; Olsson et al.,

2006). During periods of drought or hydropower regulation, density will increase. Stradmeyer

et al. (2008) showed how the importance of dominance status for maintaining food intake

740 increases as the polarization between the top ranked fish and others increased. As a result,

fewer fish will mature and become residents, thus migration may be an alternative (B.

742 Jonsson & Jonsson, 1993).

743

744 Inter-specific competition

745 Similarly, competing species may also influence the proportion of brown trout that migrate, 746 mediated through a density-dependence process (Olsson et al., 2006). Young brown trout and 747 Atlantic salmon compete for food and space in rivers, as evidenced by the spatial segregation 748 observed between the two species (B. Jonsson & Jonsson, 2011). Young Atlantic salmon are 749 mainly found in shallow, fast flowing habitats of the main river while young trout colonize 750 tributaries and headwaters (Baglinière & Arribe-Moutounet, 1985; Baglinière & 751 Champigneulle, 1982; Kalleberg, 1958). In the Shelligan Burn (Scotland), the density of young Atlantic salmon was negatively correlated with that of young trout (Egglishaw & 752 753 Shackley, 1982). Brown trout are territorial and very aggressive, and through interference, 754 they constrain young Atlantic salmon from shallow, slow-flowing areas (Harwood, Metcalfe, Armstrong, & Griffiths, 2001; Heggenes, Baglinière, & Cunjak, 1999; Houde, Wilson, & 755

756 Neff, 2017; Kalleberg, 1958). This trend is accentuated by the early hatching of trout 757 (Baglinière et al., 1994), which are larger and have already established a territory by the time 758 Atlantic salmon fry emerge. Furthermore, when brown trout are removed, young Atlantic 759 salmon expand their habitat into former trout-defended areas, i.e. competitive release (Hearn, 1987; Kennedy & Strange, 1986). Experimentally, Gibson and Erkinaro (2009) showed that 760 761 brown trout were four times more aggressive than corresponding Atlantic salmon. As a result of their more aggressive behaviour and larger size, brown trout have a competitive advantage 762 763 in slow flowing, shallow areas along stream banks and pools where they monopolize the food 764 resources (Gibson & Erkinaro, 2009; Höjesjö, Stradmeyer, Griffiths, & Armstrong, 2010). 765 Atlantic salmon, on the other hand, make more use of cover, deep pools and fast-flowing 766 riffles where food is difficult to defend. In addition, their more streamlined body shape and 767 larger pectoral fins are assumed to give young Atlantic salmon a selective advantage in rapidly flowing rivers (Karlström, 1977), and they may outcompete trout in fast flowing 768 streams (Montorio, Evanno, & Nevoux, 2018). The interaction between young brown trout 769 770 and Atlantic salmon in rivers is adversely competitive, i.e. increased abundance of the one 771 leads to a decrease in abundance of the other. This is exemplified in formerly acidified rivers 772 treated with calcium carbonate (limestone powder). As the abundance of Atlantic salmon 773 increased with time, the density of brown trout decreased (Hesthagen, Larsen, Bolstad, Fiske, 774 & Jonsson, 2017). However, even when the abundance of Atlantic salmon increases at the 775 expense of brown trout, the total output of salmonid smolts increases (cf. Kennedy and 776 Strange, 1986).

777

The aggressiveness of brown trout is energetically costly. Their high-energy use is assumed
to be the main reason why brown trout are outcompeted by Arctic charr because of a much
lower growth efficiency in cold, food limited habitats (A. G. Finstad et al., 2011). In shallow,

temperate localities richer in food, on the other hand, brown trout outcompete Arctic charr
because of its higher aggressiveness. In sympatry, the two segregate spatially, with charrs
shifting to the pelagic or the profundal zone (Heggberget, 1984; Klemetsen et al., 2003;

784 Langeland, L'Abée-Lund, Jonsson, & Jonsson, 1991).

785

786 There is also competition between grayling Thymallus thymallus and brown trout. Mäki-Petäys et al. (2000) suggested that competition between these species increases in streams 787 788 during winter when ice decreases the habitat availability. Competition between the two 789 species may be a reason for their partial segregation in rivers (Greenberg, Svendsen, & 790 Harby, 1996). However, recruitment of grayling and trout were positively associated in an 791 English chalk stream (Bašić, Britton, Cove, Ibbotson, & Gregory, 2018). Also, Alpine 792 bullhead (Cottus poecilopus) is a strong competitor to juvenile brown trout where these 793 species co-occur (Hesthagen & Heggenes, 2003; Holmen, Olsen, & Vøllestad, 2003). For 794 instance, Holmen et al. (2003) reported that brown trout density was five to 10 times higher 795 at the upper, allopatric site than in downstream areas where it lived in sympatry with the 796 Alpine bullhead. Diet composition suggested that brown trout in sympatry with Alpine 797 bullhead foraged more on invertebrate drift and surface arthropods than allopatric brown trout 798 did.

799

In addition, novel species that invade brown trout areas, naturally or because of humanmediated releases, may influence brown trout migration and habitat use. Brown trout living in sympatry with brook trout consumed more terrestrial prey, had smaller home ranges, and a stouter body shape (Závorka et al., 2017). Sympatric brown trout also had lower specific growth rates, suggesting a lower fitness. Other introduced species, such as rainbow trout affect brown trout negatively. Blanchet *et al.* (2007) found by use of laboratory and field studies that rainbow trout significantly affected native brown trout habitat selection and
apparent survival. In contrast, Baran *et al.* (1995) demonstrated a competition advantage of
the brown trout in mountainous rivers when the two species live in sympatry. The more
similar the ecology of the interacting species, the greater the impact of competition is.

810

811 There are examples of rivers where Atlantic salmon and resident trout co-exist, but little or no 812 sea trout run is observed, as in Ireland and Brittany (France). In the Irish systems, the 813 presence of lakes and running river sections may promote habitat segregation and reduce 814 inter-specific competition while in large salmon rivers, the occurrence of sea trout is often 815 restricted to the lower stretches and to small tributaries close to or directly entering an estuary 816 (Fahy, 1985; K. F. Whelan, 1989). Similarly, the tree-like shape of the watersheds in 817 Brittany, characterized by many small tributaries, may maximise segregation and reduce 818 competition between the two species (Baglinière et al., 1994).

819

820 It is still unclear whether interspecific competition in freshwater influences the tendency to 821 migrate to the sea. However, one may hypothesize that brown trout will exhibit a stronger 822 tendency to migrate if their resource use in freshwater is constrained by competing species, as 823 shown for increased competition from conspecifics (Olsson et al., 2006). Montorio et al. 824 (2018) provided empirical evidence for inter-specific competition-exclusion relationship. 825 They found a negative effect of trout density on growth and survival in juveniles, but a 826 positive effect of Atlantic salmon density on the probability of migration in young trout. They reported that in years following a high recruitment in salmon, the propensity of trout leaving 827 828 the stream was high. Atlantic salmon may be a main competitor in temperate areas and Arctic 829 charr in sub-Arctic and Arctic areas.

830

The role of predators in shaping patterns of partial migration has been historically neglected (Chapman et al., 2012). However, if the decision to migrate is a balance between growth opportunities and mortality risk, increased predation in freshwater should select for anadromy in trout. Predation risk may constrain habitat use and thereby reduce individual growth (Johnsson, Rydeborg, & Sundström, 2004). Alternatively, high predation in freshwater may offer more resource per capita to the remaining individuals with an opposite effect on the opportunity for growth.

839

Predation is expected to be particularly high during the smolt run and in the first few days after the smolts reach sea water (Dieperink et al., 2002; Thorstad et al., 2016). If there are alternative prey, such as Atlantic salmon smolts migrating to sea at the same time, this may reduce the mortality and increase the fitness of sea trout (B. Jonsson & Jonsson, 2009b), or it may stimulate the aggregation of predators (Jepsen, Holthe, & Økland, 2006). Thus, although effects of predation on the life history strategy of trout is not well studied, this does not mean that predation is unimportant in this respect.

847

848 Influence of the marine environment on migration outcome

The decision to migrate is a trade-off between benefit and cost. Costs associated with sea migrations include energy used for swimming and increased probability of death, e.g. owing to predation, parasitism and diseases, but also the physiological adjustments for salt excretion (Gross et al., 1988a). Distance to the coast and deep sea areas may limit the distribution of sea trout. It is generally assumed that most anadromous brown trout feed in shallow waters within 100 km from the river mouth (B. Jonsson & Jonsson, 2011; Thorstad et al., 2016). Nevertheless, there are some evidences of large within and between population variations in

856 marine migration distance (B. Jonsson & Jonsson, 2014; Kristensen et al., 2018; Potter,

857 Campbell, Sumner, & Marshall, 2017; Prodöhl et al., 2017).

858

859 Growth potential

Growth levels off with age and spawning history, but the difference in size between resident 860 861 and anadromous trout increases with age (Figure 6, left panel). This suggests that the best growth strategy in brown trout would be to leave early and stay longer at sea. In the 862 863 Burrishoole River (Ireland) and the River Bresle (France), fish with the combination of smolt 864 age and sea age that results in the highest growth gain are well represented among the 865 returning anadromous trout (Figure 6, right panel). However, some of these strategies are 866 represented by a low proportion of trout. Data support a mismatch between the age-specific 867 additional growth and the age-structure of the anadromous trout in River Vosso (Norway), 868 River Tamar (England) and River Oir (France). In these populations, data show a shorter 869 marine sojourn and/ or an older age at smolting than expected under the hypothesis of growth 870 maximisation. We argue that strong constraints in the marine environment, in terms of 871 mortality, may select against growth maximisation strategies. For instance, young smolts may 872 be too small to escape length-biased predation, thus favouring late runners. Furthermore, 873 marine survival may be too low to support long marine sojourns, i.e. very few two sea winter 874 sea trout may survive until spawning.

875

By driving the level of mortality and growth in trout that initiate seaward migration, the
marine environment influences the balance between the cost and benefit of the time at sea. If
the costs of feeding at sea exceeds the gains, selection should favour freshwater residency
(Gross et al., 1988a). In only a few generations, altered environmental conditions can
terminate the tendency to migrate (Olsson et al., 2006; Sandlund and Jonsson, 2016). The

sea is likely to contribute to the reduction in sea trout abundance over space and time.

883

884 Seascape

Quéméré et al. (2016) explored effect of seascape, e.g. the marine landscape, on the 885 886 connectivity of brown trout populations between rivers, and how the seascape influences the life history of brown trout across northern France. They found two genetically distinct 887 888 populations which strongly differ in terms of migratory propensity and stock characteristics. 889 Western populations mainly produce freshwater resident trout and small sea trout (finnock, 890 i.e. post-smolt returning in the same year as they went to sea for the first time), while eastern 891 populations produce large sea trout that perform long sea-sojourns. They hypothesised that 892 such a pattern may be driven by the spatial arrangement and feeding opportunities of marine 893 habitats, which promotes a clinal variation in migratory behaviour. In the highly productive 894 Eastern basin of the English Channel, anadromous trout can achieve very large size and long 895 sea sojourn, which may favor dispersal and gene flow among neighbouring streams In the 896 Western basin, marine feeding opportunities are poor and trout have evolved mainly a 897 freshwater resident life history (Quéméré et al., 2016). The dispersal barrier may act as a 898 forcing factor for a distinct life history (Richardson, Urban, Bolnick, & Skelly, 2014). On the 899 other hand, the rivers appear different, and habitat characteristics may also add to the 900 variation observed. Fahy (1985) also observed differences in trout size, growth and condition 901 between trout returning from different regions, such as the Irish Sea compared to the Irish west coast and even from different bays along the west coast (Poole, unpublished). 902 903

905 The sea lice *Lepeophtheirus salmonis* and *Caligus elongatus* are natural parasites of sea trout 906 that can propose major threats to trout in the marine environment. During the last 30 years, 907 salmon farming has increased the abundance of sea lice (B. Finstad & Bjørn, 2011; Fjørtoft et 908 al., 2017; Thorstad & Finstad, 2018; Thorstad et al., 2015; Tully, Poole, & Whelan, 1993; 909 Tully, Poole, Whelan, & Merigoux, 1993) in some areas. Sea lice induce ionoregulatory 910 dysfunction, physiological stress, anaemia, reduced feeding and growth, changes in post-911 smolt behaviour, increased susceptibility to secondary infections, reduced disease resistance 912 and ultimately mortality of individual sea trout (B. Finstad & Bjørn, 2011; ICES, 1997; 913 Poole, Nolan, & Tully, 2000). Wild adult sea trout in farm-free areas exhibit relatively low 914 sea lice levels (Gargan, Tully, & Poole, 2003; Schram, Knutsen, Heuch, & Mo, 1998). On the 915 other hand, in farm-intensive areas, sea lice levels on wild sea trout are typically higher 916 (Gargan et al., 2003; I. Moore et al., 2018; Tully, Poole, Whelan, et al., 1993; Tully & 917 Whelan, 1993). Sea lice infestations in these areas regularly exceed more than 0.35 lice per 918 gram of fish mass, initiating physiological disturbance to the trout with weight range from 919 16-70 gram (Wells et al., 2006, 2007). At higher lice levels mortality may occur (Taranger et 920 al., 2015). The sea lice levels on wild sea trout are particularly high within 30 km of salmon 921 farms, but elevated lice infestations extend beyond this distance (Gargan et al., 2003; I. 922 Moore et al., 2018; Serra-Llinares, Bjørn, Finstad, Nilsen, & Asplin, 2016; Serra-Llinares et al., 2014; Tully, Gargan, Poole, & Whelan, 1999). Therefore, sea lice in intensively farmed 923 924 areas negatively impact wild sea trout through a suite of primary and secondary impacts 925 reducing marine growth and survival of sea run fish (Halttunen et al., 2018; S Shephard, 926 MacIntyre, & Gargan, 2016). Quantification of these impacts remains a challenge, although 927 population-level effects have been quantified in Atlantic salmon using field experiments (Krkošek et al., 2013; Skilbrei et al., 2013; Vollset et al., 2016), showing higher survival in 928

929 groups of anti-parasitically treated fish relative to non-treated control groups. Reduced 930 growth and increased mortality in the marine environment decrease the benefits of marine 931 migration for sea trout (Halttunen et al., 2018), and high sea lice levels might select against 932 anadromy. In some populations, immature anadromous trout return frequently to rivers during the growth season. Such 'premature returning' may be an early warning signal of 933 934 heavy salmon lice infections (Halttunen et al., 2018; Maisse et al., 1991). Sea lice-induced effects on sea trout may in extreme cases lead to local loss of anadromous trout and a 935 936 transition to entirely resident trout populations (discussed in Poole *et al.* (2006) with 937 reference to the Burrishoole sea trout stock). However, recent studies showed the complexity 938 of the relationship between brown trout and sea lice infections in the wild (relationship 939 between size of trout and distance of farm cage) and emphasised the requirement of further 940 research to quantify these effects (I. Moore et al., 2018).

941

942 Influence of the spawning habitat on reproduction outcome

943 Marine migration is a strategy that allows brown trout to escape from, or reduce the influence 944 of, density dependence and resource limitation in freshwater, and by that maximize 945 reproductive output (B. Jonsson & Jonsson, 2006b; Lobón-Cerviá, 2007; Marco-Rius, 946 Caballero, Moran, & Leaniz, 2013). But to reproduce, trout must return to freshwater and 947 compete for access to suitable spawning sites and mates. Intense spawning competition may 948 introduce both density and frequency dependant pressures (Berejikian et al., 2010; Foote et 949 al., 1997; Gross, 1985), where body size is important for determining reproductive success 950 (Serbezov, Jorde, Bernatchez, Olsen, & Vøllestad, 2012). Interference competition generally 951 favours large individuals (Olsén et al., 1998), however, in small shallow tributaries, small resident trout may have higher fitness because these habitats are less accessible for large 952 953 individuals. In populations dominated by individuals with short sea sojourns, there is larger

954 spatial overlap between the spawning grounds of resident and anadromous trout than in populations dominated by longer sea sojourn. Interbreeding between the two phenotypes 955 956 (Charles, Roussel, Lebel, Baglinière, & Ombredane, 2006) produces fertile offspring 957 (Ombredane et al., 1996), but these offspring have reduced migration probability because the migratory strategy is partly inherited (Baglinière et al., 2001; B. Jonsson, 1982). Size-specific 958 959 spawning success of both sexes is influenced by habitat characteristics as well as the competitive ability and density of the competitors, although contrasting selective mechanisms 960 961 operate on males and females (B. Jonsson & Jonsson, 2015). Hence, the effect of each of 962 these selective variables is sex-specific and determined by both the environment and the 963 population structure (Young, 2005). Thus, spawning requirements play an important role in 964 determining the frequency, abundance, and sex ratio of anadromous brown trout (B. Jonsson 965 & Jonsson, 2015).

966

967 Habitat suitability

968 In order to breed successfully, female brown trout require a specific combination of

969 hydrological conditions and substrate composition (Baglinière, Champigneulle, & Nihouarn,

970 1979; Gauthey et al., 2015; Montgomery, Buffington, Peterson, Schuett-Hames, & Quinn,

971 1996; Tappel & Bjornn, 1983). In many circumstances, the availability of habitat that

adequately meets these conditions represents the primary factor limiting the size of salmonid

populations (Buffington, Montgomery, & Greenberg, 2004; Kondolf & Wolman, 1993).

974

975 Despite their difference in body size, the specific spawning habitat requirements of

976 anadromous trout versus freshwater resident trout have hardly been studied (Nika, Virbickas,

877 & Kontautas, 2011; Walker & Bayliss, 2007). It seems clear, however, that growth associated

978 with anadromy will represent differing reproductive advantages depending on the nature of

980

979

981 The extra energy expenditure of migrating a longer distance is indicated by the gradual 982 decrease in the condition factor of anadromous brown trout with increasing migratory 983 distance inland. The gonadosomatic index of males (I=mass of gonads/somatic mass) 984 decreases with migratory distance (B. Jonsson & Jonsson, 2006a). Bohlin et al. (2001) 985 hypothesized that the fitness of migrants is negatively related to the altitude of the spawning 986 area because of migratory costs, and in support of this they found that anadromous trout were 987 replaced by non-anadromous conspecifics at an altitude of ca. 150 m in southern Sweden. 988 More than distance itself, it is the energetic demand required to reach the spawning ground 989 that matters. As such, the cumulative effect of barriers (natural or artificial) can greatly 990 impede sea trout migration inland in fragmented rivers.

991

992 Flow condition and scouring risk

993 Prior to spawning, female salmonids flex their flanks and tail fin to excavate depressions 994 (nests) in the streambed into which eggs are deposited. Due to the allometric relationship 995 between fish length and caudal thrust, salmonid females can construct their nests in gravel 996 where the median particle diameter does not exceed 10% of their fork length (Kondolf & 997 Wolman, 1993). Thus, nest sizes and depths are positively correlated with body size (Crisp & 998 Carling, 1989; Elliott, 1984; Gauthey et al., 2015). The depths of gravel mobilized during 999 high flow events are influenced by the strength of the stream flow, stream gradient and 1000 substrate composition (Harrison, Legleiter, Wydzga, & Dunne, 2011). Within water courses 1001 prone to such riverbed 'scour', individual scour events are likely to kill trout eggs located 1002 above the maximum scour depth, thereby exerting a strong selective pressure for a locally-

available spawning habitat.

determined minimum female body size (Montgomery et al., 1996). Thus, due to their ability
to dig deeper nests and move heavier gravel, large anadromous females may enjoy a distinct
advantage over small resident females as scour depth and gravel particle size increase. This
size-dependent mechanism may interfere with habitat selection in trout within a river basin,
leading to positive correlation between female body size and river size at the spawning area.

1009 Furthermore, female brown trout show a marked preference for spawning in previously 1010 utilized nests containing incubating eggs (Gortázar, Alonso, & Jalón, 2012). This behaviour, 1011 known as redd superimposition, includes a period of excavation that leads to high mortality 1012 rates amongst eggs deposited by earlier spawners (McNeil, 1964; Nomoto et al., 2010). This 1013 behaviour has also been reported where brown trout and Atlantic salmon share the same 1014 spawning ground (Baglinière et al., 1979). The intensity of redd superimposition depends on 1015 the density of spawners, the size of the spawning area, the duration of the spawning period 1016 and the distribution of spawners within the river system. As with environmentally-based 1017 scour, the survival of eggs depends on whether they are deposited below the depth of gravel 1018 mobility during subsequent excavations. As a result, the ability of large (i.e. anadromous) 1019 females to construct deeper nests than smaller freshwater resident females may represent a 1020 distinct inter-specific and intra-specific advantage where population density is high or where 1021 suitable spawning habitat is in short supply.

1022

1023 Thermal condition and spawning phenology

In some populations, freshwater residents appear to spawn later in the spawning period than
their anadromous counterparts (N. Jonsson & Jonsson, 1999). The asynchronous spawning of
anadromous and resident females may lead to the eggs of anadromous females hatching
earlier than those of resident females, potentially conferring a competitive advantage on the

1028 progeny of anadromous females through early growth opportunities. But this different

1029 temporal spawning pattern between resident and anadromous trout does not exist everywhere.

1030 In France the spawning period of sea trout (mid-December to late January) occurs within the

1031 larger spawning period of resident trout (late November to mid-February) (Baglinière, Pers.

1032 com.).

1033

1034 The efficiency of yolk conversion to body tissue declines as water temperature increases 1035 (Fleming & Gross, 1990), meaning that rising water temperatures during spring are likely to 1036 have a more pronounced negative impact on the growth performance of eggs spawned later in 1037 the breeding period. It is possible that the higher energy density found in the eggs of resident 1038 females relative to those of anadromous females from the same population (N. Jonsson & 1039 Jonsson, 1997b, 1999) represents an adaptation which mitigates the competitive 1040 disadvantages that result from late spawning. Possibly, thermal differences between eggs 1041 fertilized early or late during the spawning period may also influence later growth and 1042 reproductive allocations, as they appear to do in Atlantic salmon (A. G. Finstad & Jonsson, 1043 2012; B. Jonsson & Jonsson, 2014, 2018).

1044

1045 Oxygen and siltation

In addition to benefitting from increased fecundity, large anadromous females, which tend to accumulate high somatic mass during marine feeding, are likely to produce larger eggs than smaller resident females (Hendry & Day, 2003; N. Jonsson & Jonsson, 1999). While increased redd depth is beneficial in protecting eggs from washout during scour events, the threat of egg asphyxiation due to sedimentation or weakly oxygenated water increases with burial depth (Haury, Ombredane, & Baglinière, 1999). Einum *et al.* (2002) found that the fitness of brown trout eggs at differing oxygen levels is size-dependent, with larger eggs 1053 exhibiting significantly higher survival than small eggs when the oxygen concentration is 1054 low, possibly as a result of their increased surface area which facilitates diffusion. Laboratory 1055 experiments with brown trout embryos suggest that sublethal stress in the gravel nest caused 1056 by hypoxia can alter the behaviour and survival of trout during the early juvenile period 1057 (Roussel, 2007). It follows that the nature of local scour regimes, dissolved oxygen levels, 1058 siltation rates and riverbed gravel composition may all influence the relative fitness and 1059 frequency of differing adult female phenotypes. These factors should select for increased 1060 body size and egg mass associated with anadromy where scour is deep, gravel particles are 1061 large, and siltation or deoxygenation represent a significant threat to egg survival.

1062

1063 Male mating strategies: frequency dependence, density dependence and shelter

1064 There is strong regional and local variation in age at maturity and freshwater residency rates 1065 of males, indicating that populations are locally adapted with regard to body size and 1066 migratory strategy (Bohlin et al., 1990; Dellefors & Faremo, 1988). Where the habitat 1067 permits, small males may safely remain close to spawning females by utilising crevices, 1068 rocks, debris, or shallow areas as refuges where the aggression of large males is avoided 1069 (Gross, 1985). Thus, the availability of appropriate refuge habitats near spawning sites may 1070 be a factor influencing the fitness and, consequently, the proportion of male brown trout that 1071 forego marine migration or mature at a small size (B. Jonsson & Jonsson, 2011).

1072

While the abiotic characteristics of spawning habitat and local hydrological regimes are likely to influence the relative fitness of anadromous and freshwater resident phenotypes in brown trout, direct competition on spawning grounds may exert density-dependent selection on these phenotypes. But also, polymorphism amongst mature males appears to be maintained by frequency-dependent selection during spawning that increases the relative fitness of the 1078 rarer phenotype and promotes the co-occurrence of both mating strategies (Berejikian et al., 1079 2010; Foote et al., 1997; Gross, 1985; Hutchings & Myers, 1985). For instance, in Atlantic 1080 salmon, aggressive competition for mates between large males occupies much of their focus, 1081 allowing young and small males to go unnoticed and successfully employ their sneaking 1082 tactic when courting females. As the density of large males increases, this alternative mating 1083 behaviour may confer a greater mating success, balanced against the risk of getting killed by 1084 a large male. Conversely, the fitness of large males is likely to be reduced by competition for 1085 mates as the frequency of large males increases (Engqvist & Taborsky, 2016). Similarly, as 1086 the frequency of precocious males increases, their individual genetic contribution is likely to 1087 diminish due to increased competition from other precocious males (Hutchings & Myers, 1088 1994). In brown trout, males do not mature as early and small as in salmon, but the contrast 1089 between small resident and large migrant individuals provides a similar framework. 1090 1091 It has been argued that the development and maintenance of an evolutionarily stable 1092 alternative male reproductive strategy requires a strong underlying genetic basis, although the 1093 expression of these strategies is conditional, in the sense that environmental conditions associated with growth and survival interact with genetically based threshold responses to 1094 1095 determine individual strategy (Engqvist & Taborsky, 2016; Tomkins & Hazel, 2007). 1096 However, one cannot rule out epigenetic mechanisms, which might provide a rapid response 1097 to changing environmental conditions (Moran & Pérez-Figueroa, 2011). Hutchings and 1098 Myers (1994) suggested that within a given trout population, an evolutionarily stable 1099 continuum of strategy frequencies is maintained by polygenic thresholds (which are likely to 1100 differ significantly between sexes as well as between individuals) of environmentally

1101 controlled traits (primarily associated with growth). In this sense, the polyphenic mating

1102 strategies employed by male brown trout may be regarded as both locally adapted and

phenotypically plastic, resulting from the interplay between genotype, environmental
conditions and both frequency and density dependent pressures. While density dependent
pressures are likely to lead to high rates of migration and anadromy where feeding
opportunities are poor in the home river, frequency dependent pressures largely maintain
alternative mating strategies and associated polyphenism through intrasexual competition on
the spawning beds (Forseth et al., 1999; Landergren, 2001; Wysujack, Greenberg, Bergman,
& Olsson, 2009).

1110

1111 Human impacts on sea trout

1112 Climate change

1113 Possible effects of climate change on sea trout populations have been extensively reviewed (Elliott & Elliott, 2010; Graham & Harrod, 2009; B. Jonsson & Jonsson, 2009a; Lassalle & 1114 1115 Rochard, 2009). The expected climate change in the Atlantic region is for warmer, drier 1116 spring and summer seasons, and for milder and wetter winters, with more precipitation falling 1117 as rain and less as snow, shorter ice-covered periods, and more frequent periods of extreme 1118 weather, including droughts, heavy rain events and strong spates. A shorter ice-covered 1119 period may increase the activity and energy use of trout in rivers and decrease production 1120 unless feeding opportunities increase to a similar extent (Watz et al., 2015). 1121

1122 Climate change may also be responsible for increased rates of land erosion, which is

1123 reinforced by land-use change in agriculture and forestry practices. As a result, an increase in

1124 river sediment is likely. Suspended sediment affects trout behaviour and survival directly.

- 1125 Sediment loads clog gravel beds impeding brown trout from spawning and reducing
- 1126 recruitment success (Scheurer, Alewell, Bänninger, & Burkhardt-Holm, 2009). Invertebrate
- 1127 communities and growth opportunities for trout are also affected by increasing sediment input

(J. I. Jones et al., 2012). An increase in water temperature, notably during the spawning
period, is likely to impact reproductive behaviour and egg mortality as observed in
experimental conditions in Atlantic salmon (Beall & De Gaudemar, 1999), ultimately leading
to a drastic reduction in the species distribution. Overall, Jonsson and Jonsson (2009a)
predicted a north and northeastward movement of the distributional range of sea trout, with
decreased production and population extinction in the southern regions and invasion of new
spawning and nursery rivers and feeding areas in the north.

1135

1136 Fisheries

1137 Trout are exploited through fishing both in freshwater and marine habitats. Overexploitation, 1138 i.e., when a population is exploited to the extent that the optimal recruitment cannot be 1139 maintained due to a lack of spawners and thereby insufficient egg deposition, leads to 1140 population declines, and/ or a shift the balance between anadromous and freshwater resident 1141 compartments of populations. Because brown trout are iteroparous, the impact of exploitation 1142 is cumulative on older and larger fish (Solomon & Czerwinski, 2007). Catch statistics, as 1143 well as information on catch per unit effort and catch rates, are important for the monitoring 1144 of sea trout and assessments of levels of exploitation. In the Burrishoole catchment, fishing 1145 effort was found to be the most important determinant of catch with a weaker but significant 1146 relationship between catch and stock (Mills, Mahon, & Piggins, 1986). The quality of catch 1147 statistics for trout is adequate in some fisheries and countries, but needs improvement in 1148 others (Milner et al., 2007). The extent of illegal sea trout fishing is unknown, but can be 1149 significant in some countries.

1150

1151 Aquaculture

1152 Salmon farming affects sea trout by spreading infectious diseases caused by viruses, bacteria, 1153 fungi and parasites, as well as often functioning as a barrier to migration (Aarestrup & Koed, 1154 2003). Such diseases may affect both the freshwater and marine stages of the sea trout life 1155 cycle. Although numerous studies document the impact of salmon lice (see previous section 1156 on sea lice), little knowledge and monitoring is available on other effects of fish farming on 1157 wild brown trout populations (Bakke & Harris, 1998). However, considering the importance 1158 of the parasitism impact on wild salmonids in Norway, an operational salmon lice model has 1159 been developed to provide an improved monitoring system for risk assessment (Myksvoll et 1160 al., 2018) as well as the implemented "traffic light system" for a sustainable fish farming in 1161 Norway (Vollset et al., 2018). In Ireland, Single Bay Management (Anon, 2008) was 1162 implemented in the early 1990s which included close monitoring and advice for to harvest or 1163 treat lice, especially in the immediate months before and during the wild smolt runs.

1164

1165 Degradation of water quality

Water acidification, pollution (e.g. from agriculture, sewage treatment works, roads and
mining), hydropower developments, flow regulation, water abstraction, migratory barriers,
and habitat alterations as diverse as high diffuse organic load or clearing riffles of stones for
passing logs through, affect trout populations negatively (reviewed by Thorstad et al., 2014;
K. F. Whelan & Poole, 1993).

1171

Freshwater acidification arising from industrial emissions of sulphur dioxide and nitrogen
oxides to the atmosphere, and subsequent acid precipitation wiped out trout populations in
southern Fennoscandia during the last century. Although emissions were reduced after 1990,
many rivers in this area are still affected by chronic or episodic acidification (e.g. Hesthagen

1176 et al., 2017; Kroglund et al., 2007; Rosseland & Kroglund, 2010). Brown trout are vulnerable 1177 to acidification, although to a lesser degree than Atlantic salmon (Henrikson & Brodin, 1178 1995). Hence it is likely that any liming strategy adopted specifically for Atlantic salmon 1179 may also restore water to a quality suitable for sea trout. However, liming strategies can still 1180 be optimised to improve conditions for sea trout in tributaries and small streams that 1181 presently are not included in the liming programme of the main stem of rivers. Furthermore, 1182 competition with more acid sensitive species than brown trout, such as Atlantic salmon, 1183 seems to increase as a consequence of liming and reduce the abundance of trout in acidified 1184 waters treated with calcium carbonate (Hesthagen et al., 2017). 1185

1186 Contaminants derived from intensive agriculture, afforestation, mining and other industries 1187 impact sea trout negatively, both as a consequence of direct and indirect effects in freshwater (B. Jonsson, Jonsson, & Ugedal, 2011). High nitrite concentrations are lethal for embryos, 1188 1189 this impact being higher when dissolved oxygen concentrations are low (Massa, Baglinière, 1190 Prunet, & Grimaldi, 2000). Livestock grazing contributes to river eutrophication due to 1191 runoff from livestock manure. Interestingly, intensive agriculture systems relying on a large 1192 quantity of nutrients led to an increase in productivity in river Scorff, France (Roussel et al., 1193 2013). This coincided with an increase in growth performance in juvenile Atlantic salmon 1194 and a decrease in mean age at smolting (Rivot et al., 2009). However, this positive effect on 1195 fish may only hold within a reasonable level of eutrophication. In the Burrishoole, Ireland, 1196 land use 1197 change was found to have no significant impact on the freshwater survival of either salmon or 1198 trout (de Eyto et al., 2016). Nevertheless, contaminants also alter the parr-smolt 1199 transformation, and influence the run-timing and survival of smolts once they enter sea water 1200 (S. D. McCormick, Hansen, Quinn, & Saunders, 1998; Rosseland & Kroglund, 2010).

1201 Contaminants may damage fish gills and thereby compromise the ionoregulatory capacity of 1202 smolts (Kroglund et al., 2007; S. D. McCormick et al., 1998; A. Moore, Lower, Mayer, & 1203 Greenwood, 2007). Sublethal exposure to contaminants during smolting may therefore have 1204 implications for the survival for trout at sea and their tendency to migrate. Livestock 1205 trampling of stream bank may induce direct destruction of eggs and fry (Gregory & Gamett, 1206 2009), potentially leading to population decline, as illustrated in a cutthroat trout model 1207 (Peterson, Rieman, Young, & Brammer, 2010). Stream bank trampling also increases 1208 siltation sedimentation of salmonids redds, which limits the interchange of surface and 1209 intragravel water and, therefore, the supply of dissolved oxygen to the embryo (Heywood & 1210 Walling, 2007). Stock exclusion from watercourses using riparian fencing and limited pool 1211 excavation has the potential to preserve and enhance wild trout populations in headwaters 1212 (Summers, Giles, & Stubbing, 2008). This rehabilitation work has become a widely 1213 recommended fisheries management practice (Summers, Giles, & Willis, 1996).

1214

1215 Road and railway activities contribute to freshwater pollution, but there is little available 1216 information on effects for trout. Meland et al. (2010) suggested that traffic-related 1217 contaminants, especially those emanating from contaminated tunnel wash water, reduce 1218 growth of juvenile trout. In winter, salt from the roads may also be an issue. In addition, poor water quality in rivers may be responsible for various skin diseases. For instance, UDN-like 1219 1220 (Ulcerative Dermal Necrosis) disappeared from Europe in the 1970s, but in 1993, Roberts 1221 wrote: "it seems likely that UDN is a cyclical disease (...). It can therefore be expected to 1222 return again before the middle of the 21th century" (Roberts, 1993). Currently, a UDN-like disease is becoming an important driver of adult mortality in Baltic salmonids, this disease 1223 1224 has been detected in Latvia, Poland, Germany and Sweden (ICES, 2018).

1226 Threat to connectivity

1227 Hydropower stations, dams and weirs may all comprise major obstacles to migratory trout 1228 and cause marked alteration of riverine habitats. Building of roads, railways, bridges and 1229 other installations may incur migratory barriers for sea trout. In some cases, migrations may 1230 be completely blocked, whereas in others, culverts or other constructions may not be 1231 sufficiently well-designed to enable or facilitate fish passage (Larinier, 2002). River flow 1232 regulation for irrigation, freshwater fish hatcheries, channel modification to facilitate boat 1233 traffic (e.g. locks) and the building of infrastructure along rivers all pose challenges to 1234 migratory fish like those from hydropower regulation. There are numerous studies on 1235 restoration of spawning habitats, changes in overwintering environmental conditions, fish 1236 stranding due to rapid flow decreases, migration barriers, turbine mortality and the effects of 1237 flow and temperature regimes on juvenile growth, behaviour and recruitment of trout (e.g. 1238 (Aarestrup & Koed, 2003; Alonso-González, Gortázar, Sanz, & De Jalón, 2008; Pulg, 1239 Barlaup, Sternecker, Trepl, & Unfer, 2013; Saltveit, Halleraker, Arnekleiv, & Harby, 2001). 1240 Also, passages through hydrological barriers and temperature changes affect energy 1241 expenditure and decrease the fitness gain of migration. Furthermore, recent studies show that 1242 the presence of many dams negatively affect the energetic status of sea trout kelts during their 1243 seaward migration, reducing iteroparity (Haraldstad et al., 2018).

1244

1245 Coastal development

There is little information on how anthropogenic factors and increased human development activity in coastal areas affect sea trout survival and movements. It is not known how harbours, piers, bridges, fish farms, or industrial developments and deposits from mining activity in coastal areas affect sea trout. This is compounded by the construction of port facilities with piers or tethering systems that extend far offshore, as necessitated by the

increasing size of ocean-going vessels. The construction of tidal power turbines is currently
widely debated but the indirect impact of habitat changes associated with necessary damming
and the direct impact of the spinning blades could be substantial for migratory species like
trout. An increasing portion of rivers and the ocean is also now being illuminated at night. It
is not known how such facilities impact sea trout, but negative effects of light pollution have
been demonstrated on salmon migratory behaviour (Riley, Bendall, Ives, Edmonds, &
Maxwell, 2012) and may be expected to exert the same negative effects on trout.

1258

1259 Interactions among impact factors

1260 Often, several anthropogenic factors impact trout populations simultaneously, but studies of 1261 synergistic effects of anthropogenic disturbances on fish populations are often complicated 1262 and expensive to conduct, and very few such studies on any species exist. Synergistic effects of two or more impact factors are complex, non-linear and unpredictable, and certainly not 1263 1264 apparent from combining knowledge of the effects of single factors studied in isolation. As 1265 one example Finstad et al. (2012, 2007) investigated effects of salmon lice and acidification 1266 on Atlantic salmon post-smolt survival. These studies showed that smolts exposed to freshwater acidification were subsequently more vulnerable to salmon lice than were control 1267 1268 groups held in good quality freshwater before migrating to sea. Vulnerability to salmon lice 1269 was, however, reduced if there was a recovery period following the acidification episode and 1270 prior to exposure to salmon lice. The interaction between higher temperature and reduced sea 1271 louse generation time has been implicated in exacerbating the farm lice sea trout impact 1272 (Tully, 1992) in a context of global warming. The interaction between run timing of sea trout 1273 smolts and the point in time at which those smolts gain a sea louse burden has been 1274 demonstrated to lead to additional stress on those smolts (Poole et al., 2000). Therefore, we 1275 conclude that in years when environmental conditions are less favourable for sea trout

- 1276 transition from freshwater to saltwater and a high abundance of sea lice is present in the area,
- 1277 the combined impact on trout will be greater than in more "normal" years or locations.
- 1278

1279 Research Questions and future directions

1280 Assessment of partial anadromy

1281 In brown trout, partial anadromy is driven by complex interactions between intrinsic and 1282 extrinsic factors, leading to a wide variety of life history patterns. In particular, partial 1283 migration is influenced by juvenile growth in freshwater, largely determined by temperature 1284 and feeding opportunities (B. Jonsson, 1981) and resulting in a continuum of life history 1285 strategies in brown trout (Cucherousset et al., 2005). However, there are few empirical data 1286 sets available because studies rarely consider simultaneously sea trout and resident trout from 1287 the same population. The development of passive integrated transponder (PIT) tag monitoring 1288 programs across Europe is promising but currently limited to small to medium coastal 1289 streams or tributaries and do rarely encompass catchment or population level surveys. Scale 1290 samples and the relative concentrations of strontium (Sr) and calcium (Ca) in the otoliths can 1291 be used as proxies of movement of anadromous fish between freshwater and salt water. Also, 1292 investigations of nutritional status which differs between freshwater resident and anadromous 1293 fish or spawning ground surveys of types of spawners may offer alternative methods to study 1294 partial anadromy. In parallel, future research should explicitly incorporate both resident and 1295 anadromous individuals (and possibly all intermediate life histories), and use these in 1296 population models to improve our understanding of the complex dynamics within partly 1297 anadromous population. For instance, a recent meta-analysis highlighted that partial 1298 migration may confer a selective advantage in a stochastically varying environment (Gilroy, 1299 Gill, Butchart, Jones, & Franco, 2016). Thus, investigating whether partial anadromy under

chronic global change may promote a greater resilience in the brown trout compared toanadromous fish, should be an important issue for ecologists and managers.

1302

1303 Responses to climate change

1304 By influencing metabolic rate, growth and feeding opportunities, temperature is considered a 1305 key driver of migration decisions in brown trout. With climate change, temperature tends to 1306 increase more rapidly in rivers than in the ocean, but the effect of temperature on growth 1307 opportunities depends on whether it is above or below some optimal temperature. If 1308 temperature increases above the optimal temperature for growth, we hypothesize that the 1309 degree of anadromy will increase, provided that the risk of mortality in fresh relative to 1310 marine waters remains similar. If the temperature in freshwater is below the optimal 1311 temperature for growth, freshwater residency should be favoured as the temperature 1312 increases. The latter is the case in masu salmon, where the proportion of resident fish 1313 increases with temperature increase in freshwater (Morita et al., 2014). Furthermore, flow 1314 conditions are likely to interact with temperature, by either accentuating or attenuating the 1315 impact of temperature on brown trout depending on the situation. But surely, expected 1316 changes in flow regime would increase environmental stochasticity in freshwater. As a result, 1317 we predict that the influence of climate change on anadromy rates will differ from northern to 1318 southern populations in brown trout. The flexibility of partial anadromy is an issue for future 1319 research.

1320

Climate also influences the phenology of fish species (Anderson, Gurarie, Bracis, Burke, &
Laidre, 2013; B. Jonsson, Jonsson, & Jonsson, 2017). In a warmer climate, eggs may hatch
earlier (Alp, Erer, & Kamalak, 2010), the length of the growth season may increase until a
certain point (Elliott & Hurley, 1998), smolts should migrate earlier to sea (B. Jonsson &

1325 Ruud-Hansen, 1985; Otero et al., 2014) and adults return later for spawning (Alm, 1950), 1326 thereby increasing the duration of the sea-sojourn. In rivers, water flow appears to influence 1327 time of freshwater entry and upstream migration, especially in small streams (B. Jonsson et 1328 al., 2001, 2018). Furthermore, the tendency to spend the winter at sea may increase when the sea water temperature increases (Olsen et al., 2006). The spawning period in southern 1329 1330 populations may be prolonged as observed in Spain (Larios-López, de Figueroa, Galiana-1331 García, Gortázar, & Alonso, 2015). Investigations of latitudinal gradients in phenological and 1332 life history traits will help anticipate how climate change will influence sea trout in the future. 1333 However, because migratory distances to the home rivers differ, such gradient studies do not 1334 give direct evidence of changes in traits affected by migratory costs. Therefore, experimental 1335 research involving model species may also be helpful in studies of future phenology and life 1336 history of sea trout.

1337

1338 In a future warmer climate, age at smolting throughout much of the brown trout's distribution 1339 should decrease because of higher growth rate in freshwater; however, the opposite may be 1340 the case in the southern part of their distribution, as optimum temperature for growth will be exceeded more frequently. Whether smolt size will change or not, depends not only on 1341 1342 growth rate, but selection for optimal size depending on size-specific mortality at sea (B. 1343 Jonsson et al., 2016). If mortality of small smolts increases more than that of large smolts, 1344 selection will favour larger smolts and high survival at sea, and vice versa if mortality 1345 increases most for large smolts. Furthermore, the ability to osmoregulate may be impaired at higher temperatures (S. D. McCormick, Shrimpton, & Zydlewski, 1996) with possible full 1346 1347 loss of anadromy in southern areas, similar to Arctic charr.

1348

1349 Flow conditions in rivers will change in the future climate. With increasing flow in small 1350 streams, the size of both residents, smolts as well as adult trout may increase, and vice versa 1351 if the flow decreases (B. Jonsson et al., 2001). Furthermore, more water may influence the 1352 relative size of males and females. Apparently, males need more water than similar sized females to be reproductively successful because of their high activity. Therefore, males may 1353 1354 be smaller than females in small streams, and less so when stream size increases (B. Jonsson & Jonsson, 2015). Conversely, predicted increase in the frequency of extreme summer 1355 1356 drought is likely to jeopardize juvenile survival and growth. 1357 1358 Mean male size relative to mean female size of the anadromous trout may increase towards 1359 the north (B. Jonsson & Jonsson, 2015). This might be because conditions in freshwater make 1360 freshwater residency less favourable in the north. However, the mechanism determining the sex ratio of freshwater resident versus anadromous trout is still unknown and open for 1361 1362 research, as the proportion of the two varies among rivers (Table 1).

1363

1364 Epigenetics

1365 Increased winter temperature might influence embryo development and in this century,

1366 winter temperatures are expected to increase more than summer temperatures (IPCC, 2013).

1367 Heritable variation in phenotypic plasticity suggests that although increasing temperatures are

1368 likely to affect some populations negatively, they may have the potential to adapt to changing

1369 temperature regimes (L. F. Jensen et al., 2008). The mechanism of this flexibility is, however,

- 1370 unknown, but appears to involve a change in gene expression, possibly caused as an
- 1371 epigenetic response to early thermal conditions (B. Jonsson & Jonsson, 2014, 2016). There is
- 1372 still little knowledge about epigenetic influence on phenotypically plastic traits (B. Jonsson &

1373 Jonsson, 2019).

1375 Management strategies

58

1376 Stock assessments for brown trout are generally lacking across Europe. Little is known about 1377 productivity and recruitment in brown trout, as well as river specific carrying capacities. Data 1378 are lacking to quantify the mortality induced by fisheries at sea, in freshwater and as potential 1379 bycatch. There is also lack of information on sea trout stocks regarding human impacts (e.g. 1380 aquaculture and spread of diseases and parasites, pollution and water quality, agriculture, 1381 road, railway activities etc.). Estimates of straying among watercourses are sometimes large 1382 (Quéméré et al., 2016), but variable, possibly influenced by the size of the river and presence 1383 of lakes where trout spend the winter in a protected habitat. If straying between neighbouring 1384 populations is high, they may be managed together as meta-populations (Meier, Hansen, 1385 Bekkevold, Skaala, & Mensberg, 2011). However, in the future climate, autumn and winter 1386 precipitation may increase and thereby reduce the rate of straying with effects on the size of 1387 the management unit. As brown trout are partly anadromous, a future management program 1388 should be coordinate with data collection programs across Europe. To aid in its 1389 implementation, data collection suitable for supporting management of sea trout stocks 1390 should be cheap, easy and pragmatic, such as the Length Based Indicator (LBI) system 1391 proposed in Shephard et al. (2018).

1392

In many countries, sea trout management is viewed as a by-product of Atlantic salmon management, and this is completely disconnected from resident trout management. Thus, there is an urgent need to develop new recommendations dedicated to brown trout ecology and plasticity to promote the conservation of this iconic species. Because trout are only partly anadromous, more research is needed to define the relative contribution of sea trout and resident trout to population dynamics, as well as more general data comparing sea survival and growth in populations across the native distribution area. Preferably, this should be done
using individual tagged fish (such as PIT tagged) in order to directly couple life history to
individual parameters (and avoid problems with straying, age reading etc.). Future research
should define relevant management units and provide recommendations for dedicated
management actions able to account for the large plasticity in individual life history
trajectories in trout.

1405

1406 Acknowledgements

1407 This work is part of the ICES working group on sea trout WGTRUTTA. We thank Rafał 1408 Bernaś, Rasmus Lauridsen, Alan Walker, Steig Peterson, Jean-Luc Baglinière, Arne Johan 1409 Jensen and Kjell Leonardsson for their valued suggestions. We are extremely grateful to all 1410 the organisations that provided data: the Marine Institute, the Environmental Agency, the 1411 Norwegian Institute for Nature Research, the company Vattenfall Vattenkraft AB, the AFB 1412 and INRA as part of the Observatoire de Recherche en environnement DiaPFC. The 1413 contribution from MN was supported by INRA and European program Interreg France-1414 Channel-England (project: SAMARCH); BF, JD and BJ were supported by the Norwegian 1415 Environmental Agency, and JD was also funded by the Research Council of Norway (Project: 1416 CHASES); RP was supported by the Marine Institute, Ireland; RF was supported by a Marine 1417 Institute Cullen Fellowship Grant. QJ was funded by the French Agency for Biodiversity. 1418

1419 Data Availability Statement

1420 The data that support the findings of this study are available from the corresponding author1421 upon reasonable request.

1423 **References**

- 1424 Aarestrup, K., Baktoft, H., Koed, A., del Villar-Guerra, D., & Thorstad, E. B. (2014).
- 1425 Comparison of the riverine and early marine migration behaviour and survival of wild
- 1426 and hatchery-reared sea trout *Salmo trutta* smolts. *Marine Ecology Progress Series*,
- 1427 496, 197–206. https://doi.org/10.3354/meps10614
- Aarestrup, K., Birnie Gauvin, K., & Larsen, M. H. (2017). Another paradigm lost? Autumn
 downstream migration of juvenile brown trout: Evidence for a presmolt migration.
- 1430 *Ecology of Freshwater Fish*, 27(1), 513–516. https://doi.org/10.1111/eff.12335
- 1431 Aarestrup, K., & Jepsen, N. (1998). Spawning migration of sea trout (Salmo trutta (L)) in a
- 1432 Danish river. In *Advances in Invertebrates and Fish Telemetry* (pp. 275–281).
 1433 Springer.
- 1434 Aarestrup, K., & Koed, A. (2003). Survival of migrating sea trout (*Salmo trutta*) and Atlantic
 1435 salmon (*Salmo salar*) smolts negotiating weirs in small Danish rivers. *Ecology of*1436 *Freshwater Fish*, 12(3), 169–176.
- 1437 Aarestrup, K., Nielsen, C., & Koed, A. (2002). Net ground speed of downstream migrating
- 1438 radio-tagged Atlantic salmon (Salmo salar L.) and brown trout (Salmo trutta L.)
- smolts in relation to environmental factors. *Hydrobiologia*, 483(1), 95–102.
- 1440 https://doi.org/10.1023/A:1021306907338
- 1441 Aarestrup, K., Nielsen, C., & Madsen, S. S. (2000). Relationship between gill Na+,K+-
- 1442 ATPase activity and downstream movement in domesticated and first-generation
- 1443 offspring of wild anadromous brown trout (*Salmo trutta*). *Canadian Journal of*
- 1444 Fisheries and Aquatic Sciences, 57(10), 2086–2095. https://doi.org/10.1139/f00-164
- 1445 Acolas, M.-L., Labonne, J., Baglinière, J.-L., & Roussel, J. M. (2012). The role of body size
- 1446 versus growth on the decision to migrate: a case study with *Salmo trutta*.
- 1447 *Naturwissenschaften*, 99(1), 11–21. https://doi.org/10.1007/s00114-011-0861-5

1448	Acolas, ML., Roussel, JM., & Baglinière, JL. (2008). Linking migratory patterns and diet
1449	to reproductive traits in female brown trout (Salmo trutta L.) by means of stable
1450	isotope analysis on ova. Ecology of Freshwater Fish, 17(3), 382–393.
1451	https://doi.org/10.1111/j.1600-0633.2007.00290.x
1452	Aldvén, D., & Davidsen, J. (2017). Marine migrations of sea trout (Salmo trutta). In G.
1453	Harris (Ed.), Sea Trout: Science & Management: Proceedings of the 2nd
1454	International Sea Trout Symposium (pp. 267–276). Dundalk, Ireland: Troubador
1455	Publishing Ltd.
1456	Aldvén, D., Hedger, R., Økland, F., Rivinoja, P., & Höjesjö, J. (2015). Migration speed,
1457	routes, and mortality rates of anadromous brown trout Salmo trutta during outward
1458	migration through a complex coastal habitat. Marine Ecology Progress Series, 541,
1459	151–163.
1460	Alm, G. (1950). The sea-trout population in the Ava Stream. Ann. Rep. Inst. Freshwater Res.,
1461	Drottningholm, 31, 26–51.
1462	Alonso-González, C., Gortázar, J., Sanz, D. B., & De Jalón, D. G. (2008). Dam function rules
1463	based on brown trout flow requirements: design of environmental flow regimes in
1464	regulated streams. Hydrobiologia, 609(1), 253-262.
1465	Alp, A., Erer, M., & Kamalak, A. (2010). Eggs incubation, early development and growth in
1466	frys of brown trout (Salmo trutta macrostigma) and black sea trout (Salmo trutta
1467	labrax). Turkish Journal of Fisheries and Aquatic Sciences, 10(3).
1468	Álvarez, D., Cano, J. M., & Nicieza, A. G. (2006). Microgeographic variation in metabolic
1469	rate and energy storage of brown trout: countergradient selection or thermal
1470	sensitivity? Evolutionary Ecology, 20(4), 345-363. https://doi.org/10.1007/s10682-
1471	006-0004-1

- 1472 Anderson, J. J., Gurarie, E., Bracis, C., Burke, B. J., & Laidre, K. L. (2013). Modeling
- climate change impacts on phenology and population dynamics of migratory marine
 species. *Ecological Modelling*, 264, 83–97.
- 1475 Anon. (2008). A strategy for improved pest control on Irish salmon farms (p. 56pp). Dublin,
- 1476 Ireland: Department of Agriculture, Fisheries and Food.
- 1477 Artemov, A. V., Mugue, N. S., Rastorguev, S. M., Zhenilo, S., Mazur, A. M., Tsygankova, S.
- 1478 V., ... Prokhortchouk, E. B. (2017). Genome-Wide DNA Methylation Profiling
- 1479 Reveals Epigenetic Adaptation of Stickleback to Marine and Freshwater Conditions.
- 1480 *Molecular Biology and Evolution*, *34*(9), 2203–2213.
- 1481 https://doi.org/10.1093/molbev/msx156
- 1482 Baerwald, M. R., Meek, M. H., Stephens, M. R., Nagarajan, R. P., Goodbla, A. M., Tomalty,
- 1483 K. M. H., ... Nichols, K. M. (2016). Migration-related phenotypic divergence is
- 1484 associated with epigenetic modifications in rainbow trout. *Molecular Ecology*, 25(8),
- 1485 1785–1800. https://doi.org/10.1111/mec.13231
- 1486 Bagenal, T. B. (1969). Relationship between egg size and fry survival in brown trout Salmo
- 1487 *trutta* L. Journal of Fish Biology, 1(4), 349–353.
- 1488 Baglinière, J.-L. (1999). Introduction: The brown trout (Salmo trutta L)—its origin,
- 1489 distribution and economic and scientific significance. In J.-L. Baglinière & G. Maisse
- 1490 (Eds.), *Biology and Ecology of the Brown and Sea Trout* (pp. 1–12).
- 1491 https://doi.org/10.1007/978-1-4471-0775-0_1
- 1492 Baglinière, J.-L., & Arribe-Moutounet, D. (1985). Microrépartition des populations de truite
- 1493 commune (Salmo trutta L.) de juvenile de saumon atlantique (Salmo salar L.) et des
- 1494 autres espèces présentes dans la partie haute du Scorff (Bretagne). *Hydrobiologia*,
- 1495 *120*(3), 229–239. https://doi.org/10.1007/BF00045166

1496	Baglinière, JL., & Champigneulle, A. (1982). Densité des populations de truite commune
1497	(Salmo trutta L.) et de juvéniles de saumon atlantique (Salmo salar L.) sur le cours
1498	principal du Scorff (Bretagne): préférendums physiques et variations annuelles (1976-
1499	1980). Acta Oecologica: Oecologia Applicata.
1500	Baglinière, JL., Champigneulle, A., & Nihouarn, A. (1979). La fraie du saumon atlantique
1501	(Salmo salar L.) et de la truite commune (Salmo trutta L.) sur le bassin du Scorff.
1502	<i>Cybium</i> , <i>3</i> (7), 75–96.
1503	Baglinière, JL., Guyomard, R., Héland, M., Ombredane, D., & Prévost, E. (2001). Ecologie
1504	des populations de Poissons des cours d'eau à Salmonidés. L'eau Dans l'espace
1505	Rural. Vie et Milieux Aquatiques, A. Neveu, C. Riou, R. Bonhomme, P. Chassin, F.
1506	Papy (Eds), 31–49.
1507	Baglinière, JL., & Maisse, G. (1985). Precocious maturation and smoltification in wild
1508	Atlantic salmon in the Armorican massif, France. Aquaculture, 45(1), 249–263.
1509	https://doi.org/10.1016/0044-8486(85)90274-1
1510	Baglinière, JL., & Maisse, G. (1990). La croissance de la truite commune (Salmo trutta L.)
1511	sur le bassin de Scorff. Bulletin Français de la Pêche et de la Pisciculture, (318), 89-
1512	101. https://doi.org/10.1051/kmae:1990012
1513	Baglinière, JL., & Maisse, G. (1999). Biology and ecology of the brown and sea trout.
1514	Chichester, UK: Springer Praxis publishing Chichester.
1515	Baglinière, JL., & Maisse, G. (2002). La biologie de la truite commune (Salmo trutta L.)
1516	dans la rivière Scorff, Bretagne : une synthèse des études de 1972 à 1997. Productions
1517	Animales 5 (15), 319-331. (2002).

- 1518 Baglinière, J.-L., Prévost, E., & Maisse, G. (1994). Comparison of population dynamics of
- 1519 Atlantic salmon (*Salmo salar*) and brown trout (*Salmo trutta*) in a small tributary of

- 1520 the River Scorff (Brittany, France). *Ecology of Freshwater Fish*, *3*(1), 25–34.
- 1521 https://doi.org/10.1111/j.1600-0633.1994.tb00104.x
- Bakke, T. A., & Harris, P. D. (1998). Diseases and parasites in wild Atlantic salmon (*Salmo salar*) populations. *Canadian Journal of Fisheries and Aquatic Sciences*, 55(S1),
 247–266.
- 1525 Bal, G., Rivot, E., Prévost, E., Piou, C., & Baglinière, J.-L. (2011). Effect of water
- 1526 temperature and density of juvenile salmonids on growth of young-of-the-year
- 1527 Atlantic salmon Salmo salar. Journal of Fish Biology, 78(4), 1002–1022.
- 1528 https://doi.org/10.1111/j.1095-8649.2011.02902.x
- 1529 Balon, E. K. (1980). Early ontogeny of the lake chaar, *Salvelinus (Cristivomer) namaycush*.
- 1530 In Chaars: Salmonid fishes of the genus Salvelinus (pp. 475–685). The Hague,
- 1531 Netherlands: Dr. W. Junk Publishers.
- 1532 Baran, P., Delacoste, M., Lascaux, J. M., Bauba, F., & Segura, G. (1995). La compétition
- 1533 interspécifique entre la truite commune (Salmo trutta L.) et la truite arc-en-ciel
- 1534 (Oncorhynchus Mykiss Walbaum) : influence sur les modèles d'habitat. Bulletin
- 1535 Français de la Pêche et de la Pisciculture, (337-338–339), 283–290.
- 1536 https://doi.org/10.1051/kmae:1995032
- 1537 Bašić, T., Britton, J. R., Cove, R. J., Ibbotson, A., & Gregory, S. D. (2018). Roles of
- discharge and temperature in recruitment of a cold-water fish, the European grayling
- 1539 *Thymallus thymallus*, near its southern range limit. *Ecology of Freshwater Fish*,
- 1540 27(4), 940–951. https://doi.org/10.1111/eff.12405
- Beall, E., & De Gaudemar, B. (1999). Plasticity of reproductive behaviour in Atlantic salmon *Salmo salar* (Salmonidae) in relation to environmental factors. *Cybium*, 23(1), 9–28.
- 1543 Berejikian, B. A., Van Doornik, D. M., Endicott, R. C., Hoffnagle, T. L., Tezak, E. P.,
- 1544 Moore, M. E., & Atkins, J. (2010). Mating success of alternative male phenotypes and

- 1545 evidence for frequency-dependent selection in Chinook salmon, Oncorhynchus
- 1546 tshawytscha. Canadian Journal of Fisheries and Aquatic Sciences, 67(12), 1933–

1941. https://doi.org/10.1139/F10-112 1547

- 1548 Berg, O. K., & Jonsson, B. (1990). Growth and survival rates of the anadromous trout, Salmo 1549 trutta, from the Vardnes River, northern Norway. Environmental Biology of Fishes, 1550
- 29(2), 145–154.
- Berglund, I. (1995). Effects of size and spring growth on sexual maturation in 1+ Atlantic 1551 1552 salmon (Salmo salar) male parr: interactions with smoltification. Canadian Journal of
- 1553 Fisheries and Aquatic Sciences, 52(12), 2682–2694. https://doi.org/10.1139/f95-857
- 1554 Bernatchez, L. (2001). The Evolutionary History of Brown Trout (Salmo trutta L.) Inferred
- 1555 from Phylogeographic, Nested Clade, and Mismatch Analyses of Mitochondrial Dna
- 1556 Variation. Evolution, 55(2), 351-379. https://doi.org/10.1111/j.0014-
- 1557 3820.2001.tb01300.x
- 1558 Bird, A. (2002). DNA methylation patterns and epigenetic memory. Genes & Development, 1559 16(1), 6–21. https://doi.org/10.1101/gad.947102
- 1560 Birkeland, K. (1996). Consequences of premature return by sea trout (Salmo trutta) infested
- 1561 with the salmon louse (Lepeophtheirus salmonis Krøyer): migration, growth, and
- 1562 mortality. Canadian Journal of Fisheries and Aquatic Sciences, 53(12), 2808–2813.
- 1563 Birnie Gauvin, K., & Aarestrup, K. (2018). A call for a paradigm shift: Assumed-to-be
- 1564 premature migrants actually yield good returns. Ecology of Freshwater Fish, O(0).
- 1565 https://doi.org/10.1111/eff.12431
- Blanchet, S., Loot, G., Grenouillet, G., & Brosse, S. (2007). Competitive interactions 1566
- 1567 between native and exotic salmonids: a combined field and laboratory demonstration.
- Ecology of Freshwater Fish, 16(2), 133–143. 1568

1569	Boeuf, G., & Harache, Y. (1982). Criteria for adaptation of salmonids to high salinity
1570	seawater in France. Aquaculture, 28(1), 163–176. https://doi.org/10.1016/0044-
1571	8486(82)90019-9

- 1572 Bohlin, T., Dellefors, C., & Faremo, U. (1990). Large or small at maturity—theories on the
- 1573 choice of alternative male strategies in anadromous salmonids. *Annales Zoologici*1574 *Fennici*, 139–147. JSTOR.
- Bohlin, T., Pettersson, J., & Degerman, E. (2001). Population density of migratory and
 resident brown trout (*Salmo trutta*) in relation to altitude: evidence for a migration
 cost. *Journal of Animal Ecology*, 70(1), 112–121.
- 1578 Bordeleau, X., Davidsen, J. G., Eldøy, S. H., Sjursen, A. D., Whoriskey, F. G., & Crossin, G.

1579 T. (2018). Nutritional correlates of spatiotemporal variations in the marine habitat use

1580 of brown trout (*Salmo trutta*) veteran migrants. *Canadian Journal of Fisheries and*

1581 Aquatic Sciences, 75, 1744–1754. https://doi.org/10.1139/cjfas-2017-0350

Borgstrøm, R., & Jan, H. (1988). Smoltification of sea trout (*Salmo trutta*) at short length as
an adaptation to extremely low summer stream flow. *Polskie Archwum Hydrobiologii*,
35, 375–384.

1585 Buffington, J. M., Montgomery, D. R., & Greenberg, H. M. (2004). Basin-scale availability

1586 of salmonid spawning gravel as influenced by channel type and hydraulic roughness

- in mountain catchments. *Canadian Journal of Fisheries and Aquatic Sciences*, 61(11),
 2085–2096. https://doi.org/10.1139/f04-141
- 1589 Byrne, C. J., Poole, R., Dillane, M., Rogan, G., & Whelan, K. F. (2004). Temporal and
- 1590 environmental influences on the variation in sea trout (*Salmo trutta* L.) smolt
- 1591 migration in the Burrishoole system in the west of Ireland from 1971 to 2000.

1592 *Fisheries Research*, 66(1), 85–94.

1593	Byrne, C. J., Poole, W. R., Dillane, M. G., & Whelan, K. F. (2002). The Irish sea trout
1594	enhancement programme: an assessment of the parr stocking programme into the
1595	Burrishoole catchment. Fisheries Management and Ecology, 9(6), 329–341.
1596	https://doi.org/10.1046/j.1365-2400.2002.00314.x
1597	Campbell, J. S. (1977). Spawning characteristics of brown trout and sea trout Salmo trutta L.
1598	in Kirk Burn, River Tweed, Scotland. Journal of Fish Biology, 11(3), 217–229.
1599	Campbell, R. N. (1979). Ferox trout, Salmo trutta L., and charr, Salvelinm alpinus (L.), in
1600	Scottish lochs. Journal of Fish Biology, 14(1), 1–29. https://doi.org/10.1111/j.1095-
1601	8649.1979.tb03491.x
1602	Champigneulle, A., Buttiker, B., Durand, P., & Melhaoui, M. (1999). Main characteristics of
1603	the biology of the trout (Salmo trutta L.) in Lake Léman (Lake Geneva) and some of
1604	its tributaries. In JL. Baglinière & G. Maisse (Eds.), Biology and Ecology of the
1605	Brown and Sea Trout (pp. 147–174). https://doi.org/10.1007/978-1-4471-0775-0_6
1606	Chapman, B. B., Hulthén, K., Brodersen, J., Nilsson, P. A., Skov, C., Hansson, LA., &
1607	Brönmark, C. (2012). Partial migration in fishes: causes and consequences. Journal of
1608	Fish Biology, 81(2), 456–478. https://doi.org/10.1111/j.1095-8649.2012.03342.x
1609	Charles, K., Guyomard, R., Hoyheim, B., Ombredane, D., & Baglinière, JL. (2005). Lack of
1610	genetic differentiation between anadromous and resident sympatric brown trout
1611	(Salmo trutta) in a Normandy population. Aquatic Living Resources, 18(1), 65-69.
1612	https://doi.org/10.1051/alr:2005006
1613	Charles, K., Roussel, JM., Lebel, JM., Baglinière, JL., & Ombredane, D. (2006). Genetic
1614	differentiation between anadromous and freshwater resident brown trout (Salmo trutta
1615	L.): insights obtained from stable isotope analysis. Ecology of Freshwater Fish, 15(3),

1616 255–263. https://doi.org/10.1111/j.1600-0633.2006.00149.x

- 1617 Courter, I., Justice, C., & Cramer, S. (2009). *Flow and temperature effects on life history* 1618 *diversity of* Oncorhynchus mykiss *in the Yakima River basin*. Gresham, Oregon.
- 1619 Crête-Lafrenière, A., Weir, L. K., & Bernatchez, L. (2012). Framing the Salmonidae Family
- 1620 Phylogenetic Portrait: A More Complete Picture from Increased Taxon Sampling.
- 1621 PLOS ONE, 7(10), e46662. https://doi.org/10.1371/journal.pone.0046662
- 1622 Crisp, D. T., & Carling, P. A. (1989). Observations on siting, dimensions and structure of
 1623 salmonid redds. *Journal of Fish Biology*, *34*(1), 119–134.
- 1624 Cross, T. F., Mills, C. P. R., & Williams, M. de C. (1992). An intensive study of allozyme
- 1625 variation in freshwater resident and anadromous trout, *Salmo trutta* L., in western
- 1626 Ireland. Journal of Fish Biology, 40(1), 25–32. https://doi.org/10.1111/j.1095-
- 1627 8649.1992.tb02550.x
- 1628 Cucherousset, J., Ombredane, D., Charles, K., Marchand, F., & Baglinière, J.-L. (2005). A
 1629 continuum of life history tactics in a brown trout (*Salmo trutta*) population. *Canadian*1630 *Journal of Fisheries and Aquatic Sciences*, 62(7), 1600–1610.
- 1631 https://doi.org/10.1139/f05-057
- 1632 Dahl, K. (1904). A study on trout and young salmon. *Nyt Magazin for Naturvidenskaberne*,
 1633 42, 221–338.
- Davaine, P., & Beall, E. (1997). Introduction de salmonidés en milieu vierge (Îles Kerguelen,
 Subantarctique) : enjeux, résultats, perspectives. *Bulletin Français de la Pêche et de la Pisciculture*, (344–345), 93–110. https://doi.org/10.1051/kmae:1997013
- 1637 Davidsen, J. G., Daverdin, M., Sjursen, A. D., Rønning, L., Arnekleiv, J. V., & Koksvik, J. I.
- 1638 (2014). Does reduced feeding prior to release improve the marine migration of
- hatchery brown trout *Salmo trutta* smolts? *Journal of Fish Biology*, 85(6), 1992–2002.
- 1640 https://doi.org/10.1111/jfb.12485

- Davidson, I. C., Hazlewood, M. S., & Cove, R. J. (2007). Predicted Growth of Juvenile Trout
 and Salmon in Four Rivers in England and Wales Based on Past and Possible Future
- 1643 Temperature Regimes Linked to Climate Change. In *Sea Trout: Biology*,
- 1644 *Conservation and Management* (pp. 401–416).
- 1645 https://doi.org/10.1002/9780470996027.ch28
- 1646 de Eyto, E., Dalton, C., Dillane, M., Jennings, E., McGinnity, P., O'Dwyer, B., ... Taylor, D.
- 1647 (2016). The response of North Atlantic diadromous fish to multiple stressors,
- 1648 including land use change: a multidecadal study. *Canadian Journal of Fisheries and*
- 1649 Aquatic Sciences, 73(12), 1759–1769. https://doi.org/10.1139/cjfas-2015-0450
- 1650 del Villar-Guerra, D., Aarestrup, K., Skov, C., & Koed, A. (2013). Marine migrations in
- 1651 anadromous brown trout (*Salmo trutta*). Fjord residency as a possible alternative in
- 1652 the continuum of migration to the open sea. *Ecology of Freshwater Fish*, 23(4), 594–
- 1653 603. https://doi.org/10.1111/eff.12110
- 1654 Dellefors, C. (1996). Smoltification and sea migration in wild and hatchery-reared brown
- 1655 *trout*, Salmo trutta (Ph.D. thesis). University of Göteborg, Göteborg.
- Dellefors, C., & Faremo, U. (1988). Early sexual maturation in males of wild sea trout, *Salmo trutta* L., inhibits smoltification. *Journal of Fish Biology*, *33*(5), 741–749.
- Dieperink, C., Bak, B. D., Pedersen, L.-F., Pedersen, M. I., & Pedersen, S. (2002). Predation
 on Atlantic salmon and sea trout during their first days as postsmolts. *Journal of Fish Biology*, *61*(3), 848–852.
- 1661 Dieperink, C., Pedersen, S., & Pedersen, M. I. (2001). Estuarine predation on radiotagged
- wild and domesticated sea trout (*Salmo trutta* L.) smolts. *Ecology of Freshwater Fish*, *10*(3), 177–183.

1664	Dodson, J. J., Aubin-Horth, N., Thériault, V., & Páez, D. J. (2013). The evolutionary ecology
1665	of alternative migratory tactics in salmonid fishes. Biological Reviews of the

1666 *Cambridge Philosophical Society*, 88(3), 602–625. https://doi.org/10.1111/brv.12019

1667 Dominey, W. J. (1984). Alternative mating tactics and evolutionarily stable strategies.

1668 *American Zoologist*, 24(2), 385–396.

- 1669 Egglishaw, H. J., & Shackley, P. E. (1982). Influence of water depth on dispersion of juvenile
- 1670 salmonids, Salmo salar L. and S. trutta L., in a Scottish stream. Journal of Fish

1671 *Biology*, 21(2), 141–155. https://doi.org/10.1111/j.1095-8649.1982.tb03995.x

- 1672 Einum, S., Hendry, A. P., & Fleming, I. A. (2002). Egg-size evolution in aquatic
- 1673 environments: does oxygen availability constrain size? *Proceedings of the Royal*1674 Society of London B: Biological Sciences, 269(1507), 2325–2330.
- 1675 Eldøy, S. H., Davidsen, J. G., Thorstad, E. B., Whoriskey, F. G., Aarestrup, K., Naesje, T. F.,

1676 ... Arnekleiv, J. V. (2017). Marine depth use of sea trout *Salmo trutta* in fjord areas of

- 1677 central Norway: marine depth use of *salmo trutta*. *Journal of Fish Biology*, *91*, 1268–
- 1678 1283. https://doi.org/10.1111/jfb.13463
- 1679 Elliott, J. M. (1984). Numerical changes and population regulation in young migratory trout
- Salmo trutta in a Lake District stream, 1966-83. *The Journal of Animal Ecology*, 327–
 350.
- Elliott, J. M. (1993). A 25-year study of production of juvenile sea-trout, *Salmo trutta*, in an
 English Lake District stream. *Canadian Special Publication of Fisheries and Aquatic Sciences*, 109–122.
- 1685 Elliott, J. M. (1994). *Quantitative Ecology and the Brown Trout*. Oxford University Press,
 1686 USA.
- 1687 Elliott, J. M., & Elliott, J. A. (2010). Temperature requirements of Atlantic salmon Salmo
 1688 salar, brown trout Salmo trutta and Arctic charr Salvelinus alpinus: predicting the

- 1689 effects of climate change. *Journal of Fish Biology*, 77(8), 1793–1817.
- 1690 https://doi.org/10.1111/j.1095-8649.2010.02762.x
- 1691 Elliott, J. M., & Hurley, M. A. (1998). Predicting fluctuations in the size of newly emerged
 1692 sea-trout fry in a Lake District stream. *Journal of Fish Biology*, *53*(5), 1120–1133.
- 1693 Engqvist, L., & Taborsky, M. (2016). The evolution of genetic and conditional alternative
 1694 reproductive tactics. *Proc. R. Soc. B*, 283(1825), 20152945.
- 1695 Esteve, M. (2005). Observations of spawning behaviour in Salmoninae: Salmo,
- 1696 Oncorhynchus and Salvelinus. Reviews in Fish Biology and Fisheries, 15(1–2), 1–21.
- 1697 Euzenat, G., Fournel, F., & Fagard, J. L. (2006). Population dynamics and stock-recruitment
- relationship of sea trout in the River Bresle, upper Normandy, France. In *Sea Trout*. *Biology, Conservation and Management* (pp. 307–323).
- 1700 Euzenat, G., Fournel, F., & Richard, A. (1999). Sea trout (Salmo trutta L.) in Normandy and
- 1701 Picardy. In J.-L. Baglinière & G. Maisse (Eds.), *Biology and Ecology of the Brown*
- 1702 and Sea Trout (pp. 175–203). https://doi.org/10.1007/978-1-4471-0775-0_7
- 1703 Evangelista, C., Boiche, A., Lecerf, A., & Cucherousset, J. (2014). Ecological opportunities
- and intraspecific competition alter trophic niche specialization in an opportunistic
- 1705 stream predator. *Journal of Animal Ecology*, 83(5), 1025–1034.
- 1706 https://doi.org/10.1111/1365-2656.12208
- 1707 Fahy, E. (1985). Child of the Tides: a sea trout handbook. Glendale Press.
- 1708 Fealy, R., Allot, N., Borderick, C., deEyto, E., Dillane, M., Erdil, R. M., ... White, J. (2010).
- 1709 *RESCALE: Review and Simulate Climate and Catchment Responses at Burrishoole.*
- 1710 Retrieved from https://oar.marine.ie/handle/10793/31
- 1711 Ferguson, A. (2004). The Importance of Identifying Conservation Units: Brown Trout and
- 1712 Pollan Biodiversity in Ireland. *Biology and Environment: Proceedings of the Royal*
- 1713 Irish Academy, 104B(3), 33–41.

- 72
- 1714 Ferguson, A. (2006). Genetics of Sea Trout, with Particular Reference to Britain and Ireland.
- 1715 In Sea Trout: Biology, Conservation and Management (pp. 155–182).
- 1716 https://doi.org/10.1002/9780470996027.ch12
- 1717 Ferguson, A., Reed, T., Cross, T. F., McGinnity, P., & Prodöhl, P. (2019). Anadromy,
- potamodromy and residency in brown trout *Salmo trutta*: the role of genes and the
 environment. *Journal of Fish Biology*. https://doi.org/10.1111/jfb.14005
- 1720 Ferguson, A., Reed, T., McGinnity, P., & Prodöhl, P. (2017). Anadromy in brown trout
- 1721 (Salmo trutta): A review of the relative roles of genes and environmental factors and
 1722 the implications for management and conservation.
- 1723 Finstad, A. G., Forseth, T., Jonsson, B., Bellier, E., Hesthagen, T., Jensen, A. J., ... Foldvik,

1724 A. (2011). Competitive exclusion along climate gradients: energy efficiency

- influences the distribution of two salmonid fishes. *Global Change Biology*, *17*(4),
 1726 1703–1711.
- Finstad, A. G., & Jonsson, B. (2012). Effect of incubation temperature on growth
 performance in Atlantic salmon. *Marine Ecology Progress Series*, 454, 75–82.
- 1729 Finstad, B., & Bjørn, P. A. (2011). Present status and implications of salmon lice on wild
- 1730 salmonids in Norwegian coastal zones. *Salmon Lice: An Integrated Approach to*1731 *Understanding Parasite Abundance and Distribution*, 279–305.
- 1732 Finstad, B., Kroglund, F., Bjørn, P. A., Nilsen, R., Pettersen, K., Rosseland, B. O., ... Salbu,
- B. (2012). Salmon lice-induced mortality of Atlantic salmon postsmolts experiencing
 episodic acidification and recovery in freshwater. *Aquaculture*, *362*, 193–199.
- 1735 Finstad, B., Kroglund, F., Strand, R., Stefansson, S. O., Bjørn, P. A., Rosseland, B. O., ...
- 1736 Salbu, B. (2007). Salmon lice or suboptimal water quality—Reasons for reduced
- 1737 postsmolt survival? *Aquaculture*, 273(2–3), 374–383.

1738	Fjørtoft, H. I	B., Besnier	: F.	Stene. A.	. Nilsen.	F	Biørn.	P. A.	. Tveten	. AK.	Glover	. K.
1,20	1 1010010, 110 1			,,	, 1,110011,	··,	D prin,		, 1,000011	,	$, \ldots \circ \circ \circ \circ \circ$, <u></u> .

- 1739 A. (2017). The Phe362Tyr mutation conveying resistance to organophosphates occurs
- in high frequencies in salmon lice collected from wild salmon and trout. *Scientific*
- 1741 *Reports*, 7(1), 14258. https://doi.org/10.1038/s41598-017-14681-6
- 1742 Flaten, A. C., Davidsen, J. G., Thorstad, E. B., Whoriskey, F., Rønning, L., Sjursen, A. D., ...
- 1743 Arnekleiv, J. V. (2016). The first months at sea: marine migration and habitat use of
- sea trout *Salmo trutta* post-smolts. *Journal of Fish Biology*, 89(3), 1624–1640.
- 1745 https://doi.org/10.1111/jfb.13065
- 1746 Fleming, I. A. (1996). Reproductive strategies of Atlantic salmon: ecology and evolution.
- 1747 *Reviews in Fish Biology and Fisheries*, 6(4), 379–416.
- 1748 https://doi.org/10.1007/BF00164323
- Fleming, I. A., & Gross, M. R. (1990). Latitudinal Clines: A Trade-Off between Egg Number
 and Size in Pacific Salmon. *Ecology*, *71*(1), 1. https://doi.org/10.2307/1940241
- 1751 Foote, C. J., Brown, G. S., & Wood, C. C. (1997). Spawning success of males using
- 1752 alternative mating tactics in sockeye salmon, *Oncorhynchus nerka*. *Canadian Journal*
- 1753 *of Fisheries and Aquatic Sciences*, 54(8), 1785–1795. https://doi.org/10.1139/f97-080
- Forseth, T., & Jonsson, B. (1994). The Growth and Food Ration of Piscivorous Brown Trout
 (*Salmo trutta*). *Functional Ecology*, 8(2), 171–177. https://doi.org/10.2307/2389900
- 1756 Forseth, T., Larsson, S., Jensen, A. J., Jonsson, B., Näslund, I., & Berglund, I. (2009).
- 1757 Thermal growth performance of juvenile brown trout *Salmo trutta*: no support for
- thermal adaptation hypotheses. *Journal of Fish Biology*, 74(1), 133–149.
- 1759 https://doi.org/10.1111/j.1095-8649.2008.02119.x
- 1760 Forseth, T., Nesje, T. F., Jonsson, B., & Hårsaker, K. (1999). Juvenile migration in brown
- trout: a consequence of energetic state. *Journal of Animal Ecology*, 68(4), 783–793.
- 1762 https://doi.org/10.1046/j.1365-2656.1999.00329.x

- Frier, J. O. (1994). Growth of Anadromaus and Resident Brown Trout with Different Life
 Histories in a Danish Lowland Stream. *Nordic Journal of Freshwater Research*, 64,
 58–70.
- 1766 Frost, W. E., & Brown, M. E. (1967). *The Trout*. HarperCollins Publishers Limited.
- Fusco, G., & Minelli, A. (2010). Phenotypic plasticity in development and evolution: facts
 and concepts. *Philosophical Transactions of the Royal Society of London B:*

1769 Biological Sciences, 365(1540), 547–556. https://doi.org/10.1098/rstb.2009.0267

- Gargan, P. G., Poole, W. R., & Forde, G. P. (2006). A Review of the Status of Irish Sea Trout
 Stocks. In *Sea Trout: Biology, Conservation and Management* (pp. 25–44).
- 1772 https://doi.org/10.1002/9780470996027.ch3
- Gargan, P. G., Tully, O., & Poole, W. R. (2003). Relationship between sea lice infestation,
 sea lice production and sea trout survival in Ireland, 1992–2001. *Salmon at the Edge*,
- 1775 *10*, 119–135.
- 1776 Gauthey, Z., Lang, M., Elosegi, A., Tentelier, C., Rives, J., & Labonne, J. (2015). Brown

1777 trout spawning habitat selection and its effects on egg survival. *Ecology of*

1778 *Freshwater Fish*, 26(1), 133–140. https://doi.org/10.1111/eff.12262

- Gibson, R. J., & Erkinaro, J. (2009). The influence of water depths and inter-specific
 interactions on cover responses of juvenile Atlantic salmon. *Ecology of Freshwater Fish*, 18(4), 629–639.
- 1782 Giger, T., Excoffier, L., Day, P. J. R., Champigneulle, A., Hansen, M. M., Powell, R., &
- Largiadèr, C. R. (2006). Life history shapes gene expression in salmonids. *Current Biology: CB*, *16*(8), R281-282. https://doi.org/10.1016/j.cub.2006.03.053
- 1785 Gilroy, J. J., Gill, J. A., Butchart, S. H., Jones, V. R., & Franco, A. M. (2016). Migratory
- 1786 diversity predicts population declines in birds. *Ecology Letters*, *19*(3), 308–317.

- Gortázar, J., Alonso, C., & Jalón, D. G. de. (2012). Brown trout redd superimposition in
 relation to spawning habitat availability. *Ecology of Freshwater Fish*, 21(2), 283–292.
- 1789 https://doi.org/10.1111/j.1600-0633.2011.00546.x
- 1790 Gosset, C., Rives, J., & Labonne, J. (2006). Effect of habitat fragmentation on spawning
- 1791 migration of brown trout (Salmo trutta L.). Ecology of Freshwater Fish, 15(3), 247–
- 1792 254. https://doi.org/10.1111/j.1600-0633.2006.00144.x
- Graham, C. T., & Harrod, C. (2009). Implications of climate change for the fishes of the
 British Isles. *Journal of Fish Biology*, 74(6), 1143–1205.
- 1795 Greenberg, L., Svendsen, P., & Harby, A. (1996). Availability of microhabitats and their use
- by brown trout (*Salmo trutta*) and grayling (*Thymallus thymallus*). *Regulated Rivers: Research and Management 12: 287-303, 12, 287–303.*
- Gregory, J. S., & Gamett, B. L. (2009). Cattle Trampling of Simulated Bull Trout Redds.
 North American Journal of Fisheries Management, 29(2), 361–366.
- 1800 https://doi.org/10.1577/M08-020.1
- 1801 Gresswell, R. E., Liss, W. J., & Larson, G. L. (1994). Life-history organization of
- 1802 Yellowstone cutthroat trout (*Oncorhynchus clarki bouvieri*) in Yellowstone Lake.
- 1803 *Canadian Journal of Fisheries and Aquatic Sciences*, *51*(S1), 298–309.
- 1804 Gross, M. R. (1985). Disruptive selection for alternative life histories in salmon. *Nature*,
- 1805 *313*(5997), 47–48. https://doi.org/10.1038/313047a0
- 1806 Gross, M. R. (1987). Evolution of diadromy in fishes. American Fisheries Society
- 1807 *Symposium*, 1, 14–25.

66.

- 1808 Gross, M. R. (1991). Evolution of alternative reproductive strategies: frequency-dependent
- 1809 sexual selection in male bluegill sunfish. Phil. Trans. R. Soc. Lond. B, 332(1262), 59–
- 1810

- 1811 Gross, M. R., Coleman, R. M., & McDowall, R. M. (1988). Aquatic Productivity and the
- 1812 Evolution of Diadromous Fish Migration. *Science*, *239*(4845), 1291–1293.
- 1813 https://doi.org/10.1126/science.239.4845.1291
- 1814 Halttunen, E., Gjelland, K.-Ø., Hamel, S., Serra-Llinares, R.-M., Nilsen, R., Arechavala-
- 1815 Lopez, P., ... Finstad, B. (2018). Sea trout adapt their migratory behaviour in
- 1816 response to high salmon lice concentrations. *Journal of Fish Diseases*, 41(6), 953–
- 1817 967. https://doi.org/10.1111/jfd.12749
- 1818 Haraldstad, T., Höglund, E., Kroglund, F., Lamberg, A., Olsen, E. M., & Haugen, T. O.
- 1819 (2018). Condition-dependent skipped spawning in anadromous brown trout (Salmo
- 1820 *trutta*). Canadian Journal of Fisheries and Aquatic Sciences, 1–7.
- 1821 https://doi.org/10.1139/cjfas-2017-0076
- 1822 Haraldstad, T., Kroglund, F., Kristensen, T., Jonsson, B., & Haugen, T. O. (2017). Diel
- 1823 migration pattern of Atlantic salmon (Salmo salar) and sea trout (Salmo trutta)
- 1824 smolts: an assessment of environmental cues. *Ecology of Freshwater Fish*, 26(4),
- 1825 541–551. https://doi.org/10.1111/eff.12298
- Harris, G. (2017). Sea Trout: Science & Management: Proceedings of the 2nd International
 Sea Trout Symposium. S.l.: Matador.
- Harris, G., & Milner, N. (2006). Sea Trout: Biology, Conservation and Management. John
 Wiley & Sons.
- Harrison, L. R., Legleiter, C. J., Wydzga, M. A., & Dunne, T. (2011). Channel dynamics and
 habitat development in a meandering, gravel bed river. *Water Resources Research*,
 47(4).
- 1833 Harwood, A. J., Metcalfe, N. B., Armstrong, J. D., & Griffiths, S. W. (2001). Spatial and
- 1834 temporal effects of interspecific competition between Atlantic salmon (*Salmo salar*)

- and brown trout (*Salmo trutta*) in winter. *Canadian Journal of Fisheries and Aquatic Sciences*, 58(6), 1133–1140.
- Haury, J., Ombredane, D., & Baglinière, J.-L. (1999). The habitat of the brown trout (*Salmo trutta* L.) in water courses. In *Biology and ecology of the brown and sea trout* (pp.
- 1839 37–89). Springer.
- Hearn, W. E. (1987). Interspecific competition and habitat segregation among streamdwelling trout and salmon: a review. *Fisheries*, *12*(5), 24–31.
- 1842 Heggberget, T. G. (1984). Habitat selection and segregation of part of Arctic charr
- 1843 (*Salvelinus alpinus*), brown trout (*Salmo trutta*) and Atlantic salmon (*Salmo salar* L.)
 1844 in two streams in North Norway. *Biology of the Arctic Charr*, 217–231.
- 1845 Heggenes, J., Baglinière, J.-L., & Cunjak, R. A. (1999). Spatial niche variability for young
- 1846 Atlantic salmon (*Salmo salar*) and brown trout (*S. trutta*) in heterogeneous streams.
 1847 *Ecology of Freshwater Fish*, 8(1), 1–21.
- Hendry, A. P., & Day, T. (2003). Revisiting the positive correlation between female size and
 egg size. *Evolutionary Ecology Research*, 5(3), 421–429.
- Henrikson, L., & Brodin, Y.-W. (1995). Liming of surface waters in Sweden a synthesis.
 In *Liming of Acidified Surface Waters* (pp. 1–44). https://doi.org/10.1007/978-3-64279309-7_1
- 1853 Hessle, C. (1935). Gotlands Havlaxöring. *Kungl Lantbruksstyrelsen*, (7), 1–12.
- Hesthagen, T., & Heggenes, J. (2003). Competitive habitat displacement of brown trout by
 Siberian sculpin: the role of size and density. *Journal of Fish Biology*, 62(1), 222–
 236.
- 1857 Hesthagen, T., Larsen, B. M., Bolstad, G., Fiske, P., & Jonsson, B. (2017). Mitigation of
- 1858 acidified salmon rivers–effects of liming on young brown trout *Salmo trutta*. *Journal*
- 1859 *of Fish Biology*, 91(5), 1350–1364.

1860	Heywood, M. J. T., &	& Walling, D. E. (2007)	. The sedimentation of salmonic	1 spawning
------	----------------------	-------------------------	---------------------------------	------------

- 1861 gravels in the Hampshire Avon catchment, UK: implications for the dissolved oxygen
- 1862 content of intragravel water and embryo survival. *Hydrological Processes*, 21(6),
- 1863 770–788. https://doi.org/10.1002/hyp.6266
- 1864 Hindar, K., Jonsson, B., Ryman, N., & Ståhl, G. (1991). Genetic relationships among
- 1865 landlocked, resident, and anadromous Brown Trout, *Salmo trutta* L. *Heredity*, 66(1),
- 1866 83–91. https://doi.org/10.1038/hdy.1991.11
- 1867 Höjesjö, J., Stradmeyer, L., Griffiths, S., & Armstrong, J. (2010). Effects of Competing
- 1868Brown Trout on Habitat Use by Atlantic Salmon Parr: Controlled Investigations in a
- 1869Laboratory Stream. In Salmonid Fisheries (pp. 194–208).
- 1870 https://doi.org/10.1002/9781444323337.ch9
- Holmen, J., Olsen, E. M., & Vøllestad, L. A. (2003). Interspecific competition between
 stream-dwelling brown trout and Alpine bullhead. *Journal of Fish Biology*, 62(6),
 1312–1325.
- Houde, A. L. S., Wilson, C. C., & Neff, B. D. (2017). Performance of four salmonids species
 in competition with Atlantic salmon. *Journal of Great Lakes Research*, 43(1), 211–
- 1876 215.
- 1877 Hughes, M. R., Hooker, O. E., Leeuwen, T. E. V., Kettle-White, A., Thorne, A., Prodöhl, P.,
- 1878 & Adams, C. E. (2019). Alternative routes to piscivory: Contrasting growth
- 1879 trajectories in brown trout (*Salmo trutta*) ecotypes exhibiting contrasting life history
- 1880 strategies. *Ecology of Freshwater Fish*, 28(1), 4–10. https://doi.org/10.1111/eff.12421
- Hutchings, J. A., & Myers, R. A. (1985). Mating between anadromous and nonanadromous
 Atlantic salmon, *Salmo salar*. *Canadian Journal of Zoology*, 63(9), 2219–2221.
- 1883 Hutchings, J. A., & Myers, R. A. (1994). The evolution of alternative mating strategies in
- 1884 variable environments. *Evolutionary Ecology*, 8(3), 256–268.

- ICES. (1997). Report of the Workshop on the Interactions between salmon lice and salmonids
 (No. ICES CM 1997/M:4 Ref.:F). Edinburgh, United Kingdom, 11-15 November
 1887 1996.
- 1888 ICES. (2018). *Report of the Baltic Salmon and Trout Assessment Working Group (WGBAST)*1889 (p. 371p). Turku, Finland.
- 1890 IPCC. (2013). Climate change 2013: the physical science basis. Contribution of Working
- 1891 Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate
- 1892 Change. K., Tignor, M., Allen, SK, Boschung, J., Nauels, A., Xia, Y., Bex, V., and
- 1893 Midgley, PM, Cambridge University Press Cambridge, United Kingdom and New
 1894 York, NY, USA.
- 1895 Jensen, A. J., Finstad, B., Fiske, P., Hvidsten, N. A., Rikardsen, A. H., & Saksgård, L.
- 1896 (2012). Timing of smolt migration in sympatric populations of Atlantic salmon
- 1897 (*Salmo salar*), brown trout (*Salmo trutta*), and Arctic char (*Salvelinus alpinus*).

1898 *Canadian Journal of Fisheries and Aquatic Sciences*, 69(4), 711–723.

- 1899 Jensen, J. L. A., & Rikardsen, A. H. (2012). Archival tags reveal that Arctic charr Salvelinus
- *alpinus* and brown trout *Salmo trutta* can use estuarine and marine waters during
 winter. *Journal of Fish Biology*, *81*(2), 735–749.
- Jensen, K. W. (1968). *Seatrout* (Salmo trutta, *L.*) of the River Istra, western Norway. National
 Swedish Board of Fisheries.
- 1904 Jensen, L. F., Hansen, M. M., Pertoldi, G., Holdensgaard, G., Mensberg, K. D., & Loeschcke,
- 1905 V. (2008). Local adaptation in brown trout early life-history traits: implications for
- 1906 climate change adaptability. *Proceedings of the Royal Society B: Biological Sciences*,
- 1907 275(1653), 2859–2868. https://doi.org/10.1098/rspb.2008.0870

- Jepsen, N., Holthe, E., & Økland, F. (2006). Observations of predation on salmon and trout
 smolts in a river mouth. *Fisheries Management and Ecology*, *13*(5), 341–343.
 https://doi.org/10.1111/j.1365-2400.2006.00509.x
- 1911 Johnsson, J. I., Rydeborg, A., & Sundström, L. F. (2004). Predation risk and the territory
- 1912 value of cover: an experimental study. *Behavioral Ecology and Sociobiology*, 56(4),
- 1913 388–392. https://doi.org/10.1007/s00265-004-0791-8
- 1914 Jones, D. A., Bergman, E., & Greenberg, L. (2015). Food availability in spring affects
- 1915 smolting in brown trout (Salmo trutta). Canadian Journal of Fisheries and Aquatic
- 1916 Sciences, 72(11), 1694–1699. https://doi.org/10.1139/cjfas-2015-0106
- 1917 Jones, J. I., Murphy, J. F., Collins, A. L., Sear, D. A., Naden, P. S., & Armitage, P. D. (2012).
- 1918 The impact of fine sediment on macro-invertebrates. *River Research and*
- 1919 *Applications*, 28(8), 1055–1071.
- 1920 Jonsson, B. (1981). Life History Strategies of Trout (Salmo Trutta L). (PhD Thesis).
- 1921 University of Oslo Oslo, Norway.
- 1922 Jonsson, B. (1982). Diadromous and Resident Trout *Salmo trutta*: Is Their Difference Due to
- 1923 Genetics? *Oikos*, *38*(3), 297–300. https://doi.org/10.2307/3544668
- 1924 Jonsson, B. (1985). Life history patterns of freshwater resident and sea-run migrant brown
- 1925 trout in Norway. Transactions of the American Fisheries Society, 114(2), 182–194.
- 1926 Jonsson, B. (1989). Life history and habitat use of Norwegian brown trout (*Salmo trutta*).
- 1927 Freshwater Biology, 21(1), 71–86. https://doi.org/10.1111/j.1365-
- 1928 2427.1989.tb01349.x
- 1929 Jonsson, B., Jonsson, M., & Jonsson, N. (2017). Influences of migration phenology on
- 1930 survival are size-dependent in juvenile Atlantic salmon (Salmo salar). Canadian
- 1931 *Journal of Zoology*, 95(8), 581–587. https://doi.org/10.1139/cjz-2016-0136

- Jonsson, B., & Jonsson, N. (1993). Partial migration: niche shift versus sexual maturation in
 fishes. *Reviews in Fish Biology and Fisheries*, 3(4), 348–365.
- 1934 https://doi.org/10.1007/BF00043384
- 1935 Jonsson, B., & Jonsson, N. (2005). Lipid energy reserves influence life-history decision of
- 1936 Atlantic salmon (*Salmo salar*) and brown trout (*S. trutta*) in fresh water. *Ecology of*
- 1937 Freshwater Fish, 14(3), 296–301. https://doi.org/10.1111/j.1600-0633.2005.00098.x
- Jonsson, B., & Jonsson, N. (2006a). *Life history of the anadromous trout* Salmo trutta (Vol.
 1939 14). Chapter.
- Jonsson, B., & Jonsson, N. (2006b). Life-history effects of migratory costs in anadromous
 brown trout. *Journal of Fish Biology*, 69(3), 860–869. https://doi.org/10.1111/j.1095-
- 1942 8649.2006.01160.x
- 1943 Jonsson, B., & Jonsson, N. (2009a). A review of the likely effects of climate change on
- 1944 anadromous Atlantic salmon Salmo salar and brown trout Salmo trutta, with
- 1945 particular reference to water temperature and flow. *Journal of Fish Biology*, 75(10),

1946 2381–2447. https://doi.org/10.1111/j.1095-8649.2009.02380.x

- 1947 Jonsson, B., & Jonsson, N. (2009b). Migratory timing, marine survival and growth of
- 1948 anadromous brown trout *Salmo trutta* in the River Imsa, Norway. *Journal of Fish*
- 1949 *Biology*, 74(3), 621–638. https://doi.org/10.1111/j.1095-8649.2008.02152.x
- Jonsson, B., & Jonsson, N. (2011). *Ecology of Atlantic Salmon and Brown Trout. Habitat as a template for life histories*. Dordrecht: Springer Netherlands.
- 1952 Jonsson, B., & Jonsson, N. (2014). Naturally and hatchery produced European trout *Salmo*
- *trutta*: do their marine survival and dispersal differ? *Journal of Coastal Conservation*, *18*(2), 79–87.
- Jonsson, B., & Jonsson, N. (2015). Sexual size dimorphism in anadromous brown trout *Salmo trutta. Journal of Fish Biology*, 87(1), 187–193.

1957	Jonsson, B., & Jonsson, N. (2016). Trans-generational maternal effect: temperature
1958	influences egg size of the offspring in Atlantic salmon Salmo salar. Journal of Fish

1959 *Biology*, 89(2), 1482–1487. https://doi.org/10.1111/jfb.13040

- 1960 Jonsson, B., & Jonsson, N. (2018). Egg incubation temperature affects the timing of the
- 1961 Atlantic salmon *Salmo salar* homing migration. *Journal of Fish Biology*, 93(5), 1016–
- 1962 1020. https://doi.org/10.1111/jfb.13817
- Jonsson, B., & Jonsson, N. (2019). Phenotypic plasticity and epigenetics of fish: embryo
 temperature affects later developing traits. *Aquatic Biology*, 28, 21–32.
- 1965 Jonsson, B., Jonsson, N., & Albretsen, J. (2016). Environmental change influences the life

history of salmon *Salmo salar* in the North Atlantic Ocean: *salmo salar* at sea. *Journal of Fish Biology*, n/a-n/a. https://doi.org/10.1111/jfb.12854

- Jonsson, B., Jonsson, N., Brodtkorb, E., & Ingebrigtsen, P.-J. (2001). Life-history traits of
 Brown Trout vary with the size of small streams. *Functional Ecology*, 15(3), 310–
- 1970
 317.
- 1971 Jonsson, B., Jonsson, N., & Finstad, A. G. (2013). Effects of temperature and food quality on

age and size at maturity in ectotherms: an experimental test with Atlantic salmon.

1973 *Journal of Animal Ecology*, 82(1), 201–210. https://doi.org/10.1111/j.1365-

- 1974 2656.2012.02022.x
- Jonsson, B., Jonsson, N., & Finstad, A. G. (2014). Linking embryonic temperature with adult
 reproductive investment in Atlantic salmon *Salmo salar*. *Marine Ecology Progress Series*, *515*, 217–226.
- 1978 Jonsson, B., Jonsson, N., & Jonsson, M. (2018). Water level influences migratory patterns of
- anadromous brown trout in small streams. *Ecology of Freshwater Fish*.
- 1980 https://doi.org/10.1111/eff.12415

1981	Jonsson, B., Jonsson, N., & Ugedal, O. (2011). Production of juvenile salmonids in small
1982	Norwegian streams is affected by agricultural land use. Freshwater Biology, 56(12),

1983 2529–2542. https://doi.org/10.1111/j.1365-2427.2011.02678.x

- 1984 Jonsson, B., & Ruud-Hansen, J. (1985). Water Temperature as the Primary Influence on
- 1985Timing of Seaward Migrations of Atlantic Salmon (Salmo salar) Smolts. Canadian
- 1986 *Journal of Fisheries and Aquatic Sciences*, 42(3), 593–595.
- 1987 https://doi.org/10.1139/f85-076
- 1988 Jonsson, B., & Sandlund, O. T. (1979). Environmental factors and life histories of isolated
- 1989 river stocks of brown trout (*Salmo trutta* m. *fario*) in Søre Osa river system, Norway.
- 1990 *Environmental Biology of Fishes*, 4(1), 43–54. https://doi.org/10.1007/BF00005927
- 1991 Jonsson, N., & Jonsson, B. (1997a). Energy allocation in polymorphic Brown Trout.
- 1992 Functional Ecology, 11(3), 310–317. https://doi.org/10.1046/j.1365-
- 1993 2435.1997.00083.x
- 1994 Jonsson, N., & Jonsson, B. (1997b). Energy allocation in polymorphic Brown Trout.
- 1995 Functional Ecology, 11(3), 310–317. https://doi.org/10.1046/j.1365-
- 1996 2435.1997.00083.x
- Jonsson, N., & Jonsson, B. (1998). Body composition and energy allocation in life-history
 stages of brown trout. *Journal of Fish Biology*, *53*(6), 1306–1316.
- 1999 https://doi.org/10.1111/j.1095-8649.1998.tb00250.x
- Jonsson, N., & Jonsson, B. (1999). Trade-off between egg mass and egg number in brown
 trout. *Journal of Fish Biology*, 55(4), 767–783.
- Jonsson, N., & Jonsson, B. (2002). Migration of anadromous brown trout *Salmo trutta* in a
 Norwegian river. *Freshwater Biology*, 47(8), 1391–1401.

2004	Jonsson, N., Jonsson, B., & Hansen, L. P. (1994). Sea-ranching of brown trout, Salmo trutta
2005	L. Fisheries Management and Ecology, 1(1), 67–76. https://doi.org/10.1111/j.1365-
2006	2400.1970.tb00007.x
2007	Jonsson, N., Jonsson, B., & Hansen, L. P. (1998). Long-term study of the ecology of wild
2008	Atlantic salmon smolts in a small Norwegian river. Journal of Fish Biology, 52(3),
2009	638–650.
2010	Jonsson, N., Jonsson, B., Hansen, L. P., & Aass, P. (1994). Effects of seawater-
2011	acclimatization and release sites on survival of hatchery-reared brown trout Salmo
2012	trutta. Journal of Fish Biology, 44(6), 973-981. https://doi.org/10.1111/j.1095-
2013	8649.1994.tb01269.x
2014	Kalleberg, H. (1958). Observations in a stream tank of territoriality and competition in
2015	juvenile salmon and trout. Rep. Inst. Freshw. Res. Drottninigholm, 39, 55–98.
2016	Kano, Y., Shimizu, Y., & Kondou, K. (2006). Status-dependent female mimicry in
2017	landlocked red-spotted masu salmon. Journal of Ethology, 24(1), 1-7.
2018	Karlström, Ö. (1977). Habitat selection and population densities of salmon (Salmo salar L.)
2019	and trout (Salmo trutta L.) parr in Swedish rivers with some reference to human
2020	activities. Abstracts of Uppsala Dissertations from the Faculty of Science (Sweden).
2021	No. 404.
2022	Keenleyside, M. H., & Dupuis, H. M. (1988). Courtship and spawning competition in pink
2023	salmon (Oncorhynchus gorbuscha). Canadian Journal of Zoology, 66(1), 262–265.
2024	Keller, I., Taverna, A., & Seehausen, O. (2011). A pyrosequencing assay for the rapid
2025	discrimination of mitochondrial lineages in the Salmo trutta species complex.
2026	Molecular Ecology Resources, 11(1), 196–199. https://doi.org/10.1111/j.1755-
2027	0998.2010.02897.x

2028	Kendall, N. W., McMillan, J. R., Sloat, M. R., Buehrens, T. W., Quinn, T. P., Pess, G. R.,
2029	Zabel, R. W. (2014). Anadromy and residency in steelhead and rainbow trout
2030	(Oncorhynchus mykiss): a review of the processes and patterns. Canadian Journal of
2031	Fisheries and Aquatic Sciences, 72(3), 319–342. https://doi.org/10.1139/cjfas-2014-
2032	0192
2033	Kennedy, G. J. A., & Strange, C. D. (1986). The effects of intra-and inter-specific
2034	competition on the survival and growth of stocked juvenile Atlantic salmon, Salmo
2035	solar L., and resident trout, Salmo trutta L., in an upland stream. Journal of Fish
2036	Biology, 28(4), 479–489.
2037	Klemetsen, A., Amundsen, PA., Dempson, J. B., Jonsson, B., Jonsson, N., O'connell, M. F.,
2038	& Mortensen, E. (2003). Atlantic salmon Salmo salar L., brown trout Salmo trutta L.
2039	and Arctic charr Salvelinus alpinus (L.): a review of aspects of their life histories.
2040	Ecology of Freshwater Fish, 12(1), 1–59.
2041	Knudsen, C. M., Johnston, M. V., Schroder, S. L., Bosch, W. J., Fast, D. E., & Strom, C. R.
2042	(2009). Effects of Passive Integrated Transponder Tags on Smolt-to-Adult Recruit

2043 Survival, Growth, and Behavior of Hatchery Spring Chinook Salmon. *North*

2044 American Journal of Fisheries Management, 29(3), 658–669.

- 2045 https://doi.org/10.1577/M07-020.1
- Knutsen, J. A., Knutsen, H., Gjøsæter, J., & Jonsson, B. (2001). Food of anadromous brown
 trout at sea. *Journal of Fish Biology*, 59(3), 533–543. https://doi.org/10.1111/j.1095-
- 2048 8649.2001.tb02359.x
- Kondolf, G. M., & Wolman, M. G. (1993). The sizes of salmonid spawning gravels. *Water Resources Research*, 29(7), 2275–2285. https://doi.org/10.1029/93WR00402

2051	Kristensen.	M. L.,	Righton.	D., del	Villar-Guerra,	D.	Baktoft, H.	& Aarestrup	. K.	(2018)
1001	i in ibeenigenig		ingineoin,	2., 401	, mai Oaciia,		, D ancort, II.	,	, <u></u> .	(=010)

- 2052 Temperature and depth preferences of adult sea trout *Salmo trutta* during the marine 2053 migration phase. *Marine Ecology Progress Series*, *599*, 209–224.
- 2054 Krkošek, M., Revie, C. W., Gargan, P. G., Skilbrei, O. T., Finstad, B., & Todd, C. D. (2013).
- 2055 Impact of parasites on salmon recruitment in the Northeast Atlantic Ocean. *Proc. R.*

2056 Soc. B, 280(1750), 20122359. https://doi.org/10.1098/rspb.2012.2359

- 2057 Kroglund, F., Finstad, B., Stefansson, S. O., Nilsen, T. O., Kristensen, T., Rosseland, B. O.,
- 2058 ... Salbu, B. (2007). Exposure to moderate acid water and aluminum reduces Atlantic
 2059 salmon post-smolt survival. *Aquaculture*, 273(2), 360–373.
- 2060 https://doi.org/10.1016/j.aquaculture.2007.10.018
- 2061 L'Abee-Lund, J. H., Jonsson, B., Jensen, A. J., Saettem, L. M., Heggberget, T. G., Johnsen,
- 2062 B. O., & Naesje, T. F. (1989). Latitudinal Variation in Life-History Characteristics of
- 2063 Sea-Run Migrant Brown Trout Salmo trutta. The Journal of Animal Ecology, 58(2),
- 2064 525. https://doi.org/10.2307/4846
- Landergren, P. (2001). Survival and growth of sea trout parr in fresh and brackish water.
- 2066 *Journal of Fish Biology*, 58(2), 591–593.
- 2067 Landergren, P. (2004). Factors affecting early migration of sea trout *Salmo trutta* parr to

2068 brackish water. *Fisheries Research*, 67(3), 283–294.

- 2069 https://doi.org/10.1016/j.fishres.2003.10.005
- 2070 Landergren, P. (2005). Survival and growth of sea trout parr in fresh and brackish water.
- 2071 Journal of Fish Biology, 58(2), 591–593. https://doi.org/10.1111/j.1095-
- 2072 8649.2001.tb02275.x
- 2073 Landergren, P., & Vallin, L. (1998). Spawning of sea trout, Salmo trutta L., in brackish
- 2074 waters—lost effort or successful strategy? *Fisheries Research*, *35*(3), 229–236.
- 2075 https://doi.org/10.1016/S0165-7836(98)00073-3

2076	Langeland, A., L'Abée-Lund, J. H., Jonsson, B., & Jonsson, N. (1991). Resource partitioning
2077	and niche shift in Arctic charr Salvelinus alpinus and brown trout Salmo trutta. The
2078	Journal of Animal Ecology, 895–912.

- Larinier, M. (2002). Fish passage through culvets, rock weirs and estuarine obstructions.
 Bulletin Français de La Pêche et de La Pisciculture, (364 supplément), 119–134.
 https://doi.org/10.1051/kmae/2002097
- 2082 Larios-López, J. E., de Figueroa, J. M. T., Galiana-García, M., Gortázar, J., & Alonso, C.
- 2083 (2015). Extended spawning in brown trout (*Salmo trutta*) populations from the
- 2084 Southern Iberian Peninsula: the role of climate variability. *Journal of Limnology*,
 2085 74(2).
- Larsson, S., Serrano, I., & Eriksson, L.-O. (2011). olls. *Canadian Journal of Fisheries and Aquatic Sciences*, 69(1), 1–12. https://doi.org/10.1139/f2011-128
- Lassalle, G., & Rochard, E. (2009). Impact of twenty-first century climate change on
 diadromous fish spread over Europe, North Africa and the Middle East. *Global Change Biology*, *15*(5), 1072–1089.
- 2091 Lauridsen, R. B., Moore, A., Gregory, S. D., Beaumont, W. R. C., Privitera, L., & Kavanagh,
- J. A. (2017). Migration behaviour and loss rate of trout smolts in the transitional zone
 between freshwater and saltwater. In *Sea Trout: Science & Management* (pp. 292–
 307).
- Lecomte, F., Beall, E., Chat, J., Davaine, P., & Gaudin, P. (2013). The complete history of
 salmonid introductions in the Kerguelen Islands, Southern Ocean. *Polar Biology*,
- 2097 36(4), 457–475. https://doi.org/10.1007/s00300-012-1281-5
- 2098 Lemopoulos, A., Uusi-Heikkilä, S., Huusko, A., Vasemägi, A., Vainikka, A., & Yi, S.
- 2099 (2018). Comparison of Migratory and Resident Populations of Brown Trout Reveals

2100 Candidate Genes for Migration Tendency. *Genome Biology and Evolution*, 10(6),

- 2101 1493–1503. https://doi.org/10.1093/gbe/evy102
- 2102 Lemopoulos, A., Uusi-Heikkilä, S., Vasemägi, A., Huusko, A., Kokko, H., & Vainikka, A.
- 2103 (2017). Genome-wide divergence patterns support fine-scaled genetic structuring
- 2104 associated with migration tendency in brown trout. *Canadian Journal of Fisheries*
- 2105 and Aquatic Sciences, 75(10), 1680–1692. https://doi.org/10.1139/cjfas-2017-0014
- 2106 Limburg, K. E., Landergren, P., Westin, L., Elfman, M., & Kristiansson, P. (2001). Flexible
- 2107 modes of anadromy in Baltic sea trout: making the most of marginal spawning
- 2108 streams. Journal of Fish Biology, 59(3), 682–695. https://doi.org/10.1111/j.1095-
- 2109 8649.2001.tb02372.x
- 2110 Linnaeus, C. (1758). Systema naturae, Vol. 1. Systema naturae, Vol. 1.
- 2111 Lobón-Cerviá, J. (2007). Numerical changes in stream-resident brown trout (Salmo trutta):
- 2112 uncovering the roles of density-dependent and density-independent factors across
- 2113 space and time. Canadian Journal of Fisheries and Aquatic Sciences, 64(10), 1429–
- 2114 1447. https://doi.org/10.1139/f07-111
- 2115 Lobón-Cerviá, J., Rasmussen, G. H., & Mortensen, E. (2017). Discharge-Dependent
- 2116 Recruitment in Stream-Spawning Brown Trout. In J. Lobón-Cerviá & N. Sanz (Eds.),
- 2117 Brown Trout (pp. 297–318). https://doi.org/10.1002/9781119268352.ch13
- Lobón-Cerviá, J., & Sanz, N. (2017). *Brown Trout: Biology, Ecology and Management*. John
 Wiley & Sons.
- 2120 Maisse, G., & Baglinière, J.-L. (1990). The biology of brown trout, Salmo trutta L., in the
- 2121 River Scorff, Brittany: a synthesis of studies from 1973 to 1984. Aquaculture
- 2122 *Research*, 21(1), 95–106. https://doi.org/10.1111/j.1365-2109.1990.tb00385.x

Maisse, G., & Baglinière, J.-L. (1999). Biology of the brown trout (Salmo trutta L.) in French

- rivers. In J.-L. Baglinière & G. Maisse (Eds.), *Biology and Ecology of the Brown and Sea Trout* (pp. 15–35). https://doi.org/10.1007/978-1-4471-0775-0_2
- 2126 Maisse, G., Mourot, B., Breton, B., Fostier, A., Marcuzzi, O., Le Bail, P. Y., ... Richard, A.
- 2127 (1991). Sexual maturity in sea trout, Salmo trutta L., running up the River Calonne
- 2128 (Normandy, France) at the 'finnock' stage. *Journal of Fish Biology*, 39(5), 705–715.
- Mäki-Petäys, A., Vehanen, T., & Muotka, T. (2000). Microhabitat use by age-0 brown trout
 and grayling: seasonal responses to streambed restoration under different flows.
- 2131 Transactions of the American Fisheries Society, 129(3), 771–781.
- 2132 Marco-Rius, F., Caballero, P., Moran, P., & Leaniz, C. G. de. (2012). And the Last Shall Be
- 2133 First: Heterochrony and Compensatory Marine Growth in Sea Trout (*Salmo trutta*).

2134 PLOS ONE, 7(10), e45528. https://doi.org/10.1371/journal.pone.0045528

- Marco-Rius, F., Caballero, P., Moran, P., & Leaniz, C. G. de. (2013). Can migrants escape
 from density dependence? *Ecology and Evolution*, *3*(8), 2524–2534.
- 2137 https://doi.org/10.1002/ece3.652
- 2138 Massa, F., Baglinière, J.-L., Prunet, P., & Grimaldi, C. (2000). Survie embryo-larvaire de la
- truite (*Salmo trutta*) et conditions chimiques dans la frayère | French Ichthyological
 Society Cybium. *Cybium*, 24(3), 129–140.
- 2141 McCormick, S. D., Hansen, L. P., Quinn, T. P., & Saunders, R. L. (1998). Movement,
- 2142 migration, and smolting of Atlantic salmon (*Salmo salar*). *Canadian Journal of*
- 2143 *Fisheries and Aquatic Sciences*, 55(S1), 77–92. https://doi.org/10.1139/d98-011
- McCormick, S. D., Shrimpton, J. M., & Zydlewski, J. D. (1996). *Temperature effects on osmoregulatory physiology of juvenile anadromous fish.*
- 2146 McKeown, N. J., Hynes, R. A., Duguid, R. A., Ferguson, A., & Prodöhl, P. A. (2010).
- 2147 Phylogeographic structure of brown trout *Salmo trutta* in Britain and Ireland: glacial

- 2148 refugia, postglacial colonization and origins of sympatric populations. *Journal of Fish*2149 *Biology*, 76(2), 319–347. https://doi.org/10.1111/j.1095-8649.2009.02490.x
- 2150 McLean, J. E., Bentzen, P., & Quinn, T. P. (2004). Does Size Matter? Fitness-Related Factors
- 2151 in Steelhead Trout Determined by Genetic Parentage Assignment. *Ecology*, 85(11),
- 2152 2979–2985. https://doi.org/10.1890/03-0488
- 2153 McMillan, J. R., Katz, S. L., & Pess, G. R. (2007). Observational evidence of spatial and
- 2154 temporal structure in a sympatric anadromous (winter steelhead) and resident rainbow
- 2155 trout mating system on the Olympic Peninsula, Washington. *Transactions of the*
- 2156 American Fisheries Society, 136(3), 736–748.
- McNeil, W. J. (1964). Redd superimposition and egg capacity of pink salmon spawning beds. *Journal of the Fisheries Board of Canada*, *21*(6), 1385–1396.
- 2159 Meier, K., Hansen, M. M., Bekkevold, D., Skaala, Ø., & Mensberg, K. D. (2011). An

assessment of the spatial scale of local adaptation in brown trout (Salmo trutta L.):

2161 footprints of selection at microsatellite DNA loci. *Heredity*, *106*(3), 488.

2162 Meland, S., Borgstrøm, R., Heier, L. S., Rosseland, B. O., Lindholm, O., & Salbu, B. (2010).

2163 Chemical and ecological effects of contaminated tunnel wash water runoff to a small

- 2164 Norwegian stream. *Science of The Total Environment*, 408(19), 4107–4117.
- 2165 https://doi.org/10.1016/j.scitotenv.2010.05.034
- Mills, C. P. R., Mahon, G. A. T., & Piggins, D. J. (1986). Influence of stock levels, fishing
 effort and environmental factors on anglers' catches of Atlantic salmon, *Salmo salar*
- L., and sea trout, *Salmo trutta* L. *Aquaculture Research*, 17(4), 289–297.
- Mills, C. P. R., Piggins, D. J., & Cross, T. F. (1990). Burrishoole sea trout-a twenty year
 study. 20th Annual Study Course Proceedings, 61–78.
- 2171 Milner, N. J., Harris, G., Gargan, P., Beveridge, M., Pawson, M. G., Walker, A., & Whelan,
- 2172 K. F. (2007). Perspectives on Sea Trout Science and Management. In *Sea Trout:*

- 2173 *Biology, Conservation and Management* (pp. 480–490).
- 2174 https://doi.org/10.1002/9780470996027.ch34
- 2175 Montgomery, D. R., Buffington, J. M., Peterson, N. P., Schuett-Hames, D., & Quinn, T. P.
- 2176 (1996). Stream-bed scour, egg burial depths, and the influence of salmonid spawning
 2177 on bed surface mobility and embryo survival. 53, 10.
- 2178 Montorio, L., Evanno, G., & Nevoux, M. (2018). Intra- and interspecific densities shape life-
- history traits in a salmonid population. *Oecologia*, 188(2), 451–464.
- 2180 https://doi.org/10.1007/s00442-018-4213-4
- 2181 Moore, A., Lower, N., Mayer, I., & Greenwood, L. (2007). The impact of a pesticide on
- 2182 migratory activity and olfactory function in Atlantic salmon (*Salmo salar* L.) smolts.

2183 Aquaculture, 273(2), 350–359. https://doi.org/10.1016/j.aquaculture.2007.10.017

- 2184 Moore, I., Dodd, J. A., Newton, M., Bean, C. W., Lindsay, I., Jarosz, P., & Adams, C. E.
- 2185 (2018). The influence of aquaculture unit proximity on the pattern of *Lepeophtheirus*
- 2186 *salmonis* infection of anadromous *Salmo trutta* populations on the isle of Skye,
- 2187 Scotland. Journal of Fish Biology, 92(6), 1849–1865.
- 2188 https://doi.org/10.1111/jfb.13625
- 2189 Moran, P., Marco-Rius, F., Megías, M., Covelo-Soto, L., & Pérez-Figueroa, A. (2013).
- 2190 Environmental induced methylation changes associated with seawater adaptation in
- brown trout. *Aquaculture*, *392–395*(Supplement C), 77–83.
- 2192 https://doi.org/10.1016/j.aquaculture.2013.02.006
- 2193 Moran, P., & Pérez-Figueroa, A. (2011). Methylation changes associated with early
- 2194 maturation stages in the Atlantic salmon. *BMC Genetics*, *12*(1), 86.
- 2195 https://doi.org/10.1186/1471-2156-12-86
- 2196 Morinville, G. R., & Rasmussen, J. B. (2003). Early juvenile bioenergetic differences
- 2197 between anadromous and resident brook trout (*Salvelinus fontinalis*). *Canadian*

- *Journal of Fisheries and Aquatic Sciences*, 60(4), 401–410.
- 2199 https://doi.org/10.1139/f03-036
- 2200 Morita, K., Fukuwaka, M., Tanimata, N., & Yamamura, O. (2010). Size-dependent thermal
- 2201 preferences in a pelagic fish. *Oikos*, *119*(8), 1265–1272.
- 2202 https://doi.org/10.1111/j.1600-0706.2009.18125.x
- 2203 Morita, K., Tamate, T., Kuroki, M., & Nagasawa, T. (2014). Temperature-dependent
- 2204 variation in alternative migratory tactics and its implications for fitness and
- 2205 population dynamics in a salmonid fish. Journal of Animal Ecology, 83(6), 1268–
- 2206 1278. https://doi.org/10.1111/1365-2656.12240
- 2207 Myksvoll, M. S., Sandvik, A. D., Albretsen, J., Asplin, L., Johnsen, I. A., Karlsen, Ø., ...
- 2208 Ådlandsvik, B. (2018). Evaluation of a national operational salmon lice monitoring
- system—From physics to fish. *PLOS ONE*, *13*(7), e0201338.
- 2210 https://doi.org/10.1371/journal.pone.0201338
- 2211 Näslund, J., Sundström, L. F., & Johnsson, J. I. (2015). Autumn food restriction reduces
- smoltification rate, but not over-winter survival, in juvenile brown trout *Salmo trutta*.

2213 Ecology of Freshwater Fish, 26(2), 205–216. https://doi.org/10.1111/eff.12268

- 2214 Nielsen, C., Aarestrup, K., & Madsen, S. S. (2006). Comparison of physiological smolt status
- in descending and nondescending wild brown trout (*Salmo trutta*) in a Danish stream.
- 2216 Ecology of Freshwater Fish, 15(2), 229–236. https://doi.org/10.1111/j.1600-
- 2217 0633.2006.00143.x
- 2218 Nielsen, J. (1994). Vandlöbsfiskenes verden med biologen på arbejde. Copenhaguen,
 2219 Denmark: Gad.
- Nika, N., Virbickas, T., & Kontautas, A. (2011). Spawning site selection and redd gravel
 characteristics of sea trout Salmo trutta in the lowland streams of Lithuania.

- 2222 *Oceanological and Hydrobiological Studies*, 40(1), 46–56.
- 2223 https://doi.org/10.2478/s13545-011-0006-7
- Nomoto, K., Omiya, H., Sugimoto, T., Akiba, K., Edo, K., & Higashi, S. (2010). Potential
 negative impacts of introduced rainbow trout on endangered Sakhalin taimen through
- redd disturbance in an agricultural stream, eastern Hokkaido. *Ecology of Freshwater*
- 2227 Fish, 19(1), 116–126.
- Nordeng, H. (1983). Solution to the" char problem" based on Arctic char (*Salvelinus alpinus*)
 in Norway. *Canadian Journal of Fisheries and Aquatic Sciences*, 40(9), 1372–1387.
- 2230 Northcote, T. G. (1992). Migration and residency in stream salmonids- some ecological
- 2231 considerations and evolutionary consequences. *Nordic Journal of Freshwater* 2232 *Research. Drottningholm*, 67, 5–17.
- Økland, F., Jonsson, B., Jensen, A. J., & Hansen, L. P. (1993). Is there a threshold size
 regulating seaward migration of brown trout and Atlantic salmon? *Journal of Fish*

2235 Biology, 42(4), 541–550. https://doi.org/10.1111/j.1095-8649.1993.tb00358.x

- Okumuş, I., Kurtoglu, I. Z., & Atasaral, Ş. (2006). General Overview of Turkish Sea Trout
 (Salmo trutta L.) Populations. Sea Trout: Biology, Conservation and Management,
 115–127.
- Olsen, E. M., Knutsen, H., Simonsen, J. H., Jonsson, B., & Knutsen, J. A. (2006). Seasonal
 variation in marine growth of sea trout, *Salmo trutta*, in coastal Skagerrak. *Ecology of Freshwater Fish*, 15(4), 446–452.
- 2242 Olsén, K. H., Järvi, J. T., Mayer, I., Petersson, E., & Kroon, F. (1998). Spawning behaviour
- and sex hormone levels in adult and precocious brown trout (*Salmo trutta* L.) males
 and the effect of anosmia. *Chemoecology*, 8(1), 9–17.
- 2245 https://doi.org/10.1007/PL00001803

2246	Olsson, I. C., Greenberg, L. A., Bergman, E., & Wysujack, K. (2006). Environmentally
2247	induced migration: the importance of food. Ecology Letters, 9(6), 645-651.
2248	https://doi.org/10.1111/j.1461-0248.2006.00909.x
2249	Ombredane, D., Siegler, L., Baglinière, JL., & Prunet, P. (1996). Migration et smoltification
2250	des juvéniles de truite (Salmo trutta) dans deux cours d'eau de Basse-Normandie.
2251	<i>Cybium</i> , 20(3), 27–42.
2252	Otero, J., L'Abée-Lund, J. H., Castro-Santos, T., Leonardsson, K., Storvik, G. O., Jonsson,
2253	B., Vøllestad, L. A. (2014). Basin-scale phenology and effects of climate
2254	variability on global timing of initial seaward migration of Atlantic salmon (Salmo
2255	salar). Global Change Biology, 20(1), 61–75. https://doi.org/10.1111/gcb.12363
2256	Patarnello, T., Bargelloni, L., Caldara, F., & Colombo, L. (1994). Cytochrome b and 16S
2257	rRNA Sequence Variation in the Salmo trutta (Salmonidae, Teleostei) Species
2258	Complex. Molecular Phylogenetics and Evolution, 3(1), 69–74.
2259	https://doi.org/10.1006/mpev.1994.1008
2260	Peiman, K. S., Birnie Gauvin, K., Midwood, J. D., Larsen, M. H., Wilson, A. D. M.,
2261	Aarestrup, K., & Cooke, S. J. (2017). If and when: intrinsic differences and
2262	environmental stressors influence migration in brown trout (Salmo trutta). Oecologia,
2263	184(2), 375–384. https://doi.org/10.1007/s00442-017-3873-9
2264	Peterson, D. P., Rieman, B. E., Young, M. K., & Brammer, J. A. (2010). Modeling predicts
2265	that redd trampling by cattle may contribute to population declines of native trout.
2266	Ecological Applications, 20(4), 954–966. https://doi.org/10.1890/09-0679.1
2267	Pettersson, J. C. E., Hansen, M. M., & Bohlin, T. (2001). Does dispersal from landlocked
2268	trout explain the coexistence of resident and migratory trout females in a small
2269	stream? Journal of Fish Biology, 58(2), 487-495. https://doi.org/10.1111/j.1095-
2270	8649.2001.tb02267.x

2271	Poole, W. R., Byrne, C. J., Dillane, M. G., Whelan, K. F., & Gargan, P. G. (2002). The Irish
2272	sea trout enhancement programme: a review of the broodstock and ova production
2273	programmes. Fisheries Management and Ecology, 9(6), 315–328.

- 2274 Poole, W. R., Dillane, M., DeEyto, E., Rogan, G., McGinnity, P., & Whelan, K. F. (2006).
- 2275 Characteristics of the Burrishoole Sea Trout Population: Census, Marine Survival,
- 2276 Enhancement and Stock-Recruitment Relationship, 1971-2003. In Sea Trout: Biology,
- 2277 *Conservation and Management* (pp. 279–306).
- 2278 https://doi.org/10.1002/9780470996027.ch19
- 2279 Poole, W. R., & Dillane, M. G. (1998). Estimation of sperm concentration of wild and
- reconditioned brown trout, *Salmo trutta* L. *Aquaculture Research*, 29(6), 439–445.
- 2281 https://doi.org/10.1046/j.1365-2109.1998.00223.x
- Poole, W. R., Nolan, D., & Tully, O. (2000). Extrapolating baseline blood cortisol levels in
 trout: modelling capture effects in wild sea trout *Salmo trutta* (L.) infested with *Lepeophtheirus salmonis* (Krøyer). *Aquaculture Research*, *31*(11), 835–841.
- 2285 Poole, W. R., Whelan, K. F., Dillane, M. G., Cooke, D. J., & Matthews, M. (1996). The
- 2286 performance of sea trout, *Salmo trutta* L., stocks from the Burrishoole system western

Ireland, 1970–1994. *Fisheries Management and Ecology*, 3(1), 73–92.

- Potter, E., Campbell, R. N., Sumner, K., & Marshall, S. (2017). *Marine migrations and distribution of sea trout from rivers in Great Britain.*
- 2290 Prodöhl, P., Antoniacomi, A., Bradley, C., Carlson, J., Carvalho, G. R., Coughlan, J., ...
- 2291 Cross, T. F. (2017). Population genetics and Genetic Stock Identification of
- anadromous Salmo trutta from the Irish Sea and adjacent areas, using microsatellite
- 2293 DNA loci. In Sea Trout: Science and Management : Proceedings of the 2nd
- 2294 *International Sea Trout Symposium* (pp. 69–95). Retrieved from
- 2295 https://pure.qub.ac.uk/portal/en/publications/population-genetics-and-genetic-stock-

2296 identification-of-anadromous-salmo-trutta-from-the-irish-sea-and-adjacent-areas-2297 using-microsatellite-dna-loci(86dedf32-1d65-4de0-80e6-4d0583fb8c85)/export.html Prouzet, P., Le Bail, P. Y., & Heydorff, M. (1984). Sex Ratio and Potential Fecundity of 2298 2299 Atlantic Salmon (Salmo salar L.) Caught by Anglers on the Elorn River (Northern Brittany, France) during 1979 and 1980. Aquaculture Research, 15(3), 123–130. 2300 2301 https://doi.org/10.1111/j.1365-2109.1984.tb00843.x 2302 Pulg, U., Barlaup, B. T., Sternecker, K., Trepl, L., & Unfer, G. (2013). Restoration of 2303 spawning habitats of brown trout (Salmo trutta) in a regulated chalk stream. River 2304 Research and Applications, 29(2), 172–182.

- 2305 Quéméré, E., Baglinière, J.-L., Roussel, J.-M., Evanno, G., McGinnity, P., & Launey, S.
- (2016). Seascape and its effect on migratory life-history strategy influences gene flow
 among coastal brown trout (*Salmo trutta*) populations in the English Channel. *Journal of Biogeography*, *43*(3), 498–509. https://doi.org/10.1111/jbi.12632
- 2309 Quéméré, E., le Gentil, J., & Launey, S. (2011). Analyse spatiale et temporelle des
- 2310 *caractéristiques génétiques de la population de truite de mer de la Bresle* (Fiche
- 2311 Action Onema No. Rapport final; p. 124p).
- 2312 Regan, C. T. (1911). The freshwater fishes of the British Isles. London, Methuen & Co. Ltd.
- 2313 Richardson, J. L., Urban, M. C., Bolnick, D. I., & Skelly, D. K. (2014). Microgeographic
- adaptation and the spatial scale of evolution. *Trends in Ecology & Evolution*, 29(3),
- 2315 165–176. https://doi.org/10.1016/j.tree.2014.01.002
- 2316 Riley, W. D., Bendall, B., Ives, M. J., Edmonds, N. J., & Maxwell, D. L. (2012). Street
- 2317 lighting disrupts the diel migratory pattern of wild Atlantic salmon, Salmo salar L.,
- smolts leaving their natal stream. *Aquaculture*, *330–333*, 74–81.
- 2319 https://doi.org/10.1016/j.aquaculture.2011.12.009

2320	Rivot, E., Perrier, C., Jouanin, C., Roussel, JM., Riera, P., Porcher, J. P., & Baglinière, JL.
2321	(2009). Climate change may have affected growth and life history in Atlantic Salmon
2322	Juveniles over the past 30 years. American Fisheries Society Symposium, 69, 855-
2323	857.
2324	Roberts, R. J. (1993). Ulcerative dermal necrosis (UDN) in wild salmonids. Fisheries
2325	Research, 17(1), 3-14. https://doi.org/10.1016/0165-7836(93)90003-P
2326	Rohtla, M., Matetski, L., Svirgsden, R., Kesler, M., Taal, I., Saura, A., Vetemaa, M.
2327	(2017). Do sea trout Salmo trutta parr surveys monitor the densities of anadromous or
2328	resident maternal origin parr, or both? Fisheries Management and Ecology, 24(2),
2329	156–162. https://doi.org/10.1111/fme.12214
2330	Rosseland, B. O., & Kroglund, F. (2010). Lessons from Acidification and Pesticides. In
2331	Atlantic Salmon Ecology (pp. 387-407). https://doi.org/10.1002/9781444327755.ch15
2332	Rounsefell, G. A. (1958). Anadromy in North American Salmonidae. US Fish and Wildlife
2333	Service Fisheries Bulletin, 58, 171–185.
2334	Roussel, JM. (2007). Carry-over effects in brown trout (Salmo trutta): hypoxia on embryos
2335	impairs predator avoidance by alevins in experimental channels. Canadian Journal of
2336	Fisheries and Aquatic Sciences, 64(5), 786–792. https://doi.org/10.1139/f07-055
2337	Roussel, JM., Perrier, C., Erkinaro, J., Niemelä, E., Cunjak, R. A., Huteau, D., & Riera, P.
2338	(2013). Stable isotope analyses on archived fish scales reveal the long-term effect of
2339	nitrogen loads on carbon cycling in rivers. Global Change Biology, 20(2), 523-530.
2340	https://doi.org/10.1111/gcb.12293
2341	Rubin, JF., Glimsäter, C., & Jarvi, T. (2005). Spawning characteristics of the anadromous
2342	brown trout in a small Swedish stream. Journal of Fish Biology, 66(1), 107–121.

2343	Rundio, D. E., Williams, T. H., Pearse, D. E., & Lindley, S. T. (2012). Male-biased sex ratio
2344	of nonanadromous Oncorhynchus mykiss in a partially migratory population in
2345	California. Ecology of Freshwater Fish, 21(2), 293–299.

- 2346 Saltveit, S. J., Halleraker, J. H., Arnekleiv, J. V., & Harby, A. (2001). Field experiments on
- stranding in juvenile Atlantic salmon (*Salmo salar*) and brown trout (*Salmo trutta*)
- 2348 during rapid flow decreases caused by hydropeaking. *Regulated Rivers: Research &*
- 2349 Management: An International Journal Devoted to River Research and Management,

2350 17(4–5), 609–622.

- 2351 Sánchez-Hernández, J., Eloranta, A. P., Finstad, A. G., & Amundsen, P.-A. (2017).
- Community structure affects trophic ontogeny in a predatory fish. *Ecology and Evolution*, 7(1), 358–367. https://doi.org/10.1002/ece3.2600
- Sandlund, O. T., & Jonsson, B. (2016). Life history plasticity: migration ceased in response
 to environmental change? *Ecology of Freshwater Fish*, 25(2), 225–233.
- 2356 Sanz, N. (2017). Phylogeographic History of Brown Trout. In Brown Trout (pp. 15–63).

2357 https://doi.org/10.1002/9781119268352.ch2

- 2358 Scheurer, K., Alewell, C., Bänninger, D., & Burkhardt-Holm, P. (2009). Climate and land-
- use changes affecting river sediment and brown trout in alpine countries—a review.

2360 Environmental Science and Pollution Research, 16(2), 232–242.

- 2361 https://doi.org/10.1007/s11356-008-0075-3
- 2362 Schram, T. A., Knutsen, J. A., Heuch, P. A., & Mo, T. A. (1998). Seasonal occurrence of
- 2363 *Lepeophtheirus salmonis* and *Caligus elongatus* (Copepoda: Caligidae) on sea trout
- 2364 (Salmo trutta), off southern Norway. ICES Journal of Marine Science, 55(2), 163–
- 2365 175. https://doi.org/10.1006/jmsc.1997.0357
- Scott, G. R., & Johnston, I. A. (2012). Temperature during embryonic development has
 persistent effects on thermal acclimation capacity in zebrafish. *Proceedings of the*

- 2368 *National Academy of Sciences*, *109*(35), 14247–14252.
- 2369 https://doi.org/10.1073/pnas.1205012109
- 2370 Segers, F. H., & Taborsky, B. (2011). Egg size and food abundance interactively affect
 2371 juvenile growth and behaviour. *Functional Ecology*, 25(1), 166–176.
- 2372 Serbezov, D., Jorde, P. E., Bernatchez, L., Olsen, E. M., & Vøllestad, L. A. (2012). Life
- history and demographic determinants of effective/census size ratios as exemplified
- by brown trout (*Salmo trutta*). *Evolutionary Applications*, 5(6), 607–618.
- 2375 https://doi.org/10.1111/j.1752-4571.2012.00239.x
- 2376 Serra-Llinares, R. M., Bjørn, P. A., Finstad, B., Nilsen, R., & Asplin, L. (2016). Nearby
- farms are a source of lice for wild salmonids: a reply to Jansen et al.(2016).

2378 *Aquaculture Environment Interactions*, 8, 351–356.

- 2379 Serra-Llinares, R. M., Bjørn, P. A., Finstad, B., Nilsen, R., Harbitz, A., Berg, M., & Asplin,
- L. (2014). Salmon lice infection on wild salmonids in marine protected areas: an
 evaluation of the Norwegian 'National Salmon Fjords.
- 2382 Shephard, S, MacIntyre, C., & Gargan, P. (2016). Aquaculture and environmental drivers of
- 2383 salmon lice infestation and body condition in sea trout. Aquaculture Environment

2384 *Interactions*, 8, 597–610. https://doi.org/10.3354/aei00201

- Shephard, Samuel, Davidson, I. C., Walker, A. M., & Gargan, P. G. (2018). Length-based
 indicators and reference points for assessing data-poor stocks of diadromous trout
- 2387 Salmo trutta. Fisheries Research, 199, 36–43.
- 2388 https://doi.org/10.1016/j.fishres.2017.11.024
- 2389 Sigurjónsdóttir, H., & Gunnarsson, K. (1989). Alternative mating tactics of arctic charr,
- 2390 Salvelinus alpinus, in Thingvallavatn, Iceland. Environmental Biology of Fishes,
- 2391 26(3), 159–176.

2392	Skilbrei, (O. T.,	Finstad,	В.,	Urdal,	К.,	Bakke,	G.,	Kroglund,	F., 6	& S	trand,	R.	(2013).	Impact

- 2393 of early salmon louse, *Lepeophtheirus salmonis*, infestation and differences in
- survival and marine growth of sea-ranched Atlantic salmon, *Salmo salar* L., smolts
- 2395 1997–2009. Journal of Fish Diseases, 36(3), 249–260.
- 2396 https://doi.org/10.1111/jfd.12052
- 2397 Skrochowska, S. (1969). Migration of sea trout (*Salmo trutta* L.), brown trout (*Salmo trutta* 2398 *m. fario* L.) and their crosses. *Polish Archiv Hydrobiologie*, *16*, 125–192.
- 2399 Solomon, D. J. (1997). Review of sea trout fecundity (p. 22pp) [Technical report for the
- 2400 Welsh Environment Agency]. Environment Agency.
- 2401 Solomon, D. J., & Czerwinski, M. (2007). Catch and Release, Net Fishing and Sea Trout
- 2402 Fisheries Management. In Sea Trout: Biology, Conservation and Management (pp.
- 2403 434–440). https://doi.org/10.1002/9780470996027.ch30
- 2404 Stradmeyer, L., Höjesjö, J., Griffiths, S. W., Gilvear, D. J., & Armstrong, J. D. (2008).
- 2405 Competition between brown trout and Atlantic salmon parr over pool refuges during 2406 rapid dewatering. *Journal of Fish Biology*, 72(4), 848–860.
- 2407 Summers, D. W., Giles, N., & Stubbing, D. N. (2008). Rehabilitation of brown trout, Salmo
- 2408 *trutta*, habitat damaged by riparian grazing in an English chalkstream. *Fisheries*
- 2409 *Management and Ecology*, 15(3), 231–240. https://doi.org/10.1111/j.1365-
- 2410 2400.2008.00604.x
- 2411 Summers, D. W., Giles, N., & Willis, D. J. (1996). Restoration of Riverine Trout Habitats -
- 2412 A Guidance Manual (R&D Technical Report W18 No. Fisheries Technical Manual 1;
- 2413 p. 230p). Bristol: Environmental Agency.
- 2414 Svärdson, G. (1966). Öringen. Fiske, (66), 1–31.
- 2415 Taal, I., Kesler, M., Saks, L., Rohtla, M., Verliin, A., Svirgsden, R., ... Saat, T. (2014).
- 2416 Evidence for an autumn downstream migration of Atlantic salmon *Salmo salar*

2417	(Linnaeus) and brown trout Salmo trutta (Linnaeus) parr to the Baltic Sea. Helgoland
2418	Marine Research, 68(2), 373-377. https://doi.org/10.1007/s10152-014-0395-2
2419	Taal, I., Rohtla, M., Saks, L., Kesler, M., Jürgens, K., Svirgsden, R., Vetemaa, M. (2018).
2420	Parr dispersal between streams via a marine environment: A novel mechanism behind
2421	straying for anadromous brown trout? Ecology of Freshwater Fish, 27(1), 209–215.
2422	https://doi.org/10.1111/eff.12338
2423	Tamate, T., & Maekawa, K. (2004). Female-biased mortality rate and sexual size dimorphism
2424	of migratory masu salmon, Oncorhynchus masou. Ecology of Freshwater Fish, 13(2),
2425	96–103.
2426	Tanguy, J. M., Ombredane, D., Baglinière, JL., & Prunet, P. (1994). Aspects of parr-smolt
2427	transformation in anadromous and resident forms of brown trout (Salmo trutta) in
2428	comparison with Atlantic salmon (Salmo salar). Aquaculture, 121(1), 51-63.
2429	https://doi.org/10.1016/0044-8486(94)90007-8
2430	Tappel, P. D., & Bjornn, T. C. (1983). A New Method of Relating Size of Spawning Gravel
2431	to Salmonid Embryo Survival. North American Journal of Fisheries Management,
2432	3(2), 123–135. https://doi.org/10.1577/1548-8659(1983)3<123:ANMORS>2.0.CO;2
2433	Taranger, G. L., Karlsen, Ø., Bannister, R. J., Glover, K. A., Husa, V., Karlsbakk, E.,
2434	Svåsand, T. (2015). Risk assessment of the environmental impact of Norwegian
2435	Atlantic salmon farming. ICES Journal of Marine Science, 72(3), 997–1021.
2436	https://doi.org/10.1093/icesjms/fsu132
2437	Thomsen, D. S., Koed, A., Nielsen, C., & Madsen, S. S. (2007). Overwintering of sea trout
2438	(Salmo trutta) in freshwater: escaping salt and low temperature or an alternate life
2439	strategy? Canadian Journal of Fisheries and Aquatic Sciences, 64(5), 793-802.
2440	https://doi.org/10.1139/f07-059

- Thorpe, J. E., Talbot, C., & Villarreal, C. (1982). Bimodality of growth and smolting in
 Atlantic salmon, Salmo salar L. *Aquaculture*, 28(1–2), 123–132.
- 2443 https://doi.org/10.1016/0044-8486(82)90015-1
- Thorstad, E. B., & Finstad, B. (2018). *Impacts of salmon lice emanating from salmon farms*on wild Atlantic salmon and sea trout (NINA Report No. 1449; p. 22p). NINA.
- 2446 Thorstad, E. B., Todd, C. D., Bjørn, P. A., Gargan, P. G., Vollset, K. W., Halttunen, E., ...
- Finstad, B. (2014). *Effects of salmon lice on sea trout. A litterature review* (NINA
 Report No. 1044; p. 162p).
- 2449 Thorstad, E. B., Todd, C. D., Uglem, I., Bjørn, P. A., Gargan, P. G., Vollset, K. W., ...
- Finstad, B. (2016). Marine life of the sea trout. *Marine Biology*, *163*(3), 47.
- 2451 Thorstad, E. B., Todd, C., Uglem, I., Bjørn, P., Gargan, P., Vollset, K., ... Finstad, B. (2015).
- 2452 Effects of salmon lice Lepeophtheirus salmonis on wild sea trout *Salmo trutta*—a
- 2453 literature review. *Aquaculture Environment Interactions*, 7(2), 91–113.
- 2454 https://doi.org/10.3354/aei00142
- 2455 Titus, R. G., & Mosegaard, H. (1992). Fluctuating recruitment and variable life history of
- 2456 migratory brown trout, Salmo trutta L., in a small, unstable stream. Journal of Fish
- 2457 *Biology*, *41*(2), 239–255. https://doi.org/10.1111/j.1095-8649.1992.tb02654.x
- Tomkins, J. L., & Hazel, W. (2007). The status of the conditional evolutionarily stable
 strategy. *Trends in Ecology & Evolution*, 22(10), 522–528.
- Tully, O. (1992). Predicting infestation parameters and impacts of caligid copepods in wild
 and cultured fish populations. *Invertebrate Reproduction & Development*, 22(1–3),
 91–102.
- Tully, O., Gargan, P., Poole, W. R., & Whelan, K. F. (1999). Spatial and temporal variation
 in the infestation of sea trout (*Salmo trutta* L.) by the caligid copepod *Lepeophtheirus*

2465	salmonis (Krøyer) in relation to sources of infection in Ireland. Parasitology, 119(1),
2466	41–51.

- Tully, O., Poole, W. R., & Whelan, K. F. (1993). Infestation parameters for *Lepeophtheirus salmonis* (Kroyer)(Copepoda: Caligidae) parasitic on sea trout, *Salmo trutta* L., off
 the west coast of Ireland during 1990 and 1991. *Aquaculture Research*, 24(4), 545–
 555.
- Tully, O., Poole, W. R., Whelan, K. F., & Merigoux, S. (1993). Parameters and possible
 causes of epizootics of *Lepeophtheirus salmonis* (Krøyer) infesting sea trout (*Salmo trutta* L.) off the west coast of Ireland. *Pathogens of Wild and Farmed Fish: Sea Lice*,
 202–213.
- 2475 Tully, O., & Whelan, K. F. (1993). Production of nauplii of Lepeophtheirus salmonis
- (Krøyer)(Copepoda: Caligidae) from farmed and wild salmon and its relation to the
 infestation of wild sea trout (*Salmo trutta* L.) off the west coast of Ireland in 1991.
- 2478 Fisheries Research, 17(1–2), 187–200.
- 2479 Van Doornik, D. M., Berejikian, B. A., & Campbell, L. A. (2013). Gene flow between
- 2480 sympatric life history forms of *Oncorhynchus mykiss* located above and below
 2481 migratory barriers. *PloS One*, 8(11), e79931.
- 2482 Vøllestad, L. A. (2017). Understanding Brown Trout Population Genetic Structure. In J.
- 2483 Lobón-Cerviá & N. Sanz (Eds.), Brown Trout (pp. G1–G72).
- 2484 https://doi.org/10.1002/9781119268352.app
- 2485 Vollset, K. W., Dohoo, I., Karlsen, Ø., Halttunen, E., Kvamme, B. O., Finstad, B., ...
- 2486 Barlaup, B. T. (2018). Disentangling the role of sea lice on the marine survival of
- 2487 Atlantic salmon. *ICES Journal of Marine Science*, 75(1), 50–60.
- 2488 https://doi.org/10.1093/icesjms/fsx104

2489	Vollset, K. W., Krontveit, R. I., Jansen, P. A., Finstad, B., Barlaup, B. T., Skilbrei, O. T.,
2490	Dohoo, I. (2016). Impacts of parasites on marine survival of Atlantic salmon: a meta-
2491	analysis. Fish and Fisheries, 17(3), 714-730. https://doi.org/10.1111/faf.12141
2492	Walker, A. M., & Bayliss, B. D. (2007). The Spawning Habitat Requirements of Sea Trout:
2493	A Multi-Scale Approach. In Sea Trout: Biology, Conservation and Management (pp.
2494	325-341). https://doi.org/10.1002/9780470996027.ch21
2495	Watz, J., Bergman, E., Calles, O., Enefalk, Å., Gustafsson, S., Hagelin, A., Jonsson, B.
2496	(2015). Ice cover alters the behavior and stress level of brown trout Salmo trutta.
2497	Behavioral Ecology, 26(3), 820-827. https://doi.org/10.1093/beheco/arv019
2498	Wells, A., Grierson, C. E., MacKenzie, M., Russon, I. J., Reinardy, H., Middlemiss, C.,
2499	Hazon, N. (2006). Physiological effects of simultaneous, abrupt seawater entry and
2500	sea lice (Lepeophtheirus salmonis) infestation of wild, sea-run brown trout (Salmo
2501	trutta) smolts. Canadian Journal of Fisheries and Aquatic Sciences, 63(12), 2809–
2502	2821. https://doi.org/10.1139/f06-160
2503	Wells, A., Grierson, C. E., Marshall, L., MacKenzie, M., Russon, I. J., Reinardy, H.,
2504	Hazon, N. (2007). Physiological consequences of "premature freshwater return" for
2505	wild sea-run brown trout (Salmo trutta) postsmolts infested with sea lice
2506	(Lepeophtheirus salmonis). Canadian Journal of Fisheries and Aquatic Sciences,
2507	64(10), 1360–1369. https://doi.org/10.1139/f07-107
2508	Werner, E. E., & Gilliam, J. F. (1984). The Ontogenetic Niche and Species Interactions in
2509	Size-Structured Populations. Annual Review of Ecology and Systematics, 15, 393-
2510	425.
2511	Whelan, K. F. (1989). The angler in Ireland: game, coarse & sea. Country House.

- 2512 Whelan, K. F., & Poole, W. R. (1993). The sea trout stock collapse, 1989-1992. The
- *Conservation of Aquatic Systems: Proceedings of a Seminar Held On*, 18–19.

2514	Winter, E. R.	, Tummers, J.	. S., Aarestrup,	K., Baktoft,	H., &	Lucas, M.	C. (2016).
------	---------------	---------------	------------------	--------------	-------	-----------	------------

- 2515 Investigating the phenology of seaward migration of juvenile brown trout (*Salmo*
- 2516 *trutta*) in two European populations. *Hydrobiologia*, 775(1), 139–151.
- 2517 https://doi.org/10.1007/s10750-016-2720-z
- 2518 Wysujack, K., Greenberg, L. A., Bergman, E., & Olsson, I. C. (2009). The role of the
- environment in partial migration: food availability affects the adoption of a migratory
- tactic in brown trout *Salmo trutta*. *Ecology of Freshwater Fish*, *18*(1), 52–59.
- 2521 https://doi.org/10.1111/j.1600-0633.2008.00322.x
- 2522 Young, K. A. (2005). Life–history variation and allometry for sexual size dimorphism in
- 2523 Pacific salmon and trout. *Proceedings of the Royal Society of London B: Biological*2524 Sciences, 272(1559), 167–172.
- 2525 Závorka, L., Koeck, B., Cucherousset, J., Brijs, J., Näslund, J., Aldvén, D., ... Johnsson, J. I.
- 2526 (2017). Co-existence with non-native brook trout breaks down the integration of
- 2527 phenotypic traits in brown trout parr. *Functional Ecology*, *31*(8), 1582–1591.
- 2528 Zimmerman, C. E., & Reeves, G. H. (2000). Population structure of sympatric anadromous
- and nonanadromous *Oncorhynchus mykiss*: evidence from spawning surveys and
- 2530 otolith microchemistry. *Canadian Journal of Fisheries and Aquatic Sciences*, 57(10),
- 2531 2152–2162.
- 2532
- 2533

2534 **Table**

- 2535 Table 1. Sex ratio of smolts and sexually mature sea trout as reported from various rivers
- across Europe.
- 2537

River	Life stage	Percent females	Reference
Voss, Norway	Smolts	58	(B. Jonsson, 1985)
Tweed, Scotland	Smolts	59	(J. S. Campbell, 1977)
Nybroån, Sweden	Smolts	61	(Dellefors, 1996)
Istra, Norway	Smolts	61	(K. W. Jensen, 1968)
Bresle, France	Smolts	62	(Quéméré et al., 2011)
Jutland (several	Smolts	75	(J. Nielsen, 1994)
streams), Denmark			
Verkeån, Sweden	Smolts	75	(Svärdson, 1966)
Vindelälven, Sweden	Smolts	73	Palm, D., personal
			communication
Mean ± 1 SD	Smolts	65.5 ± 7.4	
Mørfjær, Sweden	Adult	44	(B. Jonsson et al., 2018)
Själsöån, Sweden	Adult	48	(Rubin et al., 2005)
Urvold lake, Norway	Adult	52	Davidsen, J.G., personal
			communication
Bresle, France	Adult	55	(Quéméré et al., 2011)
Bottenvassdraget,	Adult	56	Davidsen, J.G., personal
Norway			communication

Saltdalselva, Norway	Adult	58	Davidsen, J.G., personal
			communication
Vindelälven, Sweden	Adult	61	Norrfors fishladder,
			Vattenfall Vattenkraft AB
Åvaån, Sweden	Adult	63	(Alm, 1950)
Ihra, Sweden	Adult	64	(Hessle, 1935)
Voss, Norway	Adult	66	(B. Jonsson, 1985)
Verkeån, Sweden	Adult	73	(Svärdson, 1966)
Vistula, Poland	Adult	76	(Svärdson, 1966)
Oir, France	Adult	79	Marchand, F., personal
			communication
Mean ± 1 SD	Adult	61.2 ± 10.6	

2540 Figure legends

Figure 1. Endemic distribution of brown trout (Salmo trutta): dashed lines give distribution
area of anadromous populations and shaded areas give those of freshwater resident
populations. After Jonsson and Jonsson (2011). Localisation of six brown trout populations,
from north to south: River Hals 70°N (Norway), River Vosso 60°N (Norway), River
Burrishoole 54°N (Ireland), River Tamar 51°N (England), River Bresle 50°N (France) and
River Oir 48°N (France).
Figure 2. Schematic representation of brown trout (Salmo trutta) life history, and some of the
major threats affecting its abundance (© Bengt Finstad and Kari Sivertsen, NINA).
Figure 3. Mean length at age for resident (filled square) and anadromous (open dot) brown
trout (Salmo trutta) combined for the River Hals, River Vosso, River Tamar, River Bresle
and River Oir. The total age represents the freshwater age for resident trout and the sum of
the freshwater age and the sea age for anadromous. See Table S1 for details.
Figure 4. Mean length at age for brown trout (Salmo trutta) smolts at River Hals (green),
River Vosso (yellow), River Burrishoole (red), River Bresle (black) and River Oir (blue).
Values are presented ± 1 SD, except for River Burrishoole. See Table S1 for details.
Figure 5. Correlation between mean length at age for resident and anadromous brown trout
(Salmo trutta) at the River Hals (green), River Vosso (yellow), River Burrishoole (red), River
Tamar (pink), River Bresle (black) and River Oir (blue). The solid line represents the 1:1 line.
The dotted line represents the relative growth gain attributable to sea migration, with a slope

2565	estimated at 1.377 (SD: 0.138) and significantly different from 1 (Chi ² = 318.2, df = 1, P = $(1 + 1)^{12}$
2566	0.006). Values are presented \pm 1 SD, except for River Burrishoole. See Table S1 for details.
2567	
2568	Figure 6. Characterisation of the migration strategy in brown trout (Salmo trutta) along a
2569	latitudinal gradient of populations: River Hals 70°N (Norway), River Vosso 60°N (Norway),
2570	River Burrishoole 54°N (Ireland), River Tamar 51°N (England), River Bresle 50°N (France)
2571	and River Oir 48°N (France). Left panel: relative additional growth for anadromous trout
2572	related to sea age and smolt age, expressed as a growth difference with resident trout of
2573	similar age. Some values are missing when no resident trout of a similar total age were
2574	caught Right panel: age structure of returning anadromous trout, in terms of sea age and
2575	smolt age. The darker the cell, the higher the value.