# Synthesis of Phenothiazine Dyes Containing Thiophene and Diketopyrrolopyrrole pi-Bridge for Use in DSSCs 

Master's thesis in Chemical Engineering and Biotechnology
Supervisor: Bård Helge Hoff, Audun Formo Buene
June 2019

## Monica Eian Løberg

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Faculty of Natural Sciences
Department of Chemistry

## - NTNU

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## Preface

This master thesis has been carried out at the Department of Chemistry at NTNU (Trondheim) during the spring of 2019. This thesis is built on the specialization project carried out in the autumn of 2018.

I would like to thank my supervisors, Professor Bård Helge Hoff and PhD Candidate Audun Formo Buene. Your insight in the field of DSSCs has been crucial for this thesis, as well as your practical knowledge and motivational conversations. I would also like to thank PhD Candidate David Moe Almenningen and post doc. Amsalu Efrem for all help concerning laboratory work and theoretical questions. Additionally, I am grateful to department Engineer Roger Aarvik for supplying the chemicals, PhD Susana Villa Gonzalez for MS analysis and Senior Engineer Torun Margareta Melø for guidance regarding NMR. To the members of the solar cell laboratory, thank you for all the good conversations, both professional and non-academic.

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## Abstract

Dye-sensitized solar cells (DSSCs) is an emerging new photovoltaic (PV) technology, allowing solar cells to be semi-transparent, flexible and of tuneable colour. Under ambient light conditions, DSSCs have out-performed most other long established PV technologies. Hot applications of DSSCs include building integration and powering an increasing number of devices in the Internet of Things (IoT).

Our group at NTNU has initially focused on phenothiazine based dyes. A method to lower the band gap and increase light harvest is to incorporate electron deficient units into the conjugated system. This thesis presents chemistry to synthesize three novel phenothiazine dyes containing one (MEL1) or two thiophenes (MEL2) or diketopyrrolopyrrole (MEL3) as part of the $\pi$-bridge. After a four-step synthesis, MEL1 and MEL2 were synthesized in a total yield of $44 \%$ and 20\%, respectively. The novel dye MEL3 was synthesized in a five-step synthesis, and had the most red-shifted adsorption spectra of the three dyes.


## Sammendrag

Fargestoff-solceller er nye og voksende fotovoltaiske teknologier som gjør at solceller kan være halvtransparente, fleksible og av justerbar farge. Disse solcellene har utkonkurrert de fleste andre etablerte fotovoltaiske teknologier under naturlige lysforhold. Noen av bruksområdene til fargestoff-solcellene er bygningsintegrasjon og strøm til gjenstander i tingenes internett.

Vår forskningsgruppe på NTNU har forsket på fenotiazin-baserte fargestoffer. En metode for å senke båndgapsenergien og øke lysinnsamlingen er ved å lage fargestoff med elektronfattige enheter i det konjugerte systemet. Denne masteravhandlingen presenterer syntesen av tre nye fenotiazin fargestoffer som inneholder en (MEL1) eller to (MEL2) tiofener eller diketopyrrolopyrrole (MEL3) som en del av $\pi$ broen. Etter en firetrinns syntese ble MEL1 og MEL2 syntetisert med et totalt utbytte på henholdsvis $44 \%$ og 20\%. Fargestoffet MEL3 ble syntetisert med en femtrinns syntese, og hadde det mest rødskiftede adsorpsjonsspekteret av de tre fargestoffene.


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Numbered Compounds


















## Abbreviations and Symbols

$\delta \quad$ Chemical Shift${ }^{1}$ H NMR Proton Nuclear Magnetic Resonance
${ }^{13}$ C NMR Carbon Nuclear Magnetic Resonance
CB Conduction Band
CMD Concerted Metalation Deprotopnation
d Doublet
DCM Dichloromethane
DMF Dimetylformamid
DMSO Dimethyl Sulfoxide
DPP Diketopyrrolopyrrole
DSSC Dye-Sensitized Solar Cell
$\mathrm{E}^{+} \quad$ Electrophile
$\mathrm{e}^{-} \quad$ Electron
$\mathrm{E}_{f} \quad$ Fermi Level
$\mathrm{E}_{0-0} \quad$ Band Gap Energy
EAS Electrophilic Aromatic Substitution
eq. Equivalents
EWG Electron Withdrawing Group
FF Fill Factor

## Abbreviations and Symbols

## FTO Fluorine-Doped Tin Oxide

h Hours
HRMS High Resolution Mass Spectrometry
IoT Internet of Things
J Coupling Constant [Hz]
J Electric Current
$\mathrm{J}_{s c} \quad$ Short Circuit Current
L Ligand
LHE Light Harvesting Efficiency
m Medium
m Multiplet
$\mathrm{m}_{p} \quad$ Melting Point
MS Mass Spectrometry
NMR Nuclear Magnetic Resonance
$\mathrm{Nu}^{-}$Nucleophile
P Power
$\mathrm{P}_{\text {in }} \quad$ Density of Incident Light
$\mathrm{P}_{\max }$ Maximum Power Point
PCE Solar to Electical Energy Conversion Efficiency
ppm Parts per Million
PV Photovoltaic
$\mathrm{R}_{f} \quad$ Retention factor
r.t. Room Temperature
s Singlet
s Strong
SPhos 2-Dicyclohexylphosphino-2,6-dimethoxybiphenyl
t Triplet
TCO Transparent Conductive Oxide
td Triplet of Doublet
TEOME Triethylene Oxide Methyl Ether
TM Target Molecule

## V Voltage

$\mathrm{V}_{m} \quad$ Voltage at Maximum Power Point
$\mathrm{V}_{o c} \quad$ Open Circuit Voltage
VB Valence Band
w Weak
X Halide
XPhos 2-Dicyclohexylphosphino-2,4,6-triisopropylbiphenyl

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| :---: |
| Chapter |}

## Introduction

In recent years, dye-sensitized solar cells (DSSCs) has become an emerging photovoltaic technology allowing for harvesting of solar energy. ${ }^{[1]}$ Under ambient light conditions DSSCs have out-performed most other long established PV technologies. ${ }^{[2]}$ Dye-sensitized solar cells (DSSCs) profit from being flexible, semitransparent and of tuneable colour. ${ }^{[3]}$ The focus area for DSSCs are, among others, building integration and powering an increasing number of devices in the Internet of Things (IoT). ${ }^{[4,5]}$ One of the leading DSSC-dyes is N719 based on a Rucomplex, with a solar-energy-to-electricity conversion efficiency of $11 \%{ }^{[6,7]} \mathrm{Re}-$ cently fully organic, metal-free, dyes have shown great progress in the field of DSSCs. ${ }^{[8]}$

### 1.1 Background and Aims

The worlds population is growing and together with higher standards of living this creates a great demand of energy. The energy consumption growth form 2016 to 2017 was $2.2 \%$, and is expected to increase each year. ${ }^{[9]}$ More energy must be produced and preferably in a sustainable way from renewable energy sources, which covered $40 \%$ of the global energy consumption in 2018. ${ }^{[10]}$ The use of these energy sources is rising and the renewable power growth in 2017 was $17 \%$ from the year before. ${ }^{[9]}$

According to the International Energy Agency ${ }^{[10]}$ solar photovoltaics (PV) will be the fastest growing energy source in the following years. A way to provide more
energy is to make the solar cells more efficient. Many strategies could be used to achieve this goal. Dye-sensitized solar cells (DSSCs) can be tuned to adsorb light energy in varied wavelengths and can result in a wider light adsorption range. This is due to a large selection of conjugated systems that can be combined.

Phenothiazine-based dyes have been the main focus in our group at NTNU. Electron deficient units can be incorporated into the conjugated system, to lower the band gap and increase light harvest. This thesis presents synthesis of phenothiazine dyes containing one or two thiophenes, or thiophenes together with diketopyrrolopyrrole (DPP) as part of the $\pi$-bridge. The phenothiazine-based dyes have shown qualities suited for dye-sensitized solar cells. ${ }^{[1]}$ All the dyes in the MEL-series have a donor - $\pi$-bridge - acceptor system. The use of DPP in photovoltaics has yielded the first high-performance blue DSSC, with a power conversion efficiency above $10 \% .{ }^{[5]}$ Blue dyes have also shown an excellent ability of absorbing the incident photons in the red and near-infrared range. ${ }^{[11]}$ Dyes made of DPPs can become blue, and the color itself is desirable for building integration.

### 1.2 Solar Cells

A solar cell is a device that converts light directly into electricity. This section will first present a short introduction to general photovoltaics, then the traditional silicon based solar cells. Thereafter, dye-sensitized solar cells and their energic considerations and requirements will be presented, and lastly the design of target molecules for this thesis.

### 1.2.1 Working Principles

In most of the renewable energy sources mechanical energy is converted to electrical energy, such as for wind and hydroelectric power plants. Solar cells use photon energy, which are particles of light, to excite electrons so that they can be used in an external circuit to make electricity. ${ }^{[12]}$ The light creates mobile charge-particles in a semiconductor which can separate the charges. ${ }^{[13]}$ This charge separation allows for transportation to the external circuit and thus generation of electric current.

Organic DSSCs work fundamentally different than traditional inorganic silicon solar cells. ${ }^{[14]}$ The inorganic solar cells have a direct generation of the electronhole pair. In DSSCs the electron has to be injected into a semiconductor, and after a migration through the external circuit, the electron is re-introduced into the cell
on a metal electrode. The electron is flowing from the electrode into an electrolyte and back to the dye where the dye reverts to its ground state.

Generally, for all kinds of PV devices, under open circuit there is no current, where the potential is called $\mathrm{V}_{o c}$. This value together with the short circuit current, $\mathrm{J}_{s c}$, represent the absolute maximum values of the current and voltage for the solar cell. In order to get the maximum power from the solar cell, the values of J and V must be chosen so that $\mathrm{P}=\mathrm{I} \cdot \mathrm{V}$ is maximized, as shown in Figure 1.1.


Figure 1.1: Illustrates the IV-curve (red) of a solar cell. In addition, the values of I and V which maximize the area (power) are highlighted (blue curve) to show that the real values of I and V are smaller than $\mathrm{J}_{s c}$ and $\mathrm{V}_{o c}$, respectively. $\mathrm{J}_{m}$ and $\mathrm{V}_{m}$ are the maximum/optimized values.

The power, P , is the ultimate measurement of how good a solar cell is, and should be as high as possible. The fill factor, FF, is a measure of how ideal a solar cell is. ${ }^{[12]}$

$$
F F=\frac{J_{m} \cdot V_{m}}{J_{s c} \cdot V_{o c}}
$$

Where $\mathrm{J}_{m}$ and $\mathrm{V}_{m}$ is the current and voltage, respectively, of the maximum power point, $\mathrm{P}_{\max }$. The performance of the cell is often given by the solar-to-electrical energy conversion efficiency (PCE), also called the power conversion efficiency. ${ }^{[15]}$

$$
P C E=\frac{J_{s c} \cdot V_{o c} \cdot F F}{P_{i n}}
$$

Where $\mathrm{P}_{i n}$ is the intensity of incident light.

### 1.2.2 Silicon Solar Cells

An important feature of the silicon solar cell is the band gap, which corresponds to the energy difference between the valence band $(\mathrm{VB})$ and the conduction band (CB). Electrons are excited from the VB to the CB if an energy equivalent to or larger than the band gap is provided. Another crucial part of the solar cell are the semiconductor layers. This is where electrons are freed and current is created. There are two different classifications of semiconductors, $p$-type or $n$-type, ${ }^{[16]}$ as presented in Figure 1.2.


Figure 1.2: The silicon-based PV cell. ${ }^{[17]}$

A $p n$-junction is created by doping the two sides of the semiconductor with $n$ - and p-type on each side. By doping a semiconductor, the Fermi level, Ef, is shifted towards the VB for $p$-type, and CB for $n$-type. ${ }^{[12]}$ Because the two sides of the semiconductor are in thermal equilibrium, the Fermi level of the entire semiconductor must be equal, thus creating a potential difference between the two doped sides. This potential difference creates an electric field from the $n$-side to the $p$ side. ${ }^{[18]}$ As a consequence, any electrons excited on the $p$-side will be transported through the $p n$-junction and end up on the $n$-side while a hole will remain on the $p$-side. ${ }^{[19]}$ The separation of the electron-hole pair by an electric field increases their lifetime, making the time required for transport of the electrons to an external circuit sufficient to take advantage of its energy.

### 1.2.3 Dye-Sensitized Solar Cells

The dye-sensitized solar cell is a type of thin film solar cell, and can be considered to be a technology between the second and third generation solar cells. ${ }^{[15,20]}$ The DSSCs, also referred to as Grätzel cells, ${ }^{[21]}$ vary a lot from each other in architecture, but they have some common operating principles, as shown in Figure 1.3. The DSSC main components are the semiconducting metal oxide material, electrolyte, dye-materials and counter-electrode. ${ }^{[20]}$ The main difference from silicon based solar cells is the charge separation process, where the generated exciton injects an electron to the semiconductor for DSSCs.


Figure 1.3: Energetic diagram of $n$-type DSSC. ${ }^{[22]}$
The light goes through the transparent conductive oxide (TCO) first. The TCO works as an anode, and the most common type of TCO is fluorine-doped tin oxide (FTO). Attached to the anode is the nano-crystalline oxide semiconductor, $\mathrm{TiO}_{2}$. This type of semiconductor, $\mathrm{TiO}_{2}$, proposed in 1991, is still the most used semiconductor in DSSCs. ${ }^{[3,23]}$ The dyes can then be loaded onto the nano-crystalline oxide semiconductor. Ideally, the dyes will form a monolayer on the film. The dyes inject electrons to the semiconductor when photoexcitation occurs. Then the is in its oxidized form and is regenerated by the electrolyte, containing iodine in the anionic forms $\mathrm{I}^{-}$and $\mathrm{I}_{3}^{-}$. This is the most used electrolyte, due to its slow recombination rate with injected electrons and fast diffusion properties. ${ }^{[24,25]}$ Electrolytes with $\mathrm{Co}(\mathrm{II}) / \mathrm{Co}(\mathrm{III})$ and $\mathrm{Co}(\mathrm{I}) / \mathrm{Co}(\mathrm{II})$ have also shown to be promising, and have the advantage over Iodide-couple due to their non-volatility, non-toxicity and
higher open-circuit voltage. ${ }^{[18,26,27]}$ The higher open-circuit voltage is caused by their more positive redox potential. A counter electrode, the cathode, is at the bottom of the solar cell, and normally contains platinum, Pt, and TCO. The electron that was donated to the semiconductor will be transported to an external circuit via the TCO-anode.

The optimization of the DSSC can be done in different ways. For changing the charge transport, the electrolyte or semiconductor can be modified, and the dyes for changes in spectral properties. Despite the different working principles of the silicon solar cells and DSSCs, the DSSCs can also be divided into $p$ - and $n$-type. The process described above is an $n$-type solar cell. In contrast, the $p$-type DSSCs have the dyes and semicondutor on the catode, and injects electrons to the electrolyte. ${ }^{[28,29]}$

### 1.2.4 Energic Consideration of Dyes in DSSC

In order for the dye to work in a DSSC, they have to absorb photons from the sun-light. The light harvesting efficiency (LHE) is a measure of the absorption as a function of the concentration of the dye-molecules absorbed onto the $\mathrm{TiO}_{2}$ surface, extinction coefficient and the adsorption range of the dye. ${ }^{[22]}$ The dye should be panchromatic, meaning that the dye is sensitive to all wavelengths of visible light. Preferably, also for some wavelengths near the infrared light. ${ }^{[30]}$

To be able to inject an electron into the $\mathrm{TiO}_{2}$ a charge separation process has to take place. ${ }^{[31]}$ The charge separation energy is proportional to the exciton binding energy, which depends on both the geometry of the dye and the electronic structure. The driving force for electron injection is defined as the difference between the LUMO level of the dye and the semiconductors conduction band, as shown in Figure 1.3. ${ }^{[22]}$ To be thermodynamically possible, the LUMO level has to be above the $\mathrm{TiO}_{2}$ conducting band. The energy difference, $\mathrm{E}_{0-0}$, between the HOMO and LUMO level of the dye is one of the driving forces for excitation of electrons.

The band gap energy, $\mathrm{E}_{0-0}$, of the sensitizers can be estimated from the adsorption spectra. This is done by drawing a tangent in the inflection point to the right of the absorption maxima. The intersection between this tangent and the x-axis reveals the absorption onset wavelength, $\lambda$, that is used to calculate the band gap energy, $\mathrm{E}_{0-0}[\mathrm{eV}]$ :

$$
E_{0-0}=\frac{h \cdot c}{\lambda}
$$

### 1.2.5 Dye Design for DSSCs

In the case of a light absorbing dye, many considerations regarding the design have to be made. The following criteria for a dye have been suggested: ${ }^{[20,32]}$

- The visible region and even part of the near-infrared spectrum should be covered in the dyes absorption spectrum.
- The absorbing dye must have an anchoring group to strongly bind to the semiconductor surface. This group could be $-\mathrm{COOH},-\mathrm{H}_{2} \mathrm{PO}_{3},-\mathrm{SO}_{3} \mathrm{H}$, etc.
- The LUMO level of the sensitizer should be higher in energy than the conduction band of the metal oxide. This ensures an efficient transfer between the excited dye and the CB.
- The HOMO level of the sensitizer has to be more positive than the redox potential of the electrolyte in order to get regeneration of the dye.
- The sensitizers have to be thermally, photo- and electrochemically stable.
- Must be soluble in common organic solvents compatible with electrolytes.

Organic sensitizers are in most cases based on a donor, a $\pi$-bridge and an acceptor ( $\mathrm{D}-\pi-\mathrm{A}$ ) architecture, as shown in Figure 1.4. The backbone of this dye is phenothiazine, which works as an electron donor. Thiophene is the $\pi$-bridge and cyanoacrylic acid is the electron acceptor/anchoring group. These three parts are


Figure 1.4: Design concept for new dyes, which should include a donor, a $\pi$ bridge and an acceptor, with MEL3 as an example.
all what is needed to make a dye, but in order to increase the efficiency, some extra components could be added. The aromatic group pyrene is added as an auxiliary donor and the hexyl and the triethylene oxide methyl ether (TEOME) chain as anti aggregation components. Diketopyrrolopyrrole (DPP) extends the conjugation of the $\pi$-bridge by introducing an electron withdrawing group (EWG). Incorporation of an electron-deficient group reduces the energy gap of the dyes and broadly extends the spectral coverage. ${ }^{[33]}$ Cyanoacrylic acid is one of the most common anchoring groups in DSSCs, but a large selection of other anchoring groups can also be used. ${ }^{[34,35]}$

The largest difference between MEL3 and the two more simple dyes MEL1 and MEL2, is the DPP-unit. This unit has been evaluated in other dyes used in DSSCs. ${ }^{[5]}$ The addition of DPP makes the dye blue, unlike the red dyes MEL1 and MEL2. Blue DSSCs have an excellent ability of absorbing the incident photons in the red and near-infrared range. ${ }^{[11]}$ Dyes based on the DPP-unit have shown good efficiency in the cobalt electrolyte system, $[\mathrm{Co}(\mathrm{bpy}) 3]^{3+/ 2+},{ }^{[5]}$ which has the advantages of non-volatility and non-toxicity. The dye displayed in Figure 1.5 was used in a DSSC with the previous mentioned electrolyte, and gave an energy conversion efficiency of $10.1 \%$. Another DPP-based dye is the Dyenamo Blue, with an energy conversion efficiency of $7.3 \%$ and an adsorption maxima at 584 nm (49 000 $\mathrm{M}^{-1} \mathrm{~cm}^{-1}$, in DCM). ${ }^{[36,37]}$ The DPP-unit has been attached to a simpler phenothiazine donor group in a research done by Zang et al. ${ }^{[38]}$ This resulted in a efficiency of $5.2 \%$ with a diphenyl-DPP.


Figure 1.5: The dye DPP-17 dye with a DPP-unit performed a PCE above $10 \% .^{[5]}$
The first phenothiazine dye for DSSC was developed in 2007. ${ }^{[39]}$ Since then it has been shown that phenothiazine-based dyes are truly promising. ${ }^{[40]}$ Phenothiazine is electron donating, with its electron rich heteroatoms. The research group at

NTNU has developed many DSSCs with phenothiazine. ${ }^{[41]}$ Two of the dyes are AFB-19 and AFB-31 made by Buene and Boholm with a power conversion efficiency of $5.00 \%$ for both dyes. ${ }^{[41,42]}$ AFB-19 showed a molar extinction coefficient of $24450 \mathrm{M}^{-1} \mathrm{~cm}^{-1}$ at absorption 441 nm . A molar extinction coefficient of 19 $844 \mathrm{M}^{-1} \mathrm{~cm}^{-1}$ at 454 nm was seen for AFB-31.



Figure 1.6: Two phenothiazine dyes, AFB-19 (PCE = 5.00\%) and AFB-31 (PCE $=5.00 \%$ ), synthesized by Buene and Boholm. ${ }^{[41,42]}$

### 1.2.6 The Diketopyrrolopyrrole Unit

The DPP-chromophore was first discovered in 1974 by Farnum et al., ${ }^{[43]}$ as a byproduct. The DPP has shown to be easily synthesized, strongly absorbing and thermally stable. ${ }^{[44]}$ The first DPP-based small-molecule dye for use in DSSCs was reported in 2010. ${ }^{[6]}$ The DPP-unit as photovoltaic materials has been intensively explored since then. ${ }^{[45]}$ By functionalizing the DPP-unit it can be tuned towards many different colors, from orange-yellow via blue-red to violet. This can be accomplished by exchanging the substituents in the para- and meta-position of the aromatic group attached to the DPP chromophore. ${ }^{[46]}$

The incorporation of the DPP chromophore in a dye can lead to high absorbance and emittance in the visible and near infrared region. ${ }^{[47]}$ This is due to the tuning of the HOMO and LUMO energy levels which can be conveniently accomplished by alternating the $\pi$-conjugated systems, indicated by electrochemical measurements. ${ }^{[6]}$ The integration of the DPP-unit with an electron donating unit, in this case phenothiazine, can give rise to compounds with low band gaps. Band gaps of DPP-based materials as low as 1.13 eV have been reported. ${ }^{[48]}$

The DPP-unit is complex, and has many reactive centers. An overview of the potential reactivity of the DPP is presented in Figure 1.7. ${ }^{[49]}$


Figure 1.7: Potentially reactive centers for DPP derivatives, where $\mathrm{E}^{+}$is electrophile, $\mathrm{Nu}^{-}$is nucleophile and X is halide. ${ }^{[49]}$

### 1.2.7 Target Molecules

The target molecules for this master thesis are the three novel dyes MEL1, MEL2 and MEL3 shown in Figure 1.8. They all have the phenothiazine, pyrene, thiophene and triethylene oxide methyl ether (TEOME) chain in common, which is inspired by Buenes dyes AFB-19 and AFB-31, ${ }^{[41,42]}$ displayed in Section 1.2.6. MEL3 has a DPP-group similar to the ones in DPP-17 and Dyenamo Blue.




Figure 1.8: The target molecules are the three phenothiazine-based dyes MEL1, MEL2 and MEL3.

The dyes MEL1 and MEL2 only differ from each other by one thiophene. The dyes have a ( $\mathrm{D}^{‘}-\mathrm{D}-\pi-\mathrm{A}$ ) and ( ${ }^{‘}$ - $\mathrm{D}-\pi-\pi-\mathrm{A}$ )-system, respectively. MEL3 has a

DPP-unit between the two thiophenes and a phenyl, that differs from MEL2. The phenothiazine has a pyrene attached to increase the donor quality of the dye. ${ }^{[42]}$ The reason for attaching a TEOME-group and hexyl groups is to decrease the ability for the dyes to stack and aggregate. ${ }^{[50]}$ The TEOME-chain, together with the phenyl group, is electron-rich and can increase both the solubility and donor quality additionally. The thiophene groups lower the charge recombination, and improve the absorption of the light by red-shifting the absorption spectrum. ${ }^{[51]}$ The DPP component is electron withdrawing. The aim of adding this EWG is to cover more of the visible and near-infrared region of the adsorption spectra, then lower the band gap energy. The anchoring group is a cyanoacrylic acid group that absorbs well to the semiconductor material, and allows electron injection into $\mathrm{TiO}_{2}$. A proposed retrosynthesis of target molecule MEL1 and MEL3 is presented in Schemes 1.1, 1.2 and 1.3. A similar retrosynthesis can be used for MEL2, as described for MEL1.




Scheme 1.1: A proposed retrosynthetic route for MEL1. MEL2 has a largely similar retrosynthetic route.

The brominated phenothiazine $\mathbf{1}$ was synthesized in the specialization project. The first step described in this thesis towards the target molecules MEL1 and MEL2 is the Suzuki-Miyaura cross-coupling from the symmetric phenothiazine-backbone 1 to a monosubstituted pyrene compound 3. The Suzuki-Miyaura is followed by borylation to make the boronate functionalized derivative 4, before one more Suzuki to yield the aldehyde 5. The last step toward the dye is the Knoevenagel condensation to form the cyanoacrylic acid MEL1. The total route is a linear four-step synthesis.

Synthesis of MEL3 could be carried out in a convergent strategy. The synthetic route for the phenothiazine donor group 4 is previous described. Two proposed retrosyntheses for the dye MEL3 will be presented, retrosyntheses A and B, where A, as shown in Scheme 1.2, will be presented first. The first step towards the DPP building-block is an alkylation to form the alkylated derivative 9 of the commercially available dithiophene-DPP 7. The next synthesis is a NBS bromination, where bromines are introduced on both sides of the DPP 12. A Suzuki-Miyaura cross-coupling can be used to introduce a benzaldehyde on one side of the DPP 14, before one more Suzuki-Miyaura reaction binds the DPP aldehyde 14 with the donor group 4 to form the donor- $\pi$-bridge system $\mathbf{1 8}$. The last step is the same as previous described for the two other dyes MEL1 and MEL2, a Knoevenagel condensation to yield target dye MEL3, in a seven-step synthesis.






Scheme 1.2: The proposed retrosynthetic route A for MEL3.

The retrosynthesis route B is presented in Scheme 1.3. This is a shorter route, with only five step, and uses the direct arylation instead of Suzuki-Miyaura crosscoupling. The synthesis of the alkylated dithiophene 9 and the phenothiazine donor group 2 are previous described. To carry out the $\pi$-bridge $\mathbf{1 3}$ a direct arylation takes place from the alkylated DPP 9. Another direct arylation between the
phenothiazine-based compound 2 and the DPP-aldehyde 13 yields the dye precursor 18. As described for the other dyes, the last step is Knoevenagel condensation to form the target dye MEL3.



Scheme 1.3: The proposed retrosynthetic route B for MEL3.

### 1.3 Synthesis

### 1.3.1 Suzuki-Miyaura Cross-Coupling

The Suzuki-Miyaura cross-coupling creates a new carbon-carbon bond from an organoboron compound and aryl/vinyl halide, ${ }^{[52]}$ and is considered one of the most versatile methods for carbon-carbon bond formation. ${ }^{[53]}$ The reaction can be considered as a nucleophilic substitution of halide atoms with various carbon nucleophiles. ${ }^{[54]}$ This method can use mild reaction conditions, water can be used as solvent and some of the by-products are water soluble, these are some of the advantages to the Suzuki-Miyaura cross-coupling. Other advantages are the availability of many boronic acids and the method tolarates a variety of functional groups in the starting material, due to the generally high chemo- and regioselectivity. ${ }^{[55]}$ Some common by-products of the Suzuki reaction are the reduced product and homocoupling of arylboronic acid through an electroreductive or electrooxidative reaction. ${ }^{[56,57]}$ A proposed mechanism for the palladium-catalyzed reaction is shown in Scheme 1.4.

## Mechanism




Scheme 1.4: Proposed mechanism for Suzuki-Miyaura cross-coupling between the symmetric phenothiazine $\mathbf{1}$ and a pyreneboronic acid to form the monosubstituted phenothiazine 2. ${ }^{[58]}$

The suggested mechanism for Suzuki-Miyaura cross coupling proceeds trough a four step cycle, oxidative addition, metathesis, transmetalation and reductive elimination.

1. Oxidative addition is in most cases the rate determining step of the catalytic cycle. ${ }^{[58]}$ This step breakes the carbon-halogen bond, allowing two new bonds to form with the palladium complex, resulting in oxidized palladium from $\operatorname{Pd}(0)$ to $\operatorname{Pd}(\mathrm{II})$. The result of this transformation is a reduced electron density of the metal. Electron rich ligands, L, stabilizes the $\operatorname{Pd}(I I) .{ }^{[59]}$
2. Metathesis is the step where the halide exchanges with a base, to give a more reactive Pd-complex in the next step.
3. Transmetalation is the reaction where reactive groups are transferred from one metal to another, or more specific in the case of Suzuki-Miyaura, from the Pd complex to organoboron. The base that was added in the previous step, metathesis, is now exchanged with an aryl specie. ${ }^{[60]}$
4. Reductive elimination is the last step where the two aryl species on the Pd-complex are eliminated. This regenerates the active $\operatorname{Pd}(0)$ catalyst, and forms a bond between the two aryl units. ${ }^{[61]}$

The Suzuki cross-coupling reaction is frequently used, but the mechanism is not fully understood. ${ }^{[59]}$ Oxidative addition was earlier reported as the rate determining step, ${ }^{[62]}$ but research indicates that the reactivity is more complex and depends on many factors. ${ }^{[63]}$ Electron withdrawing groups, EWG, in the aryl halide compound will increase the rate of oxidative addition, owing to the weakening of the C-X bond. ${ }^{[64]}$

Suzuki-Miyaura cross-coupling can also be used to synthesis DPP-based $\pi$-conjugated systems, ${ }^{[54]}$ there the DPP-unit is an EWG and preferably the aryl halide. The two electronwithdrawing amide in DPP can facilitate nucleophilic aromatic substiution in the absence of palladium catalyst. ${ }^{[65]}$

## Catalyst and Ligands

There are a large number of possible variables in a Suzuki-Miyaura reaction, such as metal source, ligand, base, solvent, temperature, etc. ${ }^{[66]}$ The metal/ligand systems that facilitate the Suzuki-Miyaura cross-coupling with different electrophiles are the parameters that have been the most extensively studied. ${ }^{[67]}$ Palladium is the most widely used metal as catalyst, with a choice between $\mathrm{Pd}(0)\left(\mathrm{Pd} / \mathrm{C}, \mathrm{Pd} / \mathrm{Al}_{2} \mathrm{O}_{3}\right.$, $\left.\mathrm{Pd}_{2}(\mathrm{dba})_{3}\right)$ or $\mathrm{Pd}(\mathrm{II})\left(\mathrm{PdCl}_{2}, \mathrm{Pd}(\mathrm{OAc})_{2}\right) \cdot{ }^{[67]}$ The catalytic species can be formed in situ using the $\operatorname{Pd}(0)$ source and a necessary ligand or a preformed catalyst can be introduced using the $\mathrm{Pd}(\mathrm{II})$. Careful choice of ligand can facilitate two of the four steps of the catalytic cycle, shown in Scheme 1.4. The use of electron donating ligands can increase the electron density around the metal, and accelerate the oxidative addition. If a bulky ligand is present, it helps the elimination step. ${ }^{[68]}$ An overview of their impact on the efficacy of catalysts using these ligands are shown in Figure 1.9.

Substituents fixed configuration of $\mathrm{R}_{2} \mathrm{P}$ over bottom ring, enhancing rate of reductive elimination
$\mathrm{R}_{1}, \mathrm{R}_{2} \neq \mathrm{H}$ prevents cyclometallation, increasing stability
$\mathrm{R}_{1}, \mathrm{R}_{2}=$ large group (isopropyl increases [ $\left.\left.\mathrm{L}_{1} \mathrm{Pd}(0)\right]\right)$



Lower aryl ring:
Increases size of ligand, slow rate of oxidation by $\mathrm{O}_{2}$. Allows stabilizing Pd-arene interactions Promotes reductive elimination

Figure 1.9: Structural features of dialkylbiarylphosphines ligands and their impact on catalysis. ${ }^{[69]}$

Some common dialkylbiarylphosphine ligands, L, are shown in Figure 1.10, where all are bulky, electron-rich and monodentate (only one donor atom used to bond to the central metal atom or ion). ${ }^{[69]}$

L1

L2 (MePhos)

L3 (DavePhos)

L4 (JohnPhos)

L5

L6

L7 (SPhos)

L8 (XPhos)

L9 (RuPhos)

$L 10$

Figure 1.10: Common ligands in Suzuki-Miyaura cross-coupling reactions. ${ }^{[69]}$

### 1.3.2 Borylation

Pd-catalyzed borylation allows pinacolborane and aryl halides to react and form an arylboron compounds. This reaction has proved to be a powerful tool, due to the introducing of a Suzuki-Miyaura reactive group. ${ }^{[70]}$ Borylation needs base and tetiary amine, $\mathrm{Et}_{3} \mathrm{~N}$ is especially important since it favours boron-carbon bond synthesis. ${ }^{[71]}$ A proposed mechanism is shown in Scheme 1.5.



Scheme 1.5: Proposed mechanism for Pd-catalyzed borylation between the phenothiazine 2 and pinacolborane to form the borylated product 4. ${ }^{[72,73]}$

The mechanism for Pd-catalyzed borylation has similarities with the Suzuki-Miyaura cross-coupling mechanism, and consists of oxidative addition, transmetalation and reductive elimination. The mechanism suggest that the boride acts as the active transmetalating anion, resulting in the interaction between triethylamine and pinacolborane, which forms an ion pair. ${ }^{[72]}$

### 1.3.3 Knoevenagel Condensation

The Knoevenagel condensation synthesizes an $\alpha, \beta$-unsaturated compound by an amine catalyzed reaction of aldehydes/ketones and a carbon nucleophile. ${ }^{[52]}$ This reaction is an important tool for introducing the anchoring group in the organic dyes for use in DSSCs. ${ }^{[35,74,75]}$ The Knoevenagel condensation proceeds through a general base-catalyzed Aldol condensation. A proposed mechanism is displayed in Scheme 1.6. The amine reacts with the carbonyl group, in this case an aldehyde, and forms an iminium. The iminium and cyanoacetic acid react by a basecatalyzed reaction to form a $\mathrm{C}=\mathrm{C}$ bond, where an $\alpha, \beta$-unsaturated compound is formed.





Scheme 1.6: Proposed mechanism for Knoevenagel condensation between the aldehyde 5 and cyanoacetic acid to furnish the dye MEL1. ${ }^{[52,58,76]}$

### 1.3.4 Amide Alkylation

Amides have two possible nucleophiles, oksygen and nitrogen. Typical alkylation agents are alkyl halides. ${ }^{[52]}$ The amide alkylation, a type of nucleophilic aliphatic substitution, yields a higher substituted amide. The $\mathrm{S}_{N} 2$ mechanism for the alkylation is presented in Scheme 1.7. ${ }^{[52]}$ The DPP-unit is usually alkylated to improve the solubility. ${ }^{[77]}$


Scheme 1.7: Proposed mechanism for amide alkylation where hexyl is added to the DPP-nitrogen at coumpound 7, to form product 9. ${ }^{[52]}$

In 2010, it was assumed that the alkylation only occurs on the nitrogen. ${ }^{[78]}$ One year after it was reported that $N, N-, N, O$ - and $O, O$-alkylation occurs, with low selectivity. ${ }^{[77,79]}$ The reason for this observation is influenced by the delocalization of the $\pi$-electrons along the O-C-N bonds, as shown in Scheme 1.8. ${ }^{[80]}$

The ratio of $O$ - and N -alkylation depends on the reaction conditions, such as the reactivity of the alkylating agent and the reaction temperature. In a study by Mijin et al., ${ }^{[80]}$ was it found that the $N$-alkylation was more present than the $O$-alkylation when the alkylation agent was alkyl halide. The research indicated that the conversion of the reaction was low, and and the size of the alkyl-group does not influence the degree of conversion. On the other hand, research done by Zhao et al. ${ }^{[77]}$ showed that the selectivety of the $\mathrm{N}, \mathrm{N}-/ \mathrm{N}, \mathrm{O}-/ \mathrm{O}, \mathrm{O}$-isomers varied with different alkyl reagents. The relative amount of the isomers for 2-ethylhexyl was 36/28/11, and 63/17/trace for octyl.


Scheme 1.8: $O$ - and $N$-alkylation of amides. ${ }^{[80]}$

### 1.3.5 NBS Bromination

Bromination of aromatic compounds often occurs through an electrophilic aromatic substitution, EAS. ${ }^{[52]}$ Lewis acid is normally needed to achieve desirable rates when adding halogens to aromatic compounds, but NBS or NCS $\left(\mathrm{Br}^{+} / \mathrm{Cl}^{+}\right.$ source) can halogenate moderately active aromates without Lewis acid. The NBS bromination of thiophenes in acetic acid is considered to have a high degree of conversion. ${ }^{[81]}$ Scheme 1.9 shows a proposed mechanism for the NBS bromination. ${ }^{[52]}$


Scheme 1.9: Proposed mechanism for EAS with NBS and thiophene on DPP. ${ }^{[52]}$ The bromination was done on both sides of the DPP-unit, to synthesize the dibrominated dithiophene-DPP 12.

### 1.3.6 Direct Arylation

Direct arylation is a convenient way of synthesizing biaryl compounds, where one of the aryls is a privileged heteroaryl. ${ }^{[82,83]}$ This palladium-catalyzed arylation is a valuable and efficient emerging alternative to the established metal-catalyzed cross-coupling, such as the previous described Suzuki-Miyaura. ${ }^{[84]}$ Thiophenes are easily coupled in the direct arylation, without any adjustments. ${ }^{[85]}$ This reaction is an environmentally benign and atom-efficient pathway for synthesis of $\pi$ conjugated systems. ${ }^{[86]}$ The mechanism proceeds via activation of the C-H bond in a transmetalation catalytic cycle to form the C-C bond, as shown in Scheme $1.10 .{ }^{[86]}$

The exact mechanism of the direct C-H arylation is not known, but there has been established an agreement on pathway A or B when pivalic acid is present, as shown in Scheme 1.10. ${ }^{[86,87,89]}$ In pathway A, the pivalic acid interacts reversibly with the Pd-catayst, while in pathway B the pivalic acid is bound to palladium throughout the catalytic cycle. Pathway A has large similarities with the Suzuki-Miyaura cross-coupling, described in Section 1.3.1, and other Pd-catalyzed cross-coupling mechanisms. Both the reductive elimination and oxidative addition are quite similar, what differs is the addition of the second aryl-compound to the Pd-catalyst, called the concerted metalation deprotonation (CMD). Both aryl-compounds bind directly to the Pd-catalyst, which prevents formation of the unstable Aryl-Pd-R complex between the metathesis and transmetalation step in the Suzuki-Miyaura reaction. Research on the CMD step done by Biswas et al. ${ }^{[87]}$ showed that cleavage of the C-H bond occurs when an oxygen from the pivalic acid decoordinates from the Pd-centre, while the aryl-C still coordinates with Pd. The pivalic acid can then easily form an $\mathrm{O}-\mathrm{H}$ bond with the aryl- H .

In the mechanism described in Scheme 1.10, the deprotonation occurs on the DPPcompound 9. It is also possible to brominate the DPP and deprotonate the benzaldehyde instead. The possibility for ortho-addition on benzalehyde is present, and this method is possibly a convinient solution for polymerization. ${ }^{[90]}$ Direct arylation can also be used for combining the phenothiazine-donor 2 and the DPP $\pi$-bridge $\mathbf{1 3}$ from this reaction.



Scheme 1.10: Proposed mechanism for Pd-catalyzed direct arylation between dithiophene-DPP 9 and benzaldehyde to form the DPP-based aldehyde 13, with assistance of pivalic acid. ${ }^{[86-88]}$


## Results and Discussion

### 2.1 General

The aim of this master project was to synthesize the three novel dyes MEL1, MEL2 and MEL3, displayed in Figure 2.1. The syntheses of MEL1 and MEL2 are presented in Section 2.2.1-2.2.4, and MEL3 is covered in Section 2.2.52.2.9. Detailed information about the syntheses can be found in Section 5.2 Synthesis. The photophysical properties of the dyes in the MEL-series are presented in Section 2.3.


Figure 2.1: The target sensitizers synthesized during this master project.

### 2.2 Synthesis

### 2.2.1 Suzuki-Miyaura Cross-Coupling on Phenothiazine

Compound 2 was synthesized with a Suzuki-Miyaura cross-coupling between the brominated phenothiazine $\mathbf{1}$ and the commercially available pyrene-1-boronic acid, as shown in Figure 2.1. ${ }^{[91]}$ The catalyst-system described by Knapp et al. ${ }^{[92]}$ was used in this reaction.


Scheme 2.1: The phenothiazine compound 1 reacted with pyrene-1-boronic acid in a Suzuki cross-coupling to form product 2.

It was expected that the reaction would form three different products, mono-substituted 2, di-substituted $\mathbf{3}$ and the homocoupled pyrene, together with an appreciable amount of starting material 1. The statistically relative amounts of the unsubstituted, mono-substituted and di-substituted pyrene are respectively $1 / 2 / 1$, using one equivalent of pyrene-1-boronic acid. Higher amounts of the pyrene-1boronic acid can form more of the mono-substituted $\mathbf{2}$, but also the di-substituted 3. The reaction was run for 6 hours with 1.1 equivalents of pyrene-1-boronic acid, and all of the mentioned expected compounds were observed. The main challenge with this reaction is the purification, due to the equal polarity of the three phenothiazine-containing compounds, mainly caused by the triethylene oxide methyl ether (TEOME) chain. The $\mathrm{Et}_{2} \mathrm{O}$ eluent-system showed best separation, with $\mathrm{R}_{f}$-values of $0.24,0.29$ and 0.35 . The donor building block $\mathbf{2}$ was synthesized in a yield of $54 \%$, which is a significant improvement from the $19 \%$ yield obtained in the specialization project. ${ }^{[91]}$ The high yield can be due to a more ideal amount of pyrene-1-boronic acid and an careful purification process with a large total load of silica on a relatively small amount of crude material.

### 2.2.2 Borylation of Phenothiazine

A boronate ester was introduced to the phenothiazine compound 2 by a Pd-catalyzed borylation to give compound 4 . The conditions used were inspired by Murata et al., ${ }^{[93]}$ but with another catalyst-system. The reaction is presented in Scheme 2.2.



Scheme 2.2: Borylation of the donor molecule 2 to the boronate-functionalized compound 4.

The borylation reaction was fast, as it was stirred for only 3 hours. Due to the instability on silica, the crude was only semi-purified with a celite plug. ${ }^{1} \mathrm{H}$ NMR indicated that pinacolborane was the only impurity with $\delta_{H} 7.92$ and 1.17 ppm , and $\delta_{C} 81.8$ and 24.9 ppm . The borylated phenothiazine 4 was carried out in a $94 \%$ yield (included the pinacolborane).

### 2.2.3 Suzuki-Miyaura Cross-Coupling Between Phenothiazine and Thiophene Aldehydes

The Suzuki-Miyaura cross-coupling between phenothiazine $\mathbf{4}$ and thiophene aldehydes was carried out with the same conditions as previously described in Section 2.2.1. The reactions are presented in Scheme 2.3.


Scheme 2.3: Suzuki-Miyaura cross-coupling of the borylated compound 4 to form the donor- $\pi$-bridge systems 5 and $\mathbf{6}$.

Both reactions to form the products 5 and $\mathbf{6}$ were run at $80^{\circ} \mathrm{C}$ for 18 and five hours, respectively. This prepared the products 5 and $\mathbf{6}$ with a large difference in yield, $93 \%$ and $55 \%$, respectively. The main inequality between the two reactions was the purification with silica gel chromatography. For compound 6 an EtOAc eluent was used, and an $1: 1 \mathrm{Et}_{2} \mathrm{O}: \mathrm{EtOAc}$ eluent-system for 5 . TLC did not show a better separation for the $\mathrm{Et}_{2} \mathrm{O}: \mathrm{EtOAc}$ eluent-system than for EtOAc . It is still possible that the $\mathrm{Et}_{2} \mathrm{O}$ is a better eluent, due to the ether chains (TEOME) in the compounds 4,5 and 6 .

### 2.2.4 Knoevenagel Condensation to the Dyes MEL1 and MEL2

The Knoevenagel condensations displayed in Scheme 2.4 were used to derive the two target dyes MEL1 and MEL2.





Scheme 2.4: Knoevenagel condensations to yield the target sensitizers MEL1 and MEL2.

The Knoevenagel condensation is usually the last step in the synthesis of dyes, and it is expected to show high degree of conversion from this reactions. ${ }^{[74]} \mathrm{A}$ change in color from orange to red occured only minutes after the reactions were started. After 2 hours at reflux, both reactions showed full conversion, which was detected by TLC. The purification was performed with silica gel chromatography column with a gradient eluent system ( $\mathrm{DCM}-\mathrm{DCM}: \mathrm{MeOH}$ ). The products were not eluted in the DCM eluent, which makes all the impurities leave the column before addition of MeOH and following elution of the dyes MEL1 and MEL2. The MEL1 sensitizer was carried out from the Knoevenagel condensation in 94\% yield, while the MEL2 was synthesized in a yield of $72 \%$.

### 2.2.5 Alkylation of Diketopyrrolopyrrole

In order to try other $\pi$-bridges, the diketopyrrolopyrrole (DPP) unit with thiophenes 7 was used. This extended $\pi$-conjugated bridge can hopefully reduce the band gap energy and increase the molar extinction coefficient, in order to get more efficient dyes. The absorption maxima is also expected to be more red-shifted for this dye than for the MEL1 and MEL2 dyes. This can result in adsorption of other parts of the visible-wavelengths. The incorporation of the DPP-unit can be done on similar dyes as the previous described MEL1 and MEL2, as shown in Scheme 2.5.


Scheme 2.5: The dye MEL3 can be synthesized from phenothiazine 4 and DPP 9.

The procedure for the amide alkylation was inspired by Vakuliuk et al. ${ }^{[94]}$ The reaction was promising, as it had a yield of $79 \%$. The method used the commercially available DPP 7 and potassium carbonate in DMF. They purified the product with re-crystallization. Additionally, the method employed $\mathrm{Bu}_{4} \mathrm{~N}^{+} \mathrm{HSO}_{4}^{-}$and the alkylated agent was added in one portion. This differs from most of the other reaction done with this particularly system. ${ }^{[95-97]}$ The reaction conditions are shown in Scheme 2.6.

Table 2.1 displays some relative amounts of mono-alkylated and $\mathrm{N}, \mathrm{O}$-dialkylated DPP 9 in the crude material, which were detected by ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ). The un-alkylated compound 7 is not mentioned due to its insolubility in $\mathrm{CDCl}_{3}$. The $O, O$-dialkylated $\mathbf{1 1}$ is also left out, due to only trace amounts were detected.


Scheme 2.6: Reaction conditions for amide alkylation of DPP 7 to form the alkylated derivate 9 .

Four out of eleven reactions (Entry 1-4), shown in Table 2.1, were performed with $\mathrm{Bu}_{4} \mathrm{~N}^{+} \mathrm{HSO}_{4}^{-}$present. No clear difference was seen between the reactions with or without this phase transfer catalyst. The reaction time was varied between 3-25 hours, where the majority were between 20-25 hours. The experiment with reaction time of 3 hours (Entry 1) had a higher amount of the mono-N-alkylated product than the reactions with a longer reaction-time. The amount of $\mathrm{N}, \mathrm{O}$-dialkylated 10 was more or less the same as for the reaction with a longer reaction time. Therefore, the reaction time does not seem to affect the amount of $\mathrm{N}, \mathrm{O}$-dialkylated $\mathbf{1 0}$ equally as much. No significant difference was found between the 14 hour reaction (Entry 10) and the majority of reactions at 20-25 hours. This indicates that a reaction-time of approximately 14 hours or longer is mandatory for low yield of mono-alkylated DPP 10, hence higher yield of $\mathrm{N}, \mathrm{N}$-dialkylated 9 .

Some variation of the temperature was also tested. An increase in temperature from $120^{\circ} \mathrm{C}$ to $140^{\circ} \mathrm{C}$ decreased the amount of $\mathrm{N}, \mathrm{O}$-dialkylated 10 from an average of $18 \%$ to $3 \%$. The difference from $140^{\circ} \mathrm{C}$ to $155^{\circ} \mathrm{C}$ was not significant. This indicates that a high temperature, $140^{\circ} \mathrm{C}$, is required for $N-N$ selection of the alkylation, but increasing above $140^{\circ} \mathrm{C}$ would most likely have no effect.

The purification methods used were crystallization/re-crystallization or silica gel chromatography, as showed in Table 2.1. Some of the reactions were purified in many steps, but only the first step is presented in the table. The method described initially in this section suggested re-crystallization, with precipitation from cold DMF and re-crystallization with DCM and hexane. This was done for a reaction of 750 mg (Entry 2), in a yield of $11 \%$. This purification method was supposed to be used for another reaction of 100 mg scale (Entry 5), but the product was

Table 2.1: Summary of alkylation reactions and purification. Hx is short for hexane. Purification and yields in this table are the first purification step.

| Entry | Scale <br> [mg] | Heat <br> $\left[{ }^{\circ} \mathrm{C}\right]$ | Time [h] | $\left\lvert\, \begin{aligned} & \text { Crude [\%] } \\ & \mathbf{8}^{a} \\ & \mathbf{1 0}^{b}\end{aligned}\right.$ |  | Purification |  | Yield [\%] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  | Cryst ${ }^{\text {c }}$ | Chrom. ${ }^{d}$ |  |
| 1 | 100 | 120 | 3 | 27 | 18 | - | $\underset{7: 3}{\mathrm{CHCl}_{3}}: \mathrm{Hx}$ | 11 |
| 2 | 750 | 120 | 25 | 2 | 22 | $\begin{gathered} \text { DMF } \\ (\mathrm{DCM} / \mathrm{Hx}) \end{gathered}$ | - | 9 |
| 3 | 150 | 120 | 25 | 4 | 19 | (DCM/Hx) | - | - |
| 4 | 100 | 120 | 24 | 7 | 20 | - | - | - |
| 5 | 100 | 120 | 24 | 6 | 18 | DMF | - | 31 |
| 6 | 100 | 120 | 24 | 7 | 9 | - | $\begin{gathered} \mathrm{Hx}: \underset{9: 1}{\mathrm{EtOAc}} \\ \hline \end{gathered}$ | 46 |
| 7 | 100 | 140 | 24 | 7 | 5 | - | - | - |
| 8 | 1000 | 140 | 24 | 6 | 4 | DMF | - | 31 |
| 9 | 1000 | 140 | 21 | 6 | 3 | - | $\begin{gathered} \mathrm{Hx}: E t O A c \\ 9: 1 \end{gathered}$ | 10 |
| 10 | 2000 | 140 | 14 | 4 | 2 | DMF | , | 43 |
| 11 | 2000 | 155 | 20 | 7 | 3 | DMF | - | 46 |

${ }^{a} N$-monoalkylated 8.
${ }^{b} \mathrm{~N}, \mathrm{O}$-dialkylated $\mathbf{1 0}$.
${ }^{c}$ Crystallization (re-crystallization).
${ }^{d}$ Silica gel chromatography column.
pure after the precipitation from cold DMF, which was determined with ${ }^{1} \mathrm{H}$ NMR. This increased the yield to $31 \%$. The exact same yield was obtained for a 1000 mg scale (Entry 8) using the same crystallization conditions. Some more product could probably be isolated, due to 'bumping' while evaporation in vacuo. Two more reactions were purified with DMF crystallization (Entry 10, 11), which gave the pure $N, N$-dialkylated DPP 9 in a $43 \%$ and $46 \%$ yield. This indicates that crystallization is easier with a larger scale, which was expected. Only the $N, N$-dialkylated DPP 9 precipitated from cold DMF, meaning that crystallization is suficient purification for this particular system, and there is no need for re-crystallization.

The product 9 was isolated by silica gel chromatography following three of the reactions (Entry 1, 6, 9). The mono-alkylated compound $\mathbf{8}$ is effortless to sep-
arate from the di-alkylated compounds, due to its hydrophilic properties. 7:3 $\mathrm{CHCl}_{3}$ :hexane and 9:1 hexane:EtOAc were used as eluent-systems. The reaction with eluent system $\mathrm{CHCl}_{3}$ :hexane (Entry 1) had a large amount of mono-alkylated and $N, O$-dialkylated 10 in the crude material, and therefore only $11 \%$ was isolated of $N, N$-dialkylated product 9 . The two other reactions in a 100 mg (Entry 6) and 1000 mg scale (Entry 9) which were purified by silica gel chromatography with eluent-system hexane:EtOAc had a yield of $46 \%$ (39 g silica) and $10 \%$ ( 102 g silica), respectively. ${ }^{1} \mathrm{H}$ NMR of the crude materials showed $16 \%$ of monoand $\mathrm{N}, \mathrm{O}$-dialkylated of the 100 mg scale reaction, and $9 \%$ for the 1000 mg scale. This argues for chromatography as purification method for relatively small scale reactions, and precipitation by crystallization for large scales.

One of the 2 g scales (Entry 10) had a crude containing 4\% monoalkylated and $2 \% \mathrm{~N}, \mathrm{O}$-alkylated. This reaction was first purified with crystallization yielding $43 \% N, N$-alkylated 9 , as previous described. The mother-liquor was purified with chromatography ( 104 g silica, hexane:EtOAc, 9:1), which resulted in a total yield of $52 \%$ of the $N, N$-dialkylated DPP 9 .

It has been reported that placing the branching point away from the nitrogen on DPP has increased the solubility compared to a linear alkyl, while maintaining their electronic properties. ${ }^{[98,99]}$ This can reduce some of the solubility challenges, and may improve the alkylation and following reactions and purifications.

## Formation of Mono-Alkylated Relative to $N, N$-Alkylated

An alkylation experiment with $120{ }^{\circ} \mathrm{C}$ was run in order to get a better under-


Figure 2.2: Formation of mono-alkylated $\mathbf{8}$ and $\mathrm{N}, \mathrm{O}$-alkylated $\mathbf{1 0}$ relative to $\mathrm{N}, \mathrm{N}$ alkylated 9 .
standing of the time-dependence formation of the N -monoalkylated $\mathbf{8}$ and $\mathrm{N}, \mathrm{O}$ dialkylated 10 relative to the $\mathrm{N}, \mathrm{N}$-dialkylated 9, as displayed in Figure 2.2. Samples from the reaction were analyzed with ${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$.

As mentioned previously, the starting material, non alkylated 7, is not soluble in chloroform, and was therefore not detected in the ${ }^{1} \mathrm{H}$ NMR-analysis. After only one minutes, there was formation of both mono- $\mathbf{8}$ and $N, N$-alkylated DPP 9, with slightly more of the mono-alkylated molecule $\mathbf{8}$. The $N, N$-dialkylated product 9 continues rapidly formation after the first minute, in line with the decreasing amount of mono-alkylated compound $\mathbf{8}$. The formation of $\mathrm{N}, \mathrm{O}$-dialkylated $\mathbf{1 0}$ follows the same trend as shown for $\mathrm{N}, \mathrm{N}$-dialkylated $\mathbf{9}$, but in smaller quantities.

### 2.2.6 NBS Bromination of Diketopyrrolopyrrole

The procedure for bromination of DPP was inspired by Sun et al. ${ }^{[100]}$ The method uses NBS in $\mathrm{CHCl}_{3}$ and acetic acid, as shown in Scheme 2.7 They carried out the brominated reaction in $77 \%$ yield.



Scheme 2.7: NBS bromination of the DPP compound 9 to afford the brominated derivative 12.

As the bromination is an electrophilic aromatic substitution (EAS) reaction, and sulfur (an electron donating atom) is attached to the aromatic, is it expected that the addition of bromine will occur in the position next to the sulfur, as shown in Scheme 2.7. Bromination in this position was obtained and no other brominated derivatives were observed. The reaction was followed with ${ }^{1} \mathrm{H}$ NMR, that revealed the importance of long reaction time, 15 hours, to achieve full conversion. The reaction was run three times at room temperature in the dark, in scales of 200 $\mathrm{mg}, 500 \mathrm{mg}$ and 700 mg , and the crude was not purified for any of the reactions. The yield of the di-brominated product $\mathbf{1 2}$ was assumed $99 \%$ in all reaction. ${ }^{1} \mathrm{H}$ NMR-analysis predicted more than $98 \%$ pure product.

### 2.2.7 Suzuki-Miyaura Cross-Coupling on the DPP-Unit

The introduction of bromine in the previous mentioned reaction was done in order to react the product $\mathbf{1 2}$ further in a Suzuki-Miyaura cross coupling, to form the monosubstituted 14. Six different experiments are summarized in Table 2.2. This table shows a variation of reaction time, catalyst, ligand and their relative amounts, analyzed with ${ }^{1} \mathrm{H}$ NMR. Notice that the table referres to the dehalogenated product 13, and not target product 14. The fixed conditions are shown in Scheme 2.8.


Scheme 2.8: Suzuki-Miyaura cross coupling on the dithiophene DPP 12 in an attempt to form the DPP-based aldehyde 14.

The first experiment (Entry 1) with $\mathrm{Pd}(\mathrm{OAc})_{2}$ and SPhos showed no conversion after 9 hours. In an attempt to get the disubstituted product 14, an extra load of 4-formylphenylboronic acid (total 3 eq.) and catalyst/ligand were added to the reaction. After a total of 26 hours, $8 \%$ of both mono- and di-substituted product were formed, in addition to $23 \%$ of an unknown by-product. An experiment with the same start conditions was performed (Entry 2), to exclude the opportunity of human inaccuracy. This experiment had no conversion after 17 hours, and another reaction with a larger amount of catalyst and ligand was run (Entry 3). This resulted in a $6 \%$ conversion to the monosubstututed product 13 and $17 \%$ to the disubstituted 15, and only trace amounts of the unknown by-product. In an attempt

## Chapter 2. Results and Discussion

Table 2.2: Summary of Suzuki cross coupling experiments on DPP, with varied catalyst and ligand. One unknown compound was also present, but is not included in this table.

| Entry | Time [h] | Catalyst | (Eq.) | Ligand | (Eq.) | $\left\lvert\, \begin{gathered} \mathbf{S}^{a} \\ {[\%]} \end{gathered}\right.$ |  | $\begin{gathered} 15 \\ {[\%]} \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 26 | $\mathrm{Pd}(\mathrm{OAc})$ | (0.06) | SPhos | (0.10) | 61 | 8 | 8 |
| 2 | 17 | $\mathrm{Pd}(\mathrm{OAc})$ | (0.03) | SPhos | (0.06) | 100 | 0 | 0 |
| 3 | 17 | $\mathrm{Pd}(\mathrm{OAc})$ | (0.04) | SPhos | (0.08) | 76 | 6 | 17 |
| 4 | 1 | $\mathrm{Pd}(\mathrm{OAc})$ | (0.04) | XPhos | (0.08) | 70 | 18 | 10 |
| 5 | 48 | Pd (dppf) $\mathrm{Cl}_{2}$ | (0.06) |  | - | 63 | 0 | 0 |
| 6 | 5 | $\mathrm{Pd}\left(\mathrm{PPh}_{3}\right)_{4}$ | (0.03) | - | - | 94 | 0 | 6 |

${ }^{a}$ Starting material, 4-formylphenylboronic acid. Detected with ${ }^{1} \mathrm{H}$ NMRanalysis.
to decrease the amount of the disubstituted $\mathbf{1 5}$, an experiment with with a reaction time of only one hour and another ligand, XPhos, was performed (Entry 4). A larger amount of the monosubstituted $\mathbf{1 3}$ was obtained, $18 \%$ out of a conversion of $30 \%$.

All the described reactions above were run with the same catalyst. A reaction with the highly active palladium-catalyst $\mathrm{Pd}(\mathrm{dppf}) \mathrm{Cl}_{2}$ showed no formation of either mono- or di-substituted product, even at a long reaction time of 48 hours. The catalyst $\mathrm{Pd}\left(\mathrm{PPh}_{3}\right)_{4}$ has been used on a similar compound, ${ }^{[101]}$ but resulted in $6 \%$ conversion to the disubstituted compound.

None of the previous mentioned reaction showed the wanted halogenated product 14, but instead the reduced product 13. This indicates that the oxidative addition step is active. The reduction can occur from the unstable Aryl-Pd-R complex in the metathesis step, as mentioned in section 1.3.6.

The Suzuki-Miyaura cross-coupling was also used as a tool to bind the DPP unit $\mathbf{1 2}$ with the phenothiazine donor-group 4, as shown in Scheme 2.9.


Scheme 2.9: Suzuki-Miyaura cross-coupling between DPP 12 and phenothiazine 4 in an attempt to form product 17.

During the reaction the color changed from purple to blue-purple, which is a normal color for the mono-substituted dithiophenyl-DPP. ${ }^{[6]}$ The crude material showed seven different spots on TLC. After four rounds of silica gel chromatography columns there was only a small amount of material left and still three spots on TLC. The same observation as for the previous described Suzuki on DPP was also seen in this reaction, the dehalogenated compound $\mathbf{1 6}$ was formed. This indicates that the Suzuki-Miyaura cross coupling on dithiophene-DPP compounds is challenging.

A change of methodology had to be done in order to synthesize target molecule MEL3 with respectable yields. Both Stille cross-coupling ${ }^{[102,103]}$ and direct arylation ${ }^{[104,105]}$ on dithiophene-DPP-units have been described in the literature. Due to the highly toxic organotin agent used in Stille, ${ }^{[86]}$ the direct arylation was the first choice.

### 2.2.8 Direct Arylation of the DPP-Unit

The procedure for the Pd-catalyzed direct arylation of the DPP compound was inspired by Bohra et al. ${ }^{[106]}$ The method uses pivalic acid and $\mathrm{K}_{2} \mathrm{CO}_{3}$, as presented in Scheme 2.10

The DPP 9 was reacted in excess ( 1.2 eq .), to prevent di-substitution by the benzaldehyde. After 14 hours at $120{ }^{\circ} \mathrm{C}$, the ratio between the mono-substituted $\mathbf{1 3}$ and the di-substituted DPP $\mathbf{1 5}$ of the crude material was $3: 1$, respectively. This means that an even larger excess of DPP is necessary to inhibit the di-substitution.

The DPP unit is as previous mentioned poorly soluble in many organic solvents. Therefore, only a limited selection of eluents are possible. This makes the purification with gel column chromatography challenging, and resulted in four columns where some product was isolated each time. The first column (Hexane/EtOAc/DCM)


Scheme 2.10: Direct arylation between the DPP-unit 9 and benzaldehyde, to yield the DPP-based aldehyde 13.
was performed to remove the starting material. The two aldehyde compounds were not eluted before addition of DCM. In theory this would result in isolation of all starting material. Unfortunately, the starting material was not soluble in the eluentsystem, which resulted in DPP starting material 9 in all collected fractions. However, the largest amount of starting material was isolated during this column. The following three columns were able to isolate some of the three compounds each time. This direct alkylation gave the mono-substituted DPP-benzaldehyde $\mathbf{1 3}$ in a yield of $68 \%$ with high purity.

The exact same reaction conditions were used to couple the phenothiazine donor group 2 and the product $\mathbf{1 3}$ from the previous described reaction, as shown in Scheme 2.12.


Scheme 2.11: The DPP-based aldehyde 13 was bound to the phenothiazine donor compound 2 in a direct arylation to form compound 18.

After 12 hours at $120^{\circ} \mathrm{C}$ the color changed from blue-purple to blue. The likely unfortunate by-product of this reaction could be the coupling of phenothiazine to a pyrene-group on another donor compound. This could result in the wanted product 18, but with a double donor group couplet in series. After three silica gel chromatography columns, TLC-analysis indicated that there were still three compounds left. A ${ }^{1} \mathrm{H}$ NMR-analysis registered only one aldehyde peak, which is the functional-group who reacts in the next reaction, a Knoevenagel condensation.

The compound was therefore not further purified. The product $\mathbf{1 8}$ was synthesized in a $73 \%$ yield with a purity of $79 \%$. HRMS was inconclusive, possibly due to its size and solubility properties.

### 2.2.9 Knoevenagel Condensation to Dye MEL3

A Knoevenagel condensation gave the target dye MEL3, as shown in Scheme 2.12.


Scheme 2.12: Knoevenagel condensation to yield dye MEL3.

The starting material, 44.0 mg , was not soluble in the limited amount of acetonitrile. After one hour at reflux, additional solvent was added, but compound $\mathbf{1 8}$ was still not dissolved. Chloroform was added in a ratio of 1:3, ${ }^{[107]}$ resulting in completely dissolved mixture. The reaction was followed by TLC, and full conversion was achieved after seven hours. Silica gel chromatography column gave 46.4 mg assumed product MEL3.

NMR spectroscopy for this molecule MEL3 was inconclusive, meaning that HRMS and IR spectroscopy are the two most important analysis methods, in this case, for determining whether the dye is formed or not. The HRMS-analysis confirmed that the target dye MEL3 was synthesized. IR spectroscopy had all the expected peaks for the target MEL3, but also an extra adsorption band. This signal is from the $\mathrm{S}=\mathrm{O}$ stretch, indicating that the molecule is oxidized. Oxidation of the sulfur on phenothiazine S-5 have been seen during previous work in the research group at NTNU. There were also two other sulfur-atoms present on the thiophenes. The oxidized dye would have another molecular formula with corresponding molecular weight. The oxidized molecular weight was not found with HRMS-analysis, nor the dioxidized or trioxidized. It is therefore possible that there is more that differs between the by-product and the dye MEL3 than oxidation of sulfur.

In TLC-analysis, only one spot was seen. This indicates that the by-product is an acid, due to the large difference in polarity compared to if it was not an acid. In the attempts to get a decent NMR spectra, the product MEL3 was run in the
magnetic field of the NMR and solvents was removed in vacuo multiple times. This could have been unfortunate for the molecule, due to large exposure to light and air. The IR and MS analysis were both taken after this attempts of NMRanalysis. It is therefore not known whether the MEL3 has been synthesized pure and unfortunately reacted during the analysis, or if the product was impure after the Knoevenagel reaction and purification. Since HRMS did not work for the predecessor molecule, it is also possible that the by-product was formed during the direct arylation. However, the HRMS and IR together indicated that the target dye MEL3 is synthesized, but an unknown acid containing by-product is also present.

### 2.3 Photophysical Properties of the Dyes

This section presents UV-Vis absorption properties of sensitizers MEL1-3, AFB43 synthesized by Buene and the literature dye, Dyenamo Blue. All dyes are displayed in Figure 2.3. The obtained spectra for the the photophysical properties of the dyes are displayed in Figure 2.4, and summarized in Table 2.3.



Figure 2.3: The literature dye Dyenamo Blue, previously synthesized dye AFB43 by Buene and the dyes in the MEL-series.


Figure 2.4: Recorded UV-Vis adsorption spectra of the dyes MEL1-3 and AFB43 compared to the literature dye Dyenamo Blue.

The target dye MEL1 had an adsorption maxima at 482 nm with a corresponding molar extinction coefficient, $\varepsilon$, of $21401 \mathrm{M}^{-1} \mathrm{~cm}^{-1}$. The closely related dye MEL2 showed a maxima adsorption of 497 with the corresponding $\varepsilon$-value of 27 $838 \mathrm{M}^{-1} \mathrm{~cm}^{-1}$. Buenes dye AFB-43 produced a lower adsorption, 452 nm , than MEL1 and MEL2, but with a higher $\varepsilon$-value, $32262 \mathrm{M}^{-1} \mathrm{~cm}^{-1}$. The DPP-dye MEL3 had a more red-shifted adsorption spectra with a maxima of 622 nm and molar extinction coefficient of $18928 \mathrm{M}^{-1} \mathrm{~cm}^{-1}$. The literature dye Dyenamo Blue had a lower adsorption maxima, 589 nm , but significant higher $\varepsilon$-value, 55 $265 \mathrm{M}^{-1} \mathrm{~cm}^{-1}$, as shown in Table 2.3.

All the tested dyes had an adsorption maxima in the visible region, 400-700 nm. As displayed in Figure 2.4, two peaks are present for each dye. The peaks at 300370 are corresponding to the $\pi-\pi^{*}$ electronic transitions. The longer wavelengths at $380-600 \mathrm{~nm}$ are ascribed to the intramolecular charge transfer (ICT) from the donor to the acceptor. ${ }^{[108]}$ This ensures efficient charge-separation in the excited state.

As expected the sensitizers with DPP-based $\pi$-bridges had more red-shifted adsorption maxima than the dyes with thiophene-based $\pi$-bridges. The sensitizer

Table 2.3: Photophysical properties of dyes MEL1-3, AFB-43 and Dyenamo Blue.

| Dye | Adsorption <br> $\lambda_{\text {abs }}[\mathrm{nm}]^{a}$ | $\varepsilon\left[\mathrm{M}^{-1} \mathrm{~cm}^{-1}\right]^{b}$ | $\mathrm{E}_{0-0}[\mathrm{eV}]^{c}$ |
| :--- | :--- | :--- | :--- |
| MEL1 | 492 | 21401 | 2.08 |
| MEL2 | 497 | 27838 | 2.09 |
| MEL3 | 622 | 18928 | 1.81 |
| AFB-43 | 452 | 32262 | 2.29 |
| Dyenamo Blue | 589 | 55265 | 1.83 |

${ }^{a}$ Absorption maximum of the most red shifted peak, measured in $\mathrm{CHCl}_{3}\left(2 \cdot 10^{-5} \mathrm{M}\right)$.
${ }^{b}$ Molar extinction coefficient.
${ }^{c}$ Band gap energy.

AFB-43 has one thiophene and one phenyl, that blue-shifts the adsorption maxima. This can be due to the larger dihedral angel of the phenyl-thiophene bond, which decreases the orbital overlap. ${ }^{[109]}$ This observation can be related to the phenyl in MEL3, and an even more red-shifted adsorption maxima could possibly occur if the phenyl was not present or replaced by a five membered heterocycle. A research done on phenyl-DPP based dyes looked at the difference of attaching the three aromatic compounds furan, phenyl and thiophene. ${ }^{[110]}$ The combination of phenyl-DPP and thiophene showed the lowest conversion efficiency of all the three dyes. This supports that the phenyl-thiophene bond is not well suited for dyes to be used in DSSCs.

The band gap energy, $\mathrm{E}_{0-0}$, was calculated according to the equation in Section 1.2.4: Energic Consideration of Dyes in DSSC. The result of the estimated band gap energy showed the same trend as previously described for adsorption, due to the dependence of these two values. The only exception was for the dyes MEL1 and MEL2, where MEL1 has a lower band gap energy than MEL2, although MEL2 had a slightly more red-shifted adsorption maxima.

All the adsorption tested sensitizers showed a relatively high molar extinction coefficient, $\varepsilon$, of above $18000 \mathrm{M}^{-1} \mathrm{~cm}^{-1}$. The literature dye Dyenamo Blue had a significant higher $\varepsilon$-value than the rest. The molar extinction coefficient is a measure of how strongly a molecule can absorb light at a particular wavelength, and can be increased by extending the $\pi$-conjugation of the $\pi$-bridge. ${ }^{[111]} \mathrm{A}$ dye with good light absorption-properties, high $\varepsilon$-value, can have less dye-molecules attached to the $\mathrm{TiO}_{2}$. If fewer molecules are required for harvesting a certain amount of light energy, the thickness of the $\mathrm{TiO}_{2}$-layer can be reduced. ${ }^{[112]} \mathrm{A}$ thin $\mathrm{TiO}_{2}$-layer is desirable due to better electron and electrolyte diffusion through the semiconduc-
tor due to lower resistance, which lowers the charge recombination. ${ }^{[113]}$
Based on the photophysical properties of the dyes, the literature dye Dyenamo Blue is expected to outperform the dyes in the MEL-series, due to its red-shifted adsorption maxima and high molar extinction coefficient. The expected best dye to use in DSSCs of the MEL-series could be MEL3 due to its large red-shifted adsorption maxima. MEL3 has the lowest $\varepsilon$-value, but still higher than the literature dye N719, which is around $13000 \mathrm{M}^{-1} \mathrm{~cm}^{-1}$.

The product MEL3 is assumed to be impure, as discussed in Section 2.2.9: Knoevenagel Condensation to Dye MEL3. This can also be seen in Figure 2.4, as a double signal. It was used a 0.02 mM solution of the dyes to measure the photophysical properties, but if the product-mixture contains more than one molecule, this will result in a smaller signal with a lower extinction coefficient. This indicates that the dye MEL3 is even more suited as a dye to use in DSSCs than expected from the measurements presented in this section.

None of these dyes cover large parts of the solar spectrum, displayed in Figure 2.5. ${ }^{[114]}$ The $\mathrm{TiO}_{2}$ can be co-sensitized with two different dyes to obtain a broad spectral response in the visible wavelengths. ${ }^{[115]}$ The co-sensitization can also reduce charge recombination. ${ }^{[116]}$


Figure 2.5: Solar spectrum at sea level and outside atmosphere. The solar radiation that reaches the earth contains most visible and infrared light. ${ }^{[114]}$

As presented in Figure 2.5, most energy can be harvested at wavelengths of around 500 nm . The dye MEL2 has an absorption maxima at 497 nm , which is close to
the maxima of the solar spectra. The adsorption around 600 nm is close to zero, and in order to get a wider light absorption range the dyes MEL2 and MEL3 can be co-sensitized. The MEL3 related dye Dyenamo Blue has been co-sensitized with dye D35. In this research the Dyenamo Blue had an efficiency of $5.6 \%$, and increased to $7.3 \%$ when co-sensitizing with D35 in a ratio of 1:1. ${ }^{[117]}$ In another study the two same dyes were used with a ratio of $3: 4$ and resulted in an efficiency of $10.7 \%$. ${ }^{[118]}$ Due to assumed impurities in MEL3, the relative amounts of MEL2 and MEL3 for co-sensitization will not be discussed.

The other two dyes MEL1 and AFB-43 could also be used for co-sensitization with MEL3. MEL1 has a lower extinction coefficient and slightly lower adsorption maxima, and will probably be outperformed by MEL2. The dye AFB-43 has a significantly lower adsorption maxima and this dye combined with the MEL3 dye will have a low adsorption around 530 nm , as displayed in Figure 2.4. The MEL2 and MEL3 dyes are assumed to perform best as co-sensitizers.

## Structure Elucidation

By investigation of ${ }^{1} \mathrm{H},{ }^{13} \mathrm{C}$, COSY, HSQC and HMBC NMR spectra, IR and HRMS, the identities of the synthesized compounds were verified. The dyes MEL1, MEL2 and MEL3 were also characterized using UV-Vis spectroscopy. The spectroscopic data were analyzed using the literature by Silverstein et al. ${ }^{[119]}$

### 3.1 NMR

Figure 3.1 shows an overview of some connectivities used to elucidate the structure of new compounds. Assigned ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ are presented in Table 3.1-3.8, while NMR spectra used in stucture elucidation are given in Appendix under each specific compound. Most of the molecules have also been substantiated by MS and have been analyzed by IR spectroscopy.

All spectra contain residual signals from the used deuterated solvent. These, and signals remaining from solvents used in the synthesis were identified using litterature by Fulmer et al. ${ }^{[120]}$ The solvent signals are not disscussed further. If other significant impurities are presented in the spectra, they will be commented.


Figure 3.1: An overview of some COSY and HMBC connectivities used to elucidate the structure of new compounds.

For compounds containing pyrene, quaternary carbons in pyrene will not be assigned. No further reaction will be done directly on the pyrene group, and structure elucidation of pyrene is not crucial for further work with pyrene containing compounds. In order to identify different carbons and hydrogen of pyrene, the 2-D INADEQUATE NMR experiment can be used. ${ }^{[121]}$ The method is useful for determining which signals arise from neighboring carbons. This method is mainly used in case of compounds with similar structures, such as pyrene. ${ }^{[122]}$ The disadvantage of this experiment is the insensitivity, due to the low natural abundance of the NMR active ${ }^{13} \mathrm{C}$ isotope.

### 3.2 IR

IR spectra can be found in Appendix under each specific compound. Some bonds in the IR spectra are characteristic and shared for many of the synthesized molecules. All molecules have shown a weak absorption around $3200-2700 \mathrm{~cm}^{-1}$ resulting from aromatic C-H stretching. Aromatic C-H in-plane bending appear around $1300-1000 \mathrm{~cm}^{-1}$ and out-of-plane around $900-675 \mathrm{~cm}^{-1}$. Other absorption bonds will be presented under each specific compound.

## Chapter 3. Structure Elucidation

### 3.3 Spectroscopic Identification of 4

NMR, IR and HRMS analysis of compound $\mathbf{4}$ is shown in Appendix C: Spectroscopic Data - Compound 4. HRMS concluded molecular formula $\mathrm{C}_{47} \mathrm{H}_{47} \mathrm{BNO}_{6} \mathrm{~S}$ with exact mass 764.3217, which corresponded with the calculated mass at 764.7533.

Some characteristic signals of the functional groups on 4 were displayed in the IR spectrum. The peak 1140 is consistent with symmetric and asymmetric C-O-C streching of aliphatic ether.

The assigned ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR signals of 4 are shown in Figure 3.2 and Table 3.1. This section contains a walk-through of the structure elucidation of the NMR analysis of compound $\mathbf{4}$, apart from the quaternary carbon in the pyrene group.


Figure 3.2: Assigned protons and carbons for compound 4.

Table 3.1: ${ }^{1} \mathrm{H}(600 \mathrm{MHz})$ and ${ }^{13} \mathrm{C}$ NMR ( 150 MHz ) shifts for compound 4 (DMSO- $\mathrm{d}_{6}$ ). The position of carbons and protons are shown in Figure 3.2.

| Position | ${ }^{13} \mathrm{C}$ NMR [ppm] | ${ }^{1} \mathrm{H}$ NMR [ppm] |
| :---: | :---: | :---: |
| 1 | 116.2 | 6.30 |
| 2 | 129.7 | 7.19 |
| 3 | 135.3 | - |
| 4 | 128.3 | 7.30-7.26 |
| 4 a | 119.4 | - |
| 5a | 118.0 | - |
| 6 | 132.8 | 7.26 |
| 7 | 122.7 | - |
| 8 | 134.4 | 7.23 |
| 9 | 115.4 | 6.19 |
| 9 a | 146.7 | - |
| 10a | 143.1 | - |
| 11 | 132.5 | - |
| 12 | 132.3 (2C) | 7.45 (2H) |
| 13 | 117.3 (2C) | 7.30 (2H) |
| 14 | 158.9 |  |
| 15 | 67.9 | $4.23(2 \mathrm{H})$ |
| 16 | 69.3 | 3.81 (2H) |
| 17 | 70.4 | 3.63 (2H) |
| 18 | 70.3 | 3.58-3.56 (2H) |
| 19 | 70.1 | 3.55-3.53 (2H) |
| 20 | 71.7 | 3.45-3.44 (2H) |
| 21 | 58.5 | 3.24 (3H) |
| 22 | 124.9 | 8.12 |
| 23 | 128.0 | 8.18 |
| 24 | 127.8 (2C) | $8.21(2 \mathrm{H})$ |
| 25 | 125.4 | 8.29 |
| 26 | 126.9 | 8.09 |
| 27 | 128.1 | 8.33-8.31 |
| 28 | 127.9 | 7.97 |
| 29 | 125.5 | 8.33-8.31 |
| Pyr | 136.1 | - |
| Pyr | 131.4 | - |
| Pyr | 130.8 | - |
| Pyr | 130.5 | - |
| Pyr | 125.8 | - |
| Pyr | 124.5 | - |
| Pyr | 124.6 | - |
| 30 | 84.0 | - |
| 31 | 25.1 | 1.27 |

## Chapter 3. Structure Elucidation

The ${ }^{1} \mathrm{H}$ NMR singlet at 3.24 ppm , with intensity 3 , was produced by the aliphatic ether-proton $\mathrm{H}-21$. The protons did not couple to any other protons, as shown in Figure 3.3. As the figure displays, protons with chemical shift $3.45-3.44 \mathrm{ppm}$ $(\mathrm{H}-20)$ and $3.55-3.53 \mathrm{ppm}(\mathrm{H}-19)$ couple with each other (green square). The same is observed for protons $\mathrm{H}-18$ (3.58-3.56 ppm) and $\mathrm{H}-17$ ( 3.63 ppm ) (red square), and H-16 (3.81 ppm) and H-15 (4.23 ppm) (blue square).


Figure 3.3: COSY of aliphatic ether chain in compound 4.

As displayed in Figure 3.4, proton $\mathrm{H}-21$ belongs to carbon with signal 58.5 ppm , $\mathrm{H}-20$ to carbon at 71.7, and further on. This will not be explained for any other of the connecting protons and carbons in compound 4.


Figure 3.4: HSQC of aliphatic ether chain in compound 4.

From COSY NMR spectra the ethylene-fragments and the one $\mathrm{CH}_{3}$ group have been identified. Combining COSY and HMBC makes it possible to determine the order of the aliphatic ether chain, as shown in Figure 3.5, where coupling between carbon and neighboring protons are displayed.


Figure 3.5: HMBC of aliphatic ether chain in compound 4. Only one H shown where two or three H in the same position are equivalent.

## Chapter 3. Structure Elucidation

The blue arrow in Figure 3.5 shows the coupling between $\mathrm{H}-15$ and $\mathrm{C}-14$, which is displayed in the HMBC NMR spectra in Appendix. Both $\mathrm{H}-12$ and $\mathrm{H}-13$ couple with $\mathrm{C}-14$, as shown in Figure 3.6, but $\mathrm{H}-12$ has a stronger correlation, which is normal for the meta position in phenyl compounds.


Figure 3.6: HMBC coupling between $\mathrm{H}-12, \mathrm{H}-13$ and $\mathrm{C}-14$ and ${ }^{1} \mathrm{H}$ NMR of $\mathrm{H}-12$ and $\mathrm{H}-13$ in compound 4.

As shown in Figure 3.6 the $\mathrm{H}-12$ and $\mathrm{H}-13$ are pointing at each other, which is an indication that they couple with each other. The use of coupling constants is preferable, but due to overlap in the ${ }^{1} \mathrm{H}$ NMR spectrum, the accurate coupling constants for $\mathrm{H}-13$ could not be found. $\mathrm{C}-11$ is coupling with both $\mathrm{H}-12$ and $\mathrm{H}-13$, but strongest to $\mathrm{H}-13$. As expected, no correlation between $\mathrm{C}-11$ and the phenothiazine proton is observed. H-4 is coupling with a carbon of chemical shift of 136.1 ppm , which is one of the pyrene-carbons, as displayed in Figure 3.7. This indicates that H-4 is on the left side of phenothiazine. Coupling between H-2 and


Figure 3.7: Proton-proton coupling between $\mathrm{H}-1$ and $\mathrm{H}-2$ (COSY) and protoncarbon coupling between $\mathrm{H}-4$ and a pyrene-carbon and between $\mathrm{H}-2$ and $\mathrm{C}-4$ (HMBC) in compound 4.

C-4 is suggested in HMBC NMR spectra, and further coupling between $\mathrm{H}-2$ and $\mathrm{H}-1$ is seen in COSY NMR spectra.

To verify the suggested structure displayed in Figure 3.7 the multiplicities and their coupling constants can be used for assigning H-1, H-2 and H-4. H-2 is a doublet of doublets, and has coupling-constants 8.5 Hz and 2.1 Hz , as shown in Figure 3.8. $\mathrm{H}-1$ is a doublet with coupling-constant 8.42 , which corresponds to the couplingconstant of H-2. The other coupling-constant of $\mathrm{H}-2$ most likely corresponds to $\mathrm{H}-$ 4, but the coupling-constant of H-4 can not be determined, due to the overlapping with the signal of $\mathrm{H}-13$.


Figure 3.8: ${ }^{1} \mathrm{H}$ NMR of $\mathrm{H}-1$ and $\mathrm{H}-2$ signals in compound 4.

The quaternary carbons C-3, C-4a and C-10a were assigned using HMBC. All of them coupled with the protons in the left ring of phenothiazine. C-3 was coupling with $\mathrm{H}-29$ in the pyrene ring. C-10a was expected to have the highest chemical shift, due to having a nitrogen as the closest neighbor. The correlation between $\mathrm{C}-10 \mathrm{a}$ and the protons $\mathrm{H}-2$ and $\mathrm{H}-4$ was also stronger than between $\mathrm{C}-10 \mathrm{a}$ and $\mathrm{H}-1$ due to meta-coupling, ${ }^{[119]}$ as displayed in Figure 3.9. H-1 is in meta position with respect to C-4a, and a strong coupling between them is seen in the HMBC NMR spectra.


Figure 3.9: ${ }^{1} \mathrm{H}$ NMR of $\mathrm{H}-1$ and $\mathrm{H}-2$ signals in compound 4.

The same can be done for the right side of the phenothiazine, so this will not be explained any futher. Connected to the right side of phenothiazine is the pinacolboranegroup. H-31 had an intensity of 12 (3.4), and coupled to C-30.

For the pyrene-group, some fragments can be found using COSY NMR. The following fragments were found; $\mathrm{H}-22$ and $\mathrm{H}-23, \mathrm{H}-28$ and $\mathrm{H}-29, \mathrm{H}-26$ couplet to both H-25 and H-27, as shown in Figure 3.10 as red arrows. A singlet of intensity 2 was seen for the two protons $\mathrm{H}-24$, due to their identical environment. To connect all fragments HMBC NMR was used. The green arrows in Figure 3.10 give a summary of some of the couplings. A quaternary carbon with chemical shift 125.9 ppm coupled to both $\mathrm{H}-24$ and $\mathrm{H}-25$, which indicates that they are close to each other. The same observation was seen for the quaternary carbon with $\delta_{c} 124.9 \mathrm{ppm}$ to $\mathrm{H}-27$ and $\mathrm{H}-28$, and between the quaternary carbon with $\delta_{c} 136.1 \mathrm{ppm}$ to $\mathrm{H}-29$ and $\mathrm{H}-22$.


Figure 3.10: ${ }^{1} \mathrm{H}$ NMR of respectively $\mathrm{H}-1$ and $\mathrm{H}-2$ in compound 4.

### 3.4 Spectroscopic Identification of 5

NMR, IR and HRMS analysis of compound $\mathbf{5}$ is shown in Appendix D: Spectroscopic Data - Compound 5. HRMS predicted molecular formula $\mathrm{C}_{4} 6 \mathrm{H}_{3} 8 \mathrm{NO}_{5} \mathrm{~S}_{2}$ with exact mass 748.2191, the calculated mass is 747.9182 .

The IR spectrum in appendix displays some characteristic signals of the functional groups on compound $\mathbf{5}$. The peak 1140 is consistent with symmetric and asymmetric C-O-C streching of aliphatic ether. The strong signal on $1654 \mathrm{~cm}^{-1}$ correspond to $\mathrm{C}=\mathrm{O}$ stretching in aldehyde.

The assigned ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR signals of $\mathbf{5}$ is shown in Figure 3.11 and Table 3.2.


Figure 3.11: Assigned protons and carbons for compound 5.

Table 3.2: ${ }^{1} \mathrm{H}(600 \mathrm{MHz})$ and ${ }^{13} \mathrm{C}$ NMR ( 150 MHz ) shifts for compound 5 (DMSO- $\mathrm{d}_{6}$ ). The position of carbons and protons are shown in Figure 3.11.

| Position | ${ }^{13} \mathrm{C}$ NMR [ppm] | ${ }^{1} \mathrm{H}$ NMR [ppm] |
| :---: | :---: | :---: |
| 1 | 116.2 | 6.34 |
| 2 | 130.0 | 7.21 |
| 3 | 135.5 | - |
| 4 | 124.4 | 7.56 |
| 4 a | 118.8 | - |
| 5a | 119.9 | - |
| 6 | 128.5 | 7.34 |
| 7 | 127.4 | - |
| 8 | 126.1 | 7.39 |
| 9 | 116.3 | 6.21 |
| 9 a | 145.1 | - |
| 10a | 142.9 | - |
| 11 | 132.5 | - |
| 12 | 132.3 (2C) | 7.49 (2H) |
| 13 | 117.3 (2C) | 7.30 (2H) |
| 14 | 159.0 | - |
| 15 | 68.0 | $4.23(2 \mathrm{H})$ |
| 16 | 69.4 | 3.81 (2H) |
| 17 | 70.4 | 3.64-3.62 (2H) |
| 18 | 70.3 | 3.58-3.56 (2H) |
| 19 | 70.1 | 3.55-3.53 (2H) |
| 20 | 71.8 | $3.45-3.44(2 \mathrm{H})$ |
| 21 | 58.5 | 3.24 (3H) |
| 22 | 125.0 | 8.13 |
| 23 | 128.2 | 8.18 |
| 24 | 127.8 (2C) | $8.21(2 \mathrm{H})$ |
| 25 | 125.4 | 8.29 |
| 26 | 126.9 | 8.09 |
| 27 | 128.1 | 8.34-8.31 |
| 28 | 127.9 | 7.97 |
| 29 | 125.5 | 8.34-8.31 |
| Pyr | 136.1 | - |
| Pyr | 131.4 | - |
| Pyr | 130.9 | - |
| Pyr | 130.6 | - |
| Pyr | 125.8 | - |
| Pyr | 124.6 | - |
| Pyr | 124.5 | - |
| 30 | 152.1 |  |
| 31 | 124.8 | 7.63 |
| 32 | 139.9 | 7.99 |
| 33 | 141.5 | - |
| 34 | 184.2 | 9.86 |

### 3.5 Spectroscopic Identification of 6

NMR, IR and HRMS analysis of compound $\mathbf{6}$ is shown in Appendix E: Spectroscopic Data - Compound 6. HRMS analysis confirms product 6 with a molecular formula $\mathrm{C}_{50} \mathrm{H}_{39} \mathrm{NO}_{5} \mathrm{~S}_{3}$ with exact mass $830.2069\left([\mathrm{M}+\mathrm{H}]^{+}\right)$and calculated mass is 830.0418 .

The IR specra shows similar peaks as explained in Section 3.4. The assigned ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR signals of $\mathbf{6}$ is shown in Figure 3.12 and Table 3.3.


Figure 3.12: Assigned protons and carbons for compound 6.

Table 3.3: ${ }^{1} \mathrm{H}(600 \mathrm{MHz})$ and ${ }^{13} \mathrm{C}$ NMR ( 150 MHz ) shifts for compound 6 (DMSO- $\mathrm{d}_{6}$ ). The position of carbons and protons are shown in Figure 3.12.

| Position | ${ }^{13} \mathbf{C}$ NMR [ppm] | ${ }^{1} \mathbf{H}$ NMR [ppm] |
| :---: | :---: | :--- |
| 1 | 116.2 | 6.34 |
| 2 | 129.9 | 7.21 |
| 3 | 135.3 | - |
| 4 | 124.9 | 7.49 |
| 4 a | 118.9 | - |
| 5 a | 119.9 | - |
| 6 | 128.5 | 7.34 |
| 7 | 12.9 | - |
| 8 | 125.2 | $7.31-7.27$ |
| 9 | 116.4 | 6.21 |
| 9 a | 144.1 | - |
| 10 a | 143.1 | - |


| 11 | 132.6 | - |
| :---: | :---: | :--- |
| 12 | $132.3(2 \mathrm{C})$ | $7.49-7.47(2 \mathrm{H})$ |
| 13 | $117.4(2 \mathrm{C})$ | $7.31-7.27(2 \mathrm{H})$ |
| 14 | 159.0 | - |
| 15 | 68.0 | $4.23(2 \mathrm{H})$ |
| 16 | 69.4 | $3.82(2 \mathrm{H})$ |
| 17 | 70.4 | $3.65-3.63(2 \mathrm{H})$ |
| 18 | 70.3 | $3.58-3.56(2 \mathrm{H})$ |
| 19 | 70.1 | $3.55-3.54(2 \mathrm{H})$ |
| 20 | 71.8 | $3.46-3.44(2 \mathrm{H})$ |
| 21 | 58.5 | $3.24(3 \mathrm{H})$ |
| 22 | 125.0 | 8.13 |
| 23 | 128.2 | 8.18 |
| 24 | $127.8(2 \mathrm{C})$ | $8.21(2 \mathrm{H})$ |
| 25 | 125.4 | 8.29 |
| 26 | 126.9 | 8.09 |
| 27 | 128.1 | $8.34-8.31$ |
| 28 | 127.9 | 7.97 |
| 29 | 125.5 | $8.34-8.31$ |
| Pyr | 136.1 | - |
| Pyr | 131.4 | - |
| Pyr | 130.9 | - |
| Pyr | 130.6 | - |
| Pyr | 125.8 | - |
| Pyr | 124.6 | - |
| Pyr | 124.5 | - |
| 30 | 144.5 | - |
| 31 | 123.7 | $7.49-7.47$ |
| 32 | 128.7 | 7.59 |
| 33 | 133.9 | - |
| 34 | 146.0 | - |
| 35 | 125.4 | 7.54 |
| 36 | 139.7 | 8.00 |
| 37 | 141.5 | - |
| 38 | 184.2 | 9.88 |
|  |  |  |

### 3.6 Spectroscopic Identification of MEL1

NMR, IR and HRMS analysis of compound MEL1 is shown in Appendix F: Spectroscopic Data - Compound MEL1.

HRMS-analysis shows that the molecular ion found was decarboxylated, this is a common fragmentation for acids. ${ }^{[123,124]}$ This confirms that the product consist of MEL1, with a predicted molecular formula $\mathrm{C}_{4} 6 \mathrm{H}_{3} 8 \mathrm{NO}_{5} \mathrm{~S}_{2}$, with exact mass $771.2351([\mathrm{M}-\mathrm{COO}+\mathrm{H}])$. The corresponding calculated mass was 770.9548 .

The IR spectrum in appendix displayed some characteristic signals of the functional groups on compound MEL1. $\mathrm{C} \equiv \mathrm{N}$ stretching is shown as a weak peak at $2164 \mathrm{~cm}^{-1}$. The strong peak at 1574 is consistent with $\mathrm{C}=\mathrm{O}$ stretching of the carboxylic acid. C-O-C streching of the aliphatic ether is shown as a peak at 1102 $\mathrm{cm}^{-1}$.

The assigned ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR signals are shown in Figure 3.13 and Table 3.4.


Figure 3.13: Assigned protons and carbons for compound MEL1.

Table 3.4: ${ }^{1} \mathrm{H}(600 \mathrm{MHz})$ and ${ }^{13} \mathrm{C}$ NMR ( 150 MHz ) shifts for compound MEL1 (DMSO- $\mathrm{d}_{6}$ ). The position of carbons and protons are shown in Figure 3.13.

| Position | ${ }^{13}$ C NMR [ppm] | ${ }^{1}$ H NMR [ppm] |
| :---: | :---: | :--- |
| 1 | 116.3 | 6.34 |
| 2 | 130.0 | 7.21 |
| 3 | 135.4 | - |
| 4 | 128.5 | 7.33 |


| 4a | 118.9 | - |
| :---: | :---: | :--- |
| 5 a | 119.9 | - |
| 6 | 124.1 | $7.49-7.47$ |
| 7 | 127.7 | - |
| 8 | 125.7 | $7.32-7.29$ |
| 9 | 116.4 | 6.23 |
| 9 a | 144.7 | - |
| 10 a | 143.0 | - |
| 11 | 132.5 | - |
| 12 | $132.3(2 \mathrm{C})$ | $7.49-7.47(2 \mathrm{H})$ |
| 13 | $117.4(2 \mathrm{C})$ | $7.32-7.29(2 \mathrm{H})$ |
| 14 | 159.0 | - |
| 15 | 68.0 | $4.23(2 \mathrm{H})$ |
| 16 | 69.4 | $3.82(2 \mathrm{H})$ |
| 17 | 70.4 | $3.64-3.62(2 \mathrm{H})$ |
| 18 | 70.3 | $3.58-3.56(2 \mathrm{H})$ |
| 19 | 70.1 | $3.55-3.53(2 \mathrm{H})$ |
| 20 | 71.8 | $3.45-3.44(2 \mathrm{H})$ |
| 21 | 58.5 | $3.24(3 \mathrm{H})$ |
| 22 | 125.0 | 8.13 |
| 23 | 128.2 | 8.18 |
| 24 | $127.9(2 \mathrm{C})$ | $8.21-8.20(2 \mathrm{H})$ |
| 25 | 125.4 | 8.29 |
| 26 | 126.9 | 8.09 |
| 27 | 128.1 | $8.34-8.31$ |
| 28 | 127.8 | 7.97 |
| 29 | 125.5 | $8.34-8.31$ |
| Pyr | 136.1 | - |
| Pyr | 131.4 | - |
| Pyr | 130.9 | - |
| Pyr | 130.6 | - |
| Pyr | 125.8 | - |
| Pyr | 124.6 | - |
| Pyr | 124.5 | - |
| 30 | 149.2 | - |
| 31 | 124.4 | 7.57 |
| 32 | 138.4 | 7.77 |
| 33 | 135.4 | - |
| 34 | 143.0 | $8.21-8.20$ |
| 35 | 163.7 | - |
|  |  |  |

Position number 36 and 37 are not assigned. Number 37 did not show in the ${ }^{1} \mathrm{H}$ or ${ }^{13} \mathrm{C}$ NMR spectra, but the $\delta_{H}$ are expected to be in the range of $9-12 \mathrm{ppm}$ and a $\delta_{C}$ between 180-200 ppm. The shift for carbon number 36 is probably overlapping with another carbon, and is expected to be around 120-140 ppm. The exact $\delta_{C}$ could not be found, due to no H-C coupling in the HMBC NMR spectrum.

### 3.7 Spectroscopic Identification of MEL2

NMR, IR and HRMS analysis of compound MEL2 is shown in Appendix G: Spectroscopic Data - Compound MEL2.

HRMS analysis displayed the same phenomenon that was found for the dye MEL1, decarboxylation, and suggest the product MEL2 with molecule formula $\mathrm{C}_{53} \mathrm{H}_{40} \mathrm{~N}_{2} \mathrm{O}_{6} \mathrm{~S}_{3}$, exact mass 853.2228 ([M-COO+H]). Calculated mass was 853.0784.

The IR spectrum in appendix shows simlilar trends as described for MEL1.
The assigned ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR signals of MEL2 is shown in Figure 3.14 and Table 3.5.


Figure 3.14: Assigned protons and carbons for compound MEL2.

Table 3.5: ${ }^{1} \mathrm{H}(600 \mathrm{MHz})$ and ${ }^{13} \mathrm{C}$ NMR ( 150 MHz ) shifts for compound MEL2 (DMSO- $\mathrm{d}_{6}$ ). The position of carbons and protons are shown in Figure 3.14.

| Position | ${ }^{13} \mathbf{C}$ NMR [ppm] | ${ }^{1} \mathbf{H}$ NMR [ppm] |
| :---: | :---: | :--- |
| 1 | 116.2 | 6.33 |
| 2 | 129.9 | 7.20 |
| 3 | 135.2 | -7.33 |
| 4 | 128.5 | - |
| 4 a | 118.9 | - |
| 5 a | 119.8 | $7.49-7.47$ |
| 6 | 123.6 | - |
| 7 | 128.0 | $7.31-7.27$ |
| 8 | 125.1 | 6.20 |
| 9 | 116.4 | 6 |


| 9 a | 144.7 | - |
| :--- | :---: | :--- |
| 10 a | 143.0 | - |
| 11 | 132.6 | - |
| 12 | $132.4(2 \mathrm{C})$ | $7.49-7.47(2 \mathrm{H})$ |
| 13 | $117.4(2 \mathrm{C})$ | $7.31-7.27(2 \mathrm{H})$ |
| 14 | 159.0 | - |
| 15 | 68.0 | $4.23(2 \mathrm{H})$ |
| 16 | 69.4 | $3.82(2 \mathrm{H})$ |
| 17 | 70.4 | $3.65-3.63(2 \mathrm{H})$ |
| 18 | 70.3 | $3.58-3.56(2 \mathrm{H})$ |
| 19 | 70.1 | $3.55-3.54(2 \mathrm{H})$ |
| 20 | 71.8 | $3.46-3.44(2 \mathrm{H})$ |
| 21 | 58.5 | $3.25(3 \mathrm{H})$ |
| 22 | 125.0 | 8.13 |
| 23 | 128.2 | 8.18 |
| 24 | $127.9(2 \mathrm{C})$ | $8.21-8.20(2 \mathrm{H})$ |
| 25 | 125.4 | 8.29 |
| 26 | 126.9 | 8.09 |
| 27 | 128.1 | $8.33-8.31$ |
| 28 | 127.9 | 7.98 |
| 29 | 125.5 | $8.33-8.31$ |
| Pyr | 136.1 | - |
| Pyr | 131.4 | - |
| Pyr | 130.9 | - |
| Pyr | 130.6 | - |
| Pyr | 125.8 | - |
| Pyr | 124.6 | - |
| Pyr | 124.5 | - |
| $31 / 32$ | $124.9 / 125.0$ | $7.49-7.47(2 \mathrm{H})$ |
| 35 | 127.9 | 7.52 |
| 36 | 134.2 | 7.77 |
| 38 | 143.1 | $8.21-8.20$ |
| 39 | 163.4 | - |
|  |  |  |

The Carbons at position number 30, 33, 34, 37 and 40 were not assigned, due to large overlap in the ${ }^{13} \mathrm{C}$ NMR spectra. As described in Section 3.6, the acidic carbon and proton number 40 did not show in the ${ }^{13} \mathrm{C}$ or ${ }^{1} \mathrm{H}$ NMR spectra.

### 3.8 Spectroscopic Identification of 13

NMR, IR and HRMS analysis of compound $\mathbf{1 3}$ is shown in Appendix L: Spectroscopic Data - Compound 13. HRMS predicted molecular formula $\mathrm{C}_{3} 3 \mathrm{H}_{3} 6 \mathrm{~N}_{2} \mathrm{O}_{3} \mathrm{~S}_{2}$ with exact mass 573.2246 , which corresponded well with the calculated mass at 572.7791.

The IR spectrum in appendix displayed some characteristic signals of the functional groups on compound 13. The strong signal on $1654 \mathrm{~cm}^{-1}$ correspond to $\mathrm{C}=\mathrm{O}$ stretching in aldehyde. Amide $\mathrm{C}=\mathrm{O}$ stretch also appear within the same region as for aldehyde $\mathrm{C}=\mathrm{O}$ stretch. Amide $\mathrm{C}-\mathrm{N}$ stretch is shown in the spectra as a medium peak at $1211 \mathrm{~cm}^{-1}$.

The assigned ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR signals of $\mathbf{1 3}$ is shown in Figure 3.15 and Table 3.6. Assigned protons and carbons 20-25 in the figure and table were combined, due to overlapping signals.


Figure 3.15: Assigned protons and carbons for compound 13.

Table 3.6: ${ }^{1} \mathrm{H}(600 \mathrm{MHz})$ and ${ }^{13} \mathrm{C}$ NMR ( 150 MHz ) shifts for compound $\mathbf{1 3}$ $\left(\mathrm{CDCl}_{3}\right)$. The position of carbons and protons are shown in Figure 3.15.

| Position | ${ }^{13}$ C NMR [ppm] | ${ }^{1}$ H NMR [ppm] |
| :---: | :---: | :--- |
| 1 | 161.4 | - |
| 2 | 108.3 | - |
| 3 | 139.0 | - |
| 4 | 161.4 | - |
| 5 | 108.3 | - |
| 6 | 140.5 | - |
| 7 | 129.7 | - |
| 8 | 135.6 | 8.97 |
| 9 | 128.7 | $7.31-7.29$ |
| 10 | 131.1 | 7.66 |
| 11 | 130.7 | - |
| 12 | 136.3 | 8.94 |
| 13 | 126.3 | 7.60 |
| 14 | 147.3 | - |
| 15 | 138.7 | - |
| 16 | 126.4 | 7.84 |
| 17 | 130.6 | 7.94 |
| 18 | 136.0 | - |
| 19 | 191.2 | 10.03 |
| 20 | $42.3(2 \mathrm{C})$ | $4.13-4.07(4 \mathrm{H})$ |
| 21 | $30.0(2 \mathrm{C})$ | $1.81-1.73(4 \mathrm{H})$ |
| 22 | $26.6(2 \mathrm{C})$ | $1.49-1.41(4 \mathrm{H})$ |
| 23 | $31.4(2 \mathrm{C})$ | $1.36-1.32(4 \mathrm{H})$ |
| 24 | $22.6(2 \mathrm{C})$ | $1.36-1.32(4 \mathrm{H})$ |
| 25 | $14.0(2 \mathrm{C})$ | $0.91-0.88(6 \mathrm{H})$ |

### 3.9 Spectroscopic Identification of 15

NMR, IR and HRMS analysis of compound $\mathbf{1 5}$ is shown in Appendix M: Spectroscopic Data - Compound 15. HRMS predicted molecular formula $\mathrm{C}_{40} \mathrm{H}_{40} \mathrm{~N}_{2} \mathrm{O}_{4} \mathrm{~S}_{2}$ with exact mass 677.2508. The calculated mass was 676.8850 .

The IR spectrum for this compound showed similar signals as compound 13.
The assigned ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR signals of $\mathbf{1 5}$ is shown in Figure 3.16 and Table 3.7.


Figure 3.16: Assigned protons and carbons for compound 15.

Table 3.7: ${ }^{1} \mathrm{H}(600 \mathrm{MHz})$ and ${ }^{13} \mathrm{C}$ NMR ( 150 MHz ) shifts for compound $\mathbf{1 5}$ $\left(\mathrm{CDCl}_{3}\right)$. The position of carbons and protons are shown in Figure 3.16.

| Position | ${ }^{13}$ C NMR [ppm] | ${ }^{1} \mathbf{H}$ NMR [ppm] |
| :---: | :---: | :--- |
| 1 | 161.3 | - |
| 2 | 108.8 | - |
| 3 | 139.4 | - |
| 4 | 130.5 | - |
| 5 | 136.6 | 8.98 |
| 6 | 126.4 | 7.62 |
| 7 | 147.8 | - |
| 8 | 138.6 | - |
| 9 | 126.5 | 7.85 |
| 10 | 130.6 | 7.95 |
| 11 | 136.0 | - |
| 12 | 191.2 | 10.04 |
| 13 | $42.4(2 \mathrm{C})$ | $4.13(4 \mathrm{H})$ |
| 14 | $30.0(2 \mathrm{C})$ | $1.83-1.76(4 \mathrm{H})$ |
| 15 | $26.6(2 \mathrm{C})$ | $1.49-1.46(4 \mathrm{H})$ |
| 16 | $31.4(2 \mathrm{C})$ | $1.38-1.33(4 \mathrm{H})$ |
| 17 | $22.6(2 \mathrm{C})$ | $1.38-1.33(4 \mathrm{H})$ |
| 18 | $14.0(2 \mathrm{C})$ | $0.90(6 \mathrm{H})$ |

### 3.10 Spectroscopic Identification of 18

NMR, IR and HRMS analysis of compound $\mathbf{1 8}$ is shown in Appendix N: Spectroscopic Data - Compound 18.

The IR spectrum presents a strong peak at $1658 \mathrm{~cm}^{-1}$, indicating a $\mathrm{C}=\mathrm{O}$ stretching of aldehyde. Amide $\mathrm{C}=\mathrm{O}$ stretch also appear within the same region. Amide $\mathrm{C}-\mathrm{N}$ stretch is shown in the spectra as a medium peak at 1214 cm 1 . C-O-C stretchig of the aliphatic ether is shown as a weak peak at $1105 \mathrm{~cm}^{-1}$.

The assigned ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR signals of $\mathbf{1 8}$ is shown in Figure 3.17 and Table 3.8.


Figure 3.17: Assigned protons and carbons for compound 18.

Table 3.8: ${ }^{1} \mathrm{H}(600 \mathrm{MHz})$ and ${ }^{13} \mathrm{C}$ NMR ( 150 MHz ) shifts for compound $\mathbf{1 8}$ $\left(\mathrm{CDCl}_{3}\right)$. The position of carbons and protons are shown in Figure 3.17.

| Position | ${ }^{13} \mathrm{C}$ NMR [ppm] | ${ }^{1} \mathrm{H}$ NMR [ppm] |
| :---: | :---: | :---: |
| 1 | 115.8 | 6.36 |
| 2 | 129.3 | 7.12 |
| 3 | 135.9 |  |
| 4 | 128.5 | 7.33 |
| 4 a | 119.0 | - |
| 5a | 120.4 |  |
| 6 | 123.7 | 7.29 |
| 7 | 127.3 |  |
| 8 | 125.0 | 7.19 |
| 9 | 115.9 | 6.25 |
| 9 a | 145.1 | - |
| 10a | 143.0 | - |
| 11 | 132.9 |  |
| 12 | 132.0 (2C) | 7.41 (2H) |
| 13 | 116.9 (2C) | 7.21 (2H) |
| 14 | 158.9 |  |
| 15 | 67.9 | 4.26 (2H) |
| 16 | 69.8 | 3.95 (2H) |
| 17 | 71.0 | 3.81-3.79 ( 2 H ) |
| 18 | 70.3 | 3.74-3.72 (2H) |
| 19 | 70.7 | 3.70-3.68 (2H) |
| 20 | 72.0 | 3.59-3.57 (2H) |
| 21 | 59.1 | 3.40 (3H) |
| 22 | 127.5 | 8.03 |
| 23 | 124.9 | 8.20-8.18 |
| 24 | 127.4 (2C) | 8.08 (2H) |
| 25 | 125.0 | 8.16 |
| 26 | 125.0 | 8.01 |
| 27 | 124.9 | 8.20-8.18 |
| 28 | 127.2 | 7.91 |
| 29 | 124.7 | 8.20-8.18 |
| Pyr | 136.2 | - |
| Pyr | 131.5 | - |
| Pyr | 131.0 | - |
| Pyr | 128.8 | - |
| Pyr | 126.0 | - |
| Pyr | 125.5 | - |
| Pyr | 125.1 | - |


| 30 | 140.5 | - |
| :---: | :---: | :--- |
| 31 | 123.8 | 7.35 |
| 32 | 137.5 | 9.01 |
| 33 | 149.8 | - |
| 34 | 127.6 | - |
| $35 / 38$ | $115.8 / 115.9$ | - |
| $36 / 39$ | $161.2 / 161.5$ | - |
| 37 | 130.9 | - |
| 40 | $42.3(2 \mathrm{C})$ | $4.14-4.09(4 \mathrm{H})$ |
| 41 | $30.1(2 \mathrm{C})$ | $1.82-1.75(4 \mathrm{H})$ |
| 42 | $26.6(2 \mathrm{C})$ | $1.48-1.42(4 \mathrm{H})$ |
| 43 | $31.4(2 \mathrm{C})$ | $1.38-1.29(4 \mathrm{H})$ |
| 44 | $29.8(2 \mathrm{C})$ | $1.38-1.29(4 \mathrm{H})$ |
| 45 | $14.1(2 \mathrm{C})$ | $0.91-0.88(6 \mathrm{H})$ |
| 46 | 147.0 | - |
| 47 | 135.9 | 8.91 |
| 48 | 126.4 | 7.60 |
| 49 | 138.0 | - |
| 50 | 138.8 | - |
| 51 | $126.4(2 \mathrm{C})$ | $7.83(2 \mathrm{H})$ |
| 52 | $130.6(2 \mathrm{C})$ | $7.93(2 \mathrm{H})$ |
| 53 | 147.0 | - |
| 54 | 191.4 | 10.03 |

### 3.11 Spectroscopic Identification of MEL3

IR and HRMS analysis of compound MEL3 is shown in Appendix N: Spectroscopic Data - Compound MEL3. HRMS predicted molecular formula $\mathrm{C}_{76} \mathrm{H}_{71} \mathrm{~N}_{4} \mathrm{O}_{6} \mathrm{~S}_{3}$ with exact mass 1231.4536. This corresponds to the decarboxylated molecular ion, which is normal for cyano-acrylic acids. The calculated mass is 1231.4536 .

The IR spectra displays strong peaks at $2916-2850 \mathrm{~cm}^{-1}$ and $1701-1652 \mathrm{~cm}^{-1}$, which can correspond to $\mathrm{O}-\mathrm{H}$ stretching and $\mathrm{C}=\mathrm{O}$ stretching in a carboxylic acid. The weak peaks at 2214-2013 is consistent with the $\mathrm{C} \equiv \mathrm{N}$ stretching. The medium peak at $1081 \mathrm{~cm}^{-1}$ suggest the C-N stretching of the alkylated amide. Strong peaks at 1423-1307 $\mathrm{cm}^{-1}$ were found, and indicates the presence of $\mathrm{S}=\mathrm{O}$ streching.

NMR-analysis was inconclusive for this compound.


Figure 3.18: Compound MEL3.

## Conclusion and Further Work

### 4.1 Conclusion

The overall goal for this master thesis was to investigate the synthesis, chemistry and photophysical properties of phenothiazine-based dyes with various $\pi$-bridges. UV-Vis spectroscopy showed a large red-shifted adsorption maxima and corresponding low band gap energy for the extended $\pi$-conjugated dye MEL3. The DPP-dye MEL3 has presumably better photophysical properties, especially extinction coefficient, than measured, due to impurities of the dye. Dye MEL2 had the highest measured extinction coefficient of the dyes in the MEL-series. Dye AFB-43 with both thiophene and phenyl in the $\pi$-bridge, had the most blue-shifted adsorption maxima. This can be due to the thiophene-phenyl bond that has a larger dihedral angle, and decreases the orbital overlap. Sensitizer MEL3 also contains the thiophene-phenyl bond, and exchanging the phenyl with another aryl can possibly red-shift the adsorption maxima even more. The incorporation of the DPP-unit leads to sensitizers that adsorb other parts of the visible light spectra and can be used in co-sensitization. The two dyes suggested as the best combination for the co-sensitization are MEL2 and MEL3.

The synthesis of the novel dyes MEL1 and MEL2 are given in Scheme 4.1. They were formed in a successful four-step linear synthesis. The first step toward these sensitizers gave the mono-substituted product in a yield of $54 \%$. Due to the symmetry of phenothiazine $\mathbf{1}$, this is considered as a high yield. In the following synthesis no further difficulties were met, and the reactions had yields ranging from 55-94\%. The sensitizers MEL1 and MEL2 were synthesized in a total yield of
$44 \%$ and $20 \%$, respectively.




2-Cyanoacetic acid
Piperidine
Acetonitrile
94\%



Scheme 4.1: Four-step synthesis route to MEL1 and MEL2.

The first step towards the blue sensitizer MEL3 was the alkylation, as displayed in Scheme 4.2. An investigation regarding the reaction temperature, time and purification method was performed in order to lower the yield of the by-products, namely mono-alkylated $\mathbf{8}, N, O$-dialkylated 10 and $O, O$-dialkylated 11 . Long reaction time, 14 hours or more, was necessary to decrease the yield of mono-alkylated 8. A high reaction-temperature, $140{ }^{\circ} \mathrm{C}$, proved important for lowering the formation of $\mathrm{N}, \mathrm{O}$-dialkylated 10. The two purification methods studied, were crystallization and silica gel chromatography column. Re-crystallization showed to
be superfluous, since pure product 9 precipitates from cold DMF. Crystallization was a more efficient purification method in large scales, while chromatography columns was a better choice for small scales. Product 9 was isolated in a yield of $52 \%$.

The main synthetic challenge for this thesis has been the coupling between dithiopheneDPP unit and an aryl compound. A varied selection of Suzuki-Miyaura crosscoupling conditions were tested, and only a small amount of the dehalogenated product $\mathbf{1 3}$ was made. The synthetic route was changed, from a seven-step Suzukibased to a five-step direct arylation-based strategy. This allows direct coupling from the alkylated DPP 9 to form compound 13 in $68 \%$ yield, followed by another direct arylation to form molecule 18 in $73 \%$ yield with a purity of $79 \%$. The last step formed the target dye MEL3. Due to uncertainties regarding the impurities, the yield is uncertain.


Scheme 4.2: Five-step synthesis route to MEL3.

### 4.2 Further Work

The synthesis of the novel dyes MEL1 and MEL2 worked out well and the main focus for the further work will be regarding dye MEL3. The synthetic route for the target molecule MEL3 is still in need of optimization. The purification of the alkylation is one of the greatest weaknesses. Due to a limited selection of solvents that dissolves the DPP-based molecules it is also a limited selection of eluent-systems that can be used. It has been reported that alkyl-chains with the branching-point away from nitrogen have increased the solubility in many organic solvents. This could increase the yield in the alkylation step, as well as all the following DPPcontaining steps. The direct arylation was successfully, but further optimization can be done, especially concerning the equivalents of starting material to tune the reaction towards the mono-substituted product.

The presence of a thiophene-phenyl p-linker has shown to decrease the efficiency of the dyes in DSSCs. Exchanging or eliminating the phenyl between the anchoring group and the DPP-unit in dye MEL3 can red-shift the adsorption maxima more and thus a smaller band gap energy can be obtained. A five membered heteroaryl is preferable instead of the phenyl. The phenyl can also be left out, by binding the cyanoacrylic acid directly to the thiophene. This can be done by a formylation of the alkylated DPP group 9, as displayed in Scheme 4.3. After the formylation is the DPP-unit no longer symmetric, and an easier direct arylation is expected.


Scheme 4.3: Formylation of the DPP-unit to yield a dye with the anchoring group direct couplet to the thiophene on the DPP-unit.

There was some challenges regarding the analysis in the end of the synthetic-route towards MEL3. A blend of the target dye MEL3 and an oxidized by-product were detected. An isolation of compound MEL3 is therefore necessary in order to do further testing of MEL3 as a sensitizer in DSSCs.

DSSCs can be fabricated for the dyes in the MEL-series. The iodine-electrolyte is well tested within the research group. With the incorporation of DPP it is probably
possible to use a cobalt electrolyte system, which is non-toxic and allows for higher open-circuit voltage.

Further investigation of the DPP-unit for use in DSSCs has potential due to the promising measured photophysical properties. Fabrication of co-sensitized DSSC with MEL3 and MEL2 can increase the efficiency due to a broad spectral response in the visible wavelengths.

## Experimental

### 5.1 General Methods

The reagents and solvents used were commercially available from Sigma Aldrich. A magnetic stirrer was used in all reactions, and an oil bath was used in cases of heating above $23{ }^{\circ} \mathrm{C}$. All reactions were run under inert atmosphere $\left(\mathrm{N}_{2}\right)$, with degased solvents and under dry conditions (except Suzuki-Miyaura cross-coupling).

### 5.1.1 Chromatography

All reactions were monitored with thin layer chromatography, TLC (Merck, $\mathrm{F}_{254}$, silica on aluminum plates). UV-light ( 254 nm and 365 nm ) was used for visualization of the TLC-plates in case of non-colored substances. Most of the compounds had to be purified with column chromatography, with silica gel ( $40-60 \mu \mathrm{~m}$ ) as a stationary phase and an eluent system as described for each reaction. For flash column, $\mathrm{N}_{2}$ gas was used.

### 5.1.2 Nuclear Magnetic Resonance

${ }^{1} \mathrm{H}$ NMR ( $400 / 600 \mathrm{MHz}$ ) and ${ }^{13} \mathrm{C}$ NMR ( $100 / 150 \mathrm{MHz}$ ) were used to confirm the identity of the product, and were recorded at $22^{\circ} \mathrm{C}$ on a Bruker spectrometer. The solvent used was DMSO- $d_{6}$ or $\mathrm{CDCl}_{3}$, and calibration of the spectra was done by
using the signal of DMSO at $2.50 \mathrm{ppm}\left({ }^{1} \mathrm{H}\right)$ and $39.52 \mathrm{ppm}\left({ }^{13} \mathrm{C}\right)$, or that of TMS $(0 \mathrm{ppm})$ in $\mathrm{CDCl}_{3}$. Chemical shifts are reported as $\delta(\mathrm{ppm})$ and coupling constants as $\mathbf{J}(\mathrm{Hz})$.

### 5.1.3 Mass spectrometry

Synapt G2-S Q-TOF instrument form Waters ${ }^{T M}$ was used to determinate accurate mass in positive and negative mode. Samples were ionized by ACPI (AtmosphericPressure Chemical Ionization) and analyzed using an ASAP (Atmospheric Solids Analysis Probe). Waters ${ }^{T M}$ Software (Masslynx V4.1 SCN871) was used to calculate exact mass and spectrum processing.

### 5.1.4 Infrared absorption

Infrared absorption (IR) spectra were recorded with a FTIR Thermo Nicolet Nexus FT-IR Spectrometer using a Smart Endurance reflection cell. Reported frequencies were in the range of $4000-400 \mathrm{~cm}^{-1}$.

### 5.1.5 Melting point

Stuart automatic melting point SMP40 or Gallenkamp Melting Point apparatus were used to determine the melting points.

### 5.1.6 UV-Vis spectroscopy

UV-Vis spectroscopy was performed using a Hitachi U-1900 spectrometer with light-path 10 mm . Wavelengths in the region 300-900 nm were reported. The adsorption maxima were presented with wavelength $[\mathrm{nm}], \lambda$, and their corresponding molar extinction coefficients $\left[\mathrm{M}^{-1} \mathrm{~cm}^{-1}\right], \varepsilon$.

### 5.2 Synthesis

### 5.2.1 3-Bromo-10-(4-(2-(2-(2-methoxyethoxy)ethoxy)ethoxy)-phenyl)-7-(pyren-1-yl)-10H-phenothiazine (2) ${ }^{[91]}$



Compound 1 ( $500 \mathrm{mg}, 0.84 \mathrm{mmol}$ ), pyrene-1-boronic acid ( $227 \mathrm{mg}, 0.92 \mathrm{mmol}$ ), $\operatorname{Pd}\left(\mathrm{PPh}_{3}\right)_{4}(1.90 \mathrm{mg}, 0.01 \mathrm{mmol}), \mathrm{K}_{2} \mathrm{CO}_{3}(431 \mathrm{mg}, 3.36 \mathrm{mmol}), 1,4$-dioxane ( 3.00 $\mathrm{mL})$ and water $(3.00 \mathrm{~mL})$ were stirred for 6 hours at $80^{\circ} \mathrm{C}$. The reaction was then quenched with water ( 40 mL ) and extracted with EtOAc ( 120 mL ). The organic phase was washed with water ( 40 mL ) and dried with anhydrous $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was filtered and removed in vacuo. Silica gel column chromatography was performed two times ( 95 g silica, $\mathrm{EtOAc} / \mathrm{pentane}, 1: 1,79 \mathrm{~g}$ silica, $\mathrm{EtO}_{2}$ ) where product was collected both times to yield compound $\mathbf{2}, 324 \mathrm{mg}(0.45 \mathrm{mmol}, 54 \%)$, as a yellow solid.
${ }^{1} \mathrm{H}$ NMR ( 600 MHz, DMSO- $d_{6}$ ) $\delta: 8.32-8.26(\mathrm{~m}, 3 \mathrm{H}), 8.20(\mathrm{~s}, 2 \mathrm{H}), 8.16-8.06$ $(\mathrm{m}, 3 \mathrm{H}), 7.94(\mathrm{~d}, \mathrm{~J}=7.9,1 \mathrm{H}), 7.43(\mathrm{~d}, \mathrm{~J}=8.9,2 \mathrm{H}), 7.30-7.25(\mathrm{~m}, 4 \mathrm{H}), 7.18(\mathrm{dd}, \mathrm{J}$ $=8.5, \mathrm{~J}=2.1,1 \mathrm{H}), 7.11(\mathrm{dd}, \mathrm{J}=8.8, \mathrm{~J}=2.3,1 \mathrm{H}), 6.32(\mathrm{~d}, \mathrm{~J}=8.5,1 \mathrm{H}), 6.08(\mathrm{~d}, \mathrm{~J}=$ $8.8,1 \mathrm{H}), 4.20(\mathrm{t}, \mathrm{J}=8.9,2 \mathrm{H}), 3.80(\mathrm{t}, \mathrm{J}=8.9,2 \mathrm{H}), 3.63-3.61(\mathrm{~m}, 2 \mathrm{H}), 3.56-3.52$ $(\mathrm{m}, 4 \mathrm{H}), 3.44-3.42(\mathrm{~m}, 2 \mathrm{H}), 3.23(\mathrm{~s}, 3 \mathrm{H})$. The spectroscopic data corresponded well with that reported in the specialization project. ${ }^{[91]}$

### 5.2.2 10-(4-(2-(2-(2-Methoxyethoxy)ethoxy)ethoxy) phenyl)-3,7-di(pyren-1-yl)-10H-phenothiazine (3) ${ }^{[91]}$



Product 3 was isolated from the synthesis described in Section 5.2.1.
${ }^{1} \mathrm{H}$ NMR ( 400 MHz, DMSO- $d_{6}$ ) $\delta: 8.36-8.29(\mathrm{~m}, 3 \mathrm{H}), 8.22(\mathrm{~s}, 2 \mathrm{H}), 8.19(\mathrm{~d}$, J = $4.2,2 H), 8.10(\mathrm{t}, \mathrm{J}=7.6,1 \mathrm{H}), 8.00(\mathrm{~d}, \mathrm{~J}=7.9,1 \mathrm{H}), 7.60(\mathrm{~d}, \mathrm{~J}=8.9,1 \mathrm{H}), 7.38(\mathrm{~d}$, $\mathrm{J}=2.0,1 \mathrm{H}), 7.34(\mathrm{~d}, \mathrm{~J}=9.0,1 \mathrm{H}), 7.26(\mathrm{dd}, \mathrm{J}=8.5, \mathrm{~J}=2.2,1 \mathrm{H}), 6.43(\mathrm{~d}, \mathrm{~J}=8.5$, $1 \mathrm{H}), 4.26(\mathrm{t}, \mathrm{J}=4.8,2 \mathrm{H}), 3.84(\mathrm{t}, \mathrm{J}=4.2,2 \mathrm{H}), 3.66-3.63(\mathrm{~m}, 2 \mathrm{H}), 3.60-3.55(\mathrm{~m}$, 4H), 3.46-3.43(m, 2H), 3.25 (s, 3H). The spectroscopic data corresponded well with that reported in the specialization project. ${ }^{[91]}$

### 5.2.3 10-(4-(2-(2-(2-Methoxyethoxy)ethoxy)ethoxy)phenyl)-3-(pyren-1-yl)-7-(4,4,5,5-tetramethyl-1,3,2-dioxa-borolan-2-yl)-10H-phenothiazine (4)



2



Compund (2) ( $270 \mathrm{mg}, 0.38 \mathrm{mmol}$ ), $\mathrm{Pd}(\mathrm{MeCN})_{2} \mathrm{Cl}_{2}(1.00 \mathrm{mg}, 0.02 \mathrm{mmol})$, SPhos ( $20.0 \mathrm{mg}, 0.49 \mathrm{mmol}$ ), pinacol borane ( $72.3 \mathrm{mg}, 0.57 \mathrm{mmol}$ ) and dry dioxane and $\mathrm{NEt}_{3}$ were stirred for 3 hours at $80^{\circ} \mathrm{C}$. The reaction was cooled to room temperature before it was filtrated through a thin pad of Celite $\left(\mathrm{SiO}_{2}, \mathrm{EtOAc}\right)$. The product $\mathbf{4}$ was not purified further, due to its unstability on silica. Drying gave 270 $\mathrm{mg}(0.35 \mathrm{mmol}, 94 \%)$ of a yellow solid, mp. 59.7-62.2 ${ }^{\circ} \mathrm{C}$.
${ }^{1} \mathrm{H}$ NMR ( 600 MHz, DMSO- $d_{6}$ ) $\delta: 8.33-8.31(\mathrm{~m}, 2 \mathrm{H}), 8.29(\mathrm{~d}, \mathrm{~J}=7.5,1 \mathrm{H}), 8.21$ ( $\mathrm{s}, 2 \mathrm{H}$ ), $8.17(\mathrm{~d}, \mathrm{~J}=9.3,1 \mathrm{H}), 8.12(\mathrm{~d}, \mathrm{~J}=9.3,1 \mathrm{H}), 8.09(\mathrm{t}, \mathrm{J}=7.6,1 \mathrm{H}), 7.97(\mathrm{~d}$, $\mathrm{J}=7.9,1 \mathrm{H}), 7.45(\mathrm{~d}, \mathrm{~J}=8.9,2 \mathrm{H}), 7.30-7.26(\mathrm{~m}, 4 \mathrm{H}), 7.23(\mathrm{dd}, \mathrm{J}=8.3, \mathrm{~J}=1.4$, $1 \mathrm{H}), 7.19(\mathrm{dd}, \mathrm{J}=8.4, \mathrm{~J}=2.2,1 \mathrm{H}), 6.30(\mathrm{~d}, \mathrm{~J}=8.4,1 \mathrm{H}), 6.19(\mathrm{~d}, \mathrm{~J}=8.3,1 \mathrm{H}), 4.23$ (t, J = 4.7, 2H), $3.81(\mathrm{t}, \mathrm{J}=4.6,2 \mathrm{H}), 3.64-3.61(\mathrm{~m}, 2 \mathrm{H}), 3.58-3.56(\mathrm{~m}, 2 \mathrm{H}), 3.55$ - $3.53(\mathrm{~m}, 2 \mathrm{H}), 3.45-3.44(\mathrm{~m}, 2 \mathrm{H}), 3.24(\mathrm{~s}, 3 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( 150 MHz , DMSO$\left.d_{6}\right) \delta: 158.9,146.7,143.1,136.1,135.3,134.4,132.8,132.5,132.3$ (2C), 131.4, $130.8,130.5,129.7$, 128.3, 128.1, 128.0, 127.9, 127.8 (2C), 126.9, 125.8, 125.5, $125.4,124.9,124.6,124.5,122.7,119.4,118.0,117.3$ (2C), 116.2, 115.4, 84.0, 71.7, 70.4, 70.3, 70.1, 69.3, 67.9, 58.5; IR (neat, $\mathrm{cm}^{-1}$ ) $\nu: 3038$ (w), 1606 (w), 1507 (m), 1348 (m), 1140 (s), 846 (m), 670 (m), 549 (w); HRMS (APCI/ASAP+, $\mathrm{m} / \mathrm{z})$ : found $764.3217\left([\mathrm{M}+\mathrm{H}]^{+}\right.$, calculated $\mathrm{C}_{47} \mathrm{H}_{47} \mathrm{BNO}_{6} \mathrm{~S}$, 764.7533).

### 5.2.4 5-(10-(4-(2-(2-(2-Methoxyethoxy)ethoxy)ethoxy) phenyl)-7-(pyren-1-yl)-10H-phenothiazin-3-yl) thiophene-2-carbaldehyde (5)



Compound 4 ( $190 \mathrm{mg}, 0.25 \mathrm{mmol}$ ), 5-bromothenaldehyde ( $43.2 \mathrm{mg}, 0.23 \mathrm{mmol}$ ), $\mathrm{Pd}(\mathrm{OAc})_{2}(1.52 \mathrm{mg}, 0.01 \mathrm{mmol}), \mathrm{SPhos}(5.57 \mathrm{mg}, 0.01 \mathrm{mmol}), \mathrm{K}_{2} \mathrm{CO}_{3}(125 \mathrm{mg}$, $0.90 \mathrm{mmol})$, 1,4-dioxane ( 1.30 mL ) and water ( 1.30 mL ) were stirred for 18 hours at $80^{\circ} \mathrm{C}$. The reaction was then added water ( 40 mL ) and extracted with EtOAc $(3.40 \mathrm{~mL})$. The organic phase was washed with water $(40 \mathrm{~mL})$ and dried with anhydrous $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was filtered and removed in vacuo. Two Silica gel chromatography columns ( 39 g silica, $\mathrm{Et}_{2} \mathrm{O}: \mathrm{EtOAc}, 1: 10, \mathrm{R}_{f}=0.45,39 \mathrm{~g}$ silica, $\mathrm{EtOAc}, \mathrm{R}_{f}=0.37$ ), where product was isolated from both columns, gave the product 5 as a yellow solid, $157 \mathrm{mg}(0.21 \mathrm{mmol}, 93 \%)$, mp. $123.1-123.9^{\circ} \mathrm{C}$.
${ }^{1} \mathrm{H}$ NMR ( 600 MHz, DMSO- $d_{6}$ ) $\delta: 9.86(\mathrm{~s}, 1 \mathrm{H}), 8.34-8.31(\mathrm{~m}, 2 \mathrm{H}), 8.29(\mathrm{~d}, \mathrm{~J}=$ $7.5,1 \mathrm{H}), 8.21(\mathrm{~s}, 2 \mathrm{H}), 8.18(\mathrm{~d}, \mathrm{~J}=9.4,1 \mathrm{H}), 8.13(\mathrm{~d}, \mathrm{~J}=9.3,1 \mathrm{H}), 8.09(\mathrm{t}, \mathrm{J}=7.6$, $1 \mathrm{H}), 7.99(\mathrm{~d}, \mathrm{~J}=4.0,1 \mathrm{H}), 7.97(\mathrm{~d}, \mathrm{~J}=7.9,1 \mathrm{H}), 7.63(\mathrm{~d}, \mathrm{~J}=4.0,1 \mathrm{H}), 7.56(\mathrm{~d}, \mathrm{~J}=$ $2.2,1 \mathrm{H}), 7.49(\mathrm{~d}, \mathrm{~J}=8.8,2 \mathrm{H}), 7.39(\mathrm{dd}, \mathrm{J}=8.7, \mathrm{~J}=2.2,1 \mathrm{H}), 7.34(\mathrm{~d}, \mathrm{~J}=2.1,1 \mathrm{H})$ $7.30(\mathrm{~d}, \mathrm{~J}=8.9,2 \mathrm{H}), 7.21(\mathrm{dd}, \mathrm{J}=8.4, \mathrm{~J}=2.1,1 \mathrm{H}), 6.34(\mathrm{~d}, \mathrm{~J}=8.4,1 \mathrm{H}), 6.2(\mathrm{~d}, \mathrm{~J}$ $=8.7,1 \mathrm{H}), 4.23(\mathrm{t}, \mathrm{J}=4.3,2 \mathrm{H}), 3.81(\mathrm{t}, \mathrm{J}=4.6,2 \mathrm{H}), 3.64-3.62(\mathrm{~m}, 2 \mathrm{H}), 3.58-$ $3.56(\mathrm{~m}, 2 \mathrm{H}), 3.55-3.53(\mathrm{~m}, 2 \mathrm{H}), 3.45-3.44(\mathrm{~m}, 2 \mathrm{H}), 3.24(\mathrm{~s}, 3 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR (150 MHz, DMSO- $d_{6}$ ) $\delta: 184.2,159.0,152.1,145.1,142.9,141.5,139.8,136.1,135.5$, $132.5,132.3$ (2C), 131.4, 130.9, 130.6, 130.0, 128.5, 128.2, 128.1, 127.9, 127.8 (2C), 127.4, 126.9, 126.1, 125.8, 125.5, 125.4, 125.0, 124.8, 124.6, 124.5, 124.4, $119.9,118.8,117.3$ (2C), 116.3, 116.2, 71.8, 70.4, 70.3, 70.1, 69.4, 68.0, 58.5; IR (neat, $\left.\mathrm{cm}^{-1}\right) \nu: 3036$ (w), 2869 (w), 1654 (m/s), 1431 (s), 1303 (m), 1223 (s), 1102 (m), 1055 (m), 799 (m), 682 (w/m), 552 (w); HRMS (APCI/ASAP+, m/z): found $748.2191\left([\mathrm{M}+\mathrm{H}]^{+}\right.$, calculated $\left.\mathrm{C}_{46} \mathrm{H}_{37} \mathrm{NO}_{5} \mathrm{~S}_{2}, 747.9182\right)$.

### 5.2.5 5’-(10-(4-(2-(2-(2-Methoxyethoxy)ethoxy)ethoxy) phenyl)-7-(pyren-1-yl)-10H-phenothiazin-3-yl)- [2,2'-bithiophene]-5-carbaldehyde (6)



Compound 6 was made with the same approach as previously described for compound 5 (Section 5.2.4). Compound $4(150 \mathrm{mg}, 0.20 \mathrm{mmol})$ and the other reagents were stirred for 5 hours. Silica gel column chromatography was used two times ( 38 g silica, EtOAc, $\mathrm{R}_{f}=0.42$ ) and compound 6 was isolated after both columns, resulting in product 6 as a yellow solid, $80.8 \mathrm{mg}(0.1 \mathrm{mmol}, 55 \%), \mathrm{mp} .119 .1-$ $120.6^{\circ} \mathrm{C}$.
${ }^{1}{ }^{1} \mathrm{H}$ NMR ( 600 MHz, DMSO- $d_{6}$ ) $\delta: 9.88(\mathrm{~s}, 1 \mathrm{H}), 8.34-8.31(\mathrm{~m}, 2 \mathrm{H}), 8.29(\mathrm{~d}, \mathrm{~J}$ $=7.3,1 \mathrm{H}), 8.21(\mathrm{~s}, 2 \mathrm{H}), 8.18(\mathrm{~d}, \mathrm{~J}=9.4,1 \mathrm{H}), 8.13(\mathrm{~d}, 9.2,1 \mathrm{H}), 8.09(\mathrm{t}, \mathrm{J}=7.6$, $1 \mathrm{H}), 7.99(\mathrm{~d}, \mathrm{~J}=4.0,1 \mathrm{H}), 7.98(\mathrm{~d}, \mathrm{~J}=7.9,1 \mathrm{H}), 7.59(\mathrm{~d}, \mathrm{~J}=3.9,1 \mathrm{H}), 7.53(\mathrm{~d}, \mathrm{~J}=$ 3.6, 1H), $7.49-7.47$ (m, 4H), $7.34(\mathrm{~d}, \mathrm{~J}=2.1,1 \mathrm{H}), 7.31-7.27$ (m, 3H), 7.21 (dd, $\mathrm{J}=8.4, \mathrm{~J}=2.1,1 \mathrm{H}), 6.34(\mathrm{~d}, \mathrm{~J}=8.4,1 \mathrm{H}), 6.21(\mathrm{~d}, \mathrm{~J}=8.6,1 \mathrm{H}), 4.23(\mathrm{t}, \mathrm{J}=4.5$, $2 \mathrm{H}), 3.82(\mathrm{t}, \mathrm{J}=4.6,2 \mathrm{H}), 3.65-3.63(\mathrm{~m}, 2 \mathrm{H}), 3.58-3.56(\mathrm{~m}, 2 \mathrm{H}), 3.55-3.54(\mathrm{~m}$, 2H), 3.46-3.44 (m, 2H), 3.25 ( $\mathrm{s}, 3 \mathrm{H}$ ); ${ }^{13} \mathrm{C}$ NMR ( 150 MHz , DMSO- $d_{6}$ ) $\delta: 184.2$, $159.0,146.0,144.5,144.1,143.1,141.5,139.7,136.1,135.3,133.9,132.6,132.3$ (2C), 131.4, 130.9, 130.6, 129.9, 128.7, 128.5, 128.2, 128.1, 127.9 (2C), 127.8 (2C), 126.9, 125.8, 125.5, 125.4 (2C), 125.2, 125.0, 124.9, 124.6, 124.5, 123.7, 119.9, 118.9, 117.4 (2C), 116.4, 116.2, 71.8, 70.4, 70.3, 70.1, 69.4, 68.0, 58.5; IR (neat, $\mathrm{cm}^{-1}$ ) $\nu: 2923$ (w), 1654 (s), 1606 (w), 1505 (m), 1444 (s), 1224 (s), 1102 (m), $791(\mathrm{~m}), 551(\mathrm{w})$; HRMS (APCI/ASAP+, m/z): found $830.2069\left([\mathrm{M}+\mathrm{H}]^{+}\right.$, calculated $\mathrm{C}_{50} \mathrm{H}_{39} \mathrm{NO}_{5} \mathrm{~S}_{3}, 830.0418$ ).

### 5.2.6 (e)-2-Cyano-3-(5-(10-(4-(2-(2-(2-methoxyethoxy) ethoxy)ethoxy)phenyl)-7-(pyren-1-yl)-10H-phenothiazin-3-yl)thiophen-2-yl)acrylic acid (MEL1)




Compound 5 ( $150 \mathrm{mg}, 0.20 \mathrm{mmol}$ ), 2-cyanoacetic acid ( $162 \mathrm{mg}, 1.90 \mathrm{mmol}$ ), piperidine $(0.20 \mathrm{~mL}, 1.14 \mathrm{mmol})$ and acetonitrile $(22 \mathrm{~mL})$ were stirred for 2 hours at reflux $\left(84{ }^{\circ} \mathrm{C}\right)$. After reaching room temperature, the reaction was quenched with acidic water $(120 \mathrm{~mL}, 2 \mathrm{M} \mathrm{HCl})$, and stirred for 15 min . The mixture was extracted with chloroform ( 100 mL ) and dried over anhydrous $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was filtered and removed in vacuo. Silica gel column chromatography ( 38 g silica, gradient DCM - DCM:MeOH, 5:1) gave product MEL1 as a red solid, 153 mg ( $0.19 \mathrm{mmol}, 94 \%$ ), mp. $172.7-173.4^{\circ} \mathrm{C}$.
${ }^{1} \mathrm{H}$ NMR ( 600 MHz, DMSO- $d_{6}$ ) $\delta: 8.34-8.31(\mathrm{~m}, 2 \mathrm{H}), 8.29(\mathrm{~d}, \mathrm{~J}=7.4,1 \mathrm{H}), 8.21$ $-8.20(\mathrm{~m}, 3 \mathrm{H}), 8.18(\mathrm{~d}, \mathrm{~J}=9.4,1 \mathrm{H}), 8.13(\mathrm{~d}, \mathrm{~J}=9.2,1 \mathrm{H}), 8.09(\mathrm{t}, \mathrm{J}=7.6,1 \mathrm{H}), 7.97$ $(\mathrm{d}, \mathrm{J}=7.8,1 \mathrm{H}), 7.77(\mathrm{~d}, \mathrm{~J}=3.7,1 \mathrm{H}), 7.6(\mathrm{~d}, \mathrm{~J}=3.91,1 \mathrm{H}), 7.49-7.47(\mathrm{~m}, 3 \mathrm{H})$, $7.33(\mathrm{~d}, \mathrm{~J}=2.1,1 \mathrm{H}), 7.32-7.29(\mathrm{~m}, 3 \mathrm{H}), 7.21(\mathrm{dd}, \mathrm{J}=8.4, \mathrm{~J}=2.0,1 \mathrm{H}), 6.34(\mathrm{~d}, \mathrm{~J}$ $=8.4,1 \mathrm{H}), 6.23(\mathrm{~d}, \mathrm{~J}=8.7,1 \mathrm{H}), 4.23(\mathrm{t}, \mathrm{J}=4.2,2 \mathrm{H}), 3.82(\mathrm{t}, \mathrm{J}=4.4,2 \mathrm{H}), 3.64-$ $3.62(\mathrm{~m}, 2 \mathrm{H}), 3.58-3.56(\mathrm{~m}, 2 \mathrm{H}), 3.55-3.53(\mathrm{~m}, 2 \mathrm{H}), 3.45-3.44(\mathrm{~m}, 2 \mathrm{H}), 3.24(\mathrm{~s}$, $3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( 150 MHz, DMSO- $d_{6}$ ) $\delta: 163.7,159.0,149.2,144.7,143.0$ (2C), $138.4,136.1,135.4$ (2C), 132.5, 132.3 (2C), 131.4, 130.9, 130.6, 130.0, 128.5, 128.2, 128.1, 127.9 (2C), 127.8, 127.7, 126.9, 125.8, 125.7, 125.5, 125.4, 125.0, 124.6 , 124.5, 124.4, 124.1, 119.9, 118.9, 117.4 (2C), 116.4, 116.3, 71.8, 70.4, 70.3, 70.1, 69.4, 68.0, 58.5. IR (neat, $\mathrm{cm}^{-1}$ ) $\nu: 3035$ (w), 2915 (w), 2849 (w), 2164 (w), 1710 (w), 1574 (m/s), 1472 (m/s), 1304 (s), 1242 (s), 1102 (m), 1060 (m), 939 (w), 800 (m/s), 719 (w/m), 617 (w), 547 (w). HRMS (APCI/ASAP+, $\mathrm{m} / \mathrm{z}$ ): found $771.2351\left([\mathrm{M}-\mathrm{COO}+\mathrm{H}]\right.$, calculated $\mathrm{C}_{48} \mathrm{H}_{38} \mathrm{~N}_{2} \mathrm{O}_{4} \mathrm{~S}_{2}$, 770.9548)

### 5.2.7 (e)-2-Cyano-3-(5'-(10-(4-(2-(2-(2-methoxyethoxy) ethoxy)ethoxy)phenyl)-7-(pyren-1-yl)-10H-phenothiazin-3-yl)-[2,2'-bithiophen]-5-yl)acrylic acid (MEL2)




The dye MEL2 was made with the same reaction conditions as dye MEL1. Compound $6(79 \mathrm{mg}, 0.10 \mathrm{mmol})$ and the other reagents were stirred for 2 hours. Purification by silica gel column chromatography ( 40 g silica, DCM - DCM:Metanol, 10:1) gave dye MEL2 as a red solid, 58.3 mg ( $0.06 \mathrm{mmol}, 72 \%$ ), mp. 171.4 $172.1^{\circ} \mathrm{C}$.
${ }^{1}{ }^{1} \mathrm{H}$ NMR ( 600 MHz, DMSO- $d_{6}$ ) $\delta: 8.33-8.31(\mathrm{~m}, 2 \mathrm{H}), 8.29(\mathrm{~d}, \mathrm{~J}=7.26,1 \mathrm{H})$, $8.21(\mathrm{~s}, 3 \mathrm{H}), 8.18(\mathrm{~d}, \mathrm{~J}=9.4,1 \mathrm{H}), 8.13(\mathrm{~d}, 9.3,1 \mathrm{H}), 8.09(\mathrm{t}, \mathrm{J}=7.6,1 \mathrm{H}), 7.98(\mathrm{~d}, \mathrm{~J}$ $=7.9,1 \mathrm{H}), 7.77$ (broad, 1H), $7.52(\mathrm{~d}, \mathrm{~J}=3.8,1 \mathrm{H}), 7.49-7.47(\mathrm{~m}, 5 \mathrm{H}), 7.33(\mathrm{~d}, \mathrm{~J}=$ $2.1,1 \mathrm{H}), 7.31-7.27(\mathrm{~m}, 3 \mathrm{H}), 7.20(\mathrm{dd}, \mathrm{J}=8.4, \mathrm{~J}=2.1,1 \mathrm{H}), 6.33(\mathrm{~d}, \mathrm{~J}=8.4,1 \mathrm{H})$, $6.20(\mathrm{~d}, \mathrm{~J}=8.6,1 \mathrm{H}), 4.23(\mathrm{t}, \mathrm{J}=4.5,2 \mathrm{H}), 3.82(\mathrm{t}, \mathrm{J}=4.6,2 \mathrm{H}), 3.65-3.63(\mathrm{~m}, 2 \mathrm{H})$, $3.58-3.56(\mathrm{~m}, 2 \mathrm{H}), 3.55-3.54(\mathrm{~m}, 2 \mathrm{H}), 3.46-3.44(\mathrm{~m}, 2 \mathrm{H}), 3.25(\mathrm{~s}, 3 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( 150 MHz, DMSO- $d_{6}$ ) $\delta: 159.0,144.0,136.1,135.2,134.2,132.6,132.4$ (2C), 131.4, 130.9, 130.6, 129.9, 128.5, 128.2, 128.1, 128.0, 127.9 (3C), 126.9, $125.8,125.5,125.4$ (2C), 125.1, 125.0 (2C), 124.9, 124.6, 124.5, 123.6, 119.8, 118.9, 117.4 (2C), 116.4, 116.2, 71.8, 70.4, 70.3, 70.1, 69.4, 68.0, 58.5; IR (neat, $\left.\mathrm{cm}^{-1}\right) \nu: 2916$ (w), 2207 (w), 1711 (m), 1505 (m), 1440 (m), 1241 (s), 1052 (m), $791(\mathrm{~m} / \mathrm{s}), 549(\mathrm{w}), 447(\mathrm{w})$. HRMS (APCI/ASAP+, m/z): found 853.2228 ([M-COO+H], calculated $\mathrm{C}_{52} \mathrm{H}_{41} \mathrm{~N}_{2} \mathrm{O}_{4} \mathrm{~S}_{3}, 853.0784$ ).

### 5.2.8 2,5-Dihexyl-3,6-di(thiophen-2-yl)-2,5-dihydropyrrolo [3,4-c]pyrrole-1,4-dione (9) ${ }^{\text {[125] }}$



Table 5.1: Summary of alkylation reactions and purification. Hx is short for hexane. All purifications and yields in this table are from the first purification step, and some of them were purified further.

| Entry | Scale[mg] | Heat $\left[{ }^{\circ} \mathrm{C}\right]$ | Time [h] | Crude [\%] |  | Purification |  | Yield <br> [\%] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | $8^{a}$ | $\mathbf{1 0}^{b}$ | Cryst ${ }^{\text {c }}$ | Chrom. ${ }^{\text {d }}$ |  |
| 1 | 100 | 120 | 3 | 27 | 18 | - | $\underset{7: 3}{\mathrm{CHCl}_{3}}: \mathrm{Hx}$ | 11 |
| 2 | 750 | 120 | 25 | 2 | 22 | $\begin{gathered} \text { DMF } \\ (\mathrm{DCM} / \mathrm{Hx}) \end{gathered}$ | - | 9 |
| 3 | 150 | 120 | 25 | 4 | 19 | (DCM/Hx) | - | - |
| 4 | 100 | 120 | 24 | 7 | 20 | - | - | - |
| 5 | 100 | 120 | 24 | 6 | 18 | DMF | - | 31 |
| 6 | 100 | 120 | 24 | 7 | 9 | - | $\begin{gathered} \mathrm{Hx}: \underset{9: 1}{\mathrm{EtOAc}} \end{gathered}$ | 46 |
| 7 | 100 | 140 | 24 | 7 | 5 | - | - | - |
| 8 | 1000 | 140 | 24 | 6 | 4 | DMF | - | 31 |
| 9 | 1000 | 140 | 21 | 6 | 3 | - | $\begin{gathered} \mathrm{Hx}: \mathrm{EtOAc} \\ 9: 1 \end{gathered}$ | 10 |
| 10 | 2000 | 140 | 14 | 4 | 2 | DMF |  | 43 |
| 11 | 2000 | 155 | 20 | 7 | 3 | DMF | - | 46 |

${ }^{a} \mathrm{~N}$-monoalkylated $\mathbf{8}$.
${ }^{b} \mathrm{~N}, \mathrm{O}$-dialkylated $\mathbf{1 0}$.
${ }^{c}$ Crystallization (re-crystallization).
${ }^{d}$ Silica gel chromatography column.

The alkylation reactions were performed with varying different reaction conditions or purification methods, as shown in Table 5.1. Compound 7 and $\mathrm{K}_{2} \mathrm{CO}_{3}$ ( 10 eq.) were dissolved in DMF and heated. $\mathrm{Bu}_{4} \mathrm{~N}^{+} \mathrm{HSO}_{4}^{-}$( 0.05 eq.) was also present in experiment entry 1-4. After approximately 30 minutes 1 -bromohexane ( 10 eq.) was added to the mixture. After full convertion the reaction was removed from the heat, and allowed to reach room temperature. Silica gel column chromatography or crystallization (or re-crystallization) were used as purification methods, as shown in Table 5.1. With eluent system $\mathrm{CHCl}_{3}$ :hexane was the $\mathrm{R}_{f}$ value 0.18 and 0.14 for the $\mathrm{N}, \mathrm{N}$-dialkylated 9 and $\mathrm{N}, \mathrm{O}$-dialkylated $\mathbf{1 0}$, and 0.25 and 0.21 in hexane:EtOAc, respectively. Product 9 was isolated as dark pink/gold crystals.

Among others, entry 10 displayed in Table 5.1, were purified further. Silica gel column chromatography ( 104 g silica, hexane:EtOAc, 9:1) was used as a purification method after the DMF crystallization. Product 9 was isolated as a dark pink solid, $281 \mathrm{mg}(0.60 \mathrm{mmol}, 9 \%)$, and resulted in a total yield of $52 \%$.
${ }^{1}{ }^{H} \mathrm{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta: 8.95(\mathrm{dd}, \mathrm{J}=3.9, \mathrm{~J}=1.0,2 \mathrm{H}), 7.66(\mathrm{dd}, \mathrm{J}=5.0, \mathrm{~J}$ $=1.0,2 \mathrm{H}), 7.31(\mathrm{t}, \mathrm{J}=9.0,2 \mathrm{H}), 4.10(\mathrm{t}, \mathrm{J}=8.0,4 \mathrm{H}), 1.77$ (quintet, $\mathrm{J}=7.8,4 \mathrm{H})$, 1.46-1.41(m, 4H), 1.38-1.37(m, 8H), $0.91(\mathrm{t}, \mathrm{J}=7.0,6 \mathrm{H})$. The spectroscopic data corresponded well with that reported previously. ${ }^{[125]}$

### 5.2.9 2-Hexyl-4-(hexyloxy)-3,6-di(thiophen-2-yl)pyrrolo[3,4] pyrrol-1(2H)-one (10) ${ }^{[77]}$



Product $\mathbf{1 0}$ was isolated, $43.8 \mathrm{mg}(0.09 \mathrm{mmol})$, as a pink solid from the synthesis described in Section 5.2.8.
${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta: 8.47(\mathrm{dd}, \mathrm{J}=3.8, \mathrm{~J}=0.9,1 \mathrm{H}), 8.28(\mathrm{dd}, \mathrm{J}=3.8, \mathrm{~J}$ $=0.8,1 \mathrm{H}), 7.73(\mathrm{dd}, \mathrm{J}=5.1, \mathrm{~J}=0.8,1 \mathrm{H}), 7.51(\mathrm{dd}, \mathrm{J}=5.0, \mathrm{~J}=0.8,1 \mathrm{H}), 7.29-7.27$ $(\mathrm{m}, 1 \mathrm{H}), 7.22(\mathrm{dd}, \mathrm{J}=3.7, \mathrm{~J}=1.1,1 \mathrm{H}), 4.61(\mathrm{t}, \mathrm{J}=6.5,2 \mathrm{H}), 4.00(\mathrm{t}, \mathrm{J}=7.8,2 \mathrm{H})$, 1.91-1.84 (m, 2H), 1.77-1.70 (m, 2H), 1.40-1.30(m, 12H), 0.96-0.88 (m, 6 H ). The spectroscopic data corresponded well with that reported previously. ${ }^{[77]}$

### 5.2.10 3,6-bis(5-Bromothiophen-2-yl)-2,5-dihexyl-2,5- dihydropyrrolo [3,4]pyrrole-1,4-dione (12) ${ }^{[126]}$



Compound 9 ( $700 \mathrm{mg}, 1.49 \mathrm{mmol}$ ) and NBS ( $558 \mathrm{mg}, 3.14 \mathrm{mmol}$ ) were dissolved in degassed $\mathrm{CHCl}_{3}(10 \mathrm{~mL})$ and acetic acid $(10 \mathrm{~mL})$. The reaction was stirred for 15 hours, before it was quenched with water $(16 \mathrm{~mL})$. The water phase was extracted with DCM $(3.16 \mathrm{~mL})$ and the organic phase was washed with water (16 $\mathrm{mL})$ and brine $(16 \mathrm{~mL})$. The organic phase was dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, and filtered. DCM was removed in vacuo.

The same experiment was run with a 500 mg and 200 mg scale. All three experiments with a $99 \%$ yield of compound $\mathbf{1 2}$.
${ }^{1} \mathrm{H} \operatorname{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta: 8.67(\mathrm{~d}, \mathrm{~J}=4.2,2 \mathrm{H}), 7.24(\mathrm{~d}, \mathrm{~J}=4.2,2 \mathrm{H}), 3.98$ $(\mathrm{t}, \mathrm{J}=7.7,4 \mathrm{H}$ ), 1.71 (quintet, $\mathrm{J}=7.5,4 \mathrm{H}$ ), 1.41 (quintet, $\mathrm{J}=7.6,4 \mathrm{H}$ ), 1.34-1.32 $(\mathrm{m}, 8 \mathrm{H}), 0.98(\mathrm{t}, \mathrm{J}=6.8,6 \mathrm{H})$. The spectroscopic data corresponded well with that reported previously. ${ }^{[100]}$

### 5.2.11 Attempted Synthesis of 4-(5-(4-(5-Bromothiophen-2-yl)-2,5-dihexyl-3,6-dioxo-2,3,5,6-tetrahydropyrrolo [3,4]pyrrol-1-yl)thiophen-2-yl)benzaldehyde (14)



A summary of the different experiments are shown in Table 5.2, with varied catalysts and ligands. Compound $\mathbf{1 2}$, 4-formylphenylboronic acid (1 eq.), $\mathrm{K}_{2} \mathrm{CO}_{3}$ (3 eq.), Pd-catalyst, ligand, dioxane and water were stirred at $80^{\circ} \mathrm{C}$. The reaction mixture was extracted with chloroform before the solvent was removed in vacuo. Experiment Entry 1 was an attempt to get the disubstituted product 15 and had 3 eq. of 4-formylphenylboronic acid and more catalyst and ligand, as shown in Table 5.2. An unknown compound was also formed, it is not included in the table, but it is the remaining amount of the $100 \%$ in the crude material. Only one thing is certain about the unknown compound, it is not the homo-couplet benzaldehyde.

Table 5.2: Summary of suzuki cross coupling experiments on DPP, with varied catalyst and ligand. One unknown compound was also present, but is not included in this table.

| Entry | Time [h] | Catalyst | (Eq.) | Ligand (Eq.) |  | Crude [\%] |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |  |  |  |
| 1 | 26 | $\mathrm{Pd}(\mathrm{OAc})_{2}$ | (0.06) | SPhos | (0.10) | 61 | 8 | 8 |
| 2 | 17 | $\mathrm{Pd}(\mathrm{OAc})_{2}$ | (0.03) | SPhos | (0.06) | 100 | 0 | 0 |
| 3 | 17 | $\mathrm{Pd}(\mathrm{OAc})_{2}$ | (0.04) | SPhos | (0.08) | 76 | 6 | 17 |
| 4 | 1 | $\mathrm{Pd}(\mathrm{OAc})_{2}$ | (0.04) | XPhos | (0.08) | 70 | 18 | 10 |
| 5 | 48 | $\mathrm{Pd}(\mathrm{dppf}) \mathrm{Cl}_{2}$ | (0.06) | - | - | 63 | 0 | 0 |
| 6 | 5 | $\mathrm{Pd}\left(\mathrm{PPh}_{3}\right)_{4}$ | (0.03) | - | - | 94 | 0 | 6 |

[^0]
### 5.2.12 Attempted Synthesis of 3-(5-Bromothiophen-2-yl)-2,5-dihexyl-6-(5-(10-(4-(2-(2-(2-methoxyethoxy)ethoxy) ethoxy)phenyl)-7-(pyren-1-yl)-10H-phenothiazin-3-yl)thiophen-2-yl)-2,5-dihydro-pyrrolo[3,4]pyrrole-1,4dione (17)



Compound $\mathbf{4}$ ( $50.0 \mathrm{mg}, 0.08 \mathrm{mmol}$ ), compound $12(55,9 \mathrm{mg}, 0.07 \mathrm{mmol}), \mathrm{K}_{2} \mathrm{CO}_{3}$ ( $33.1 \mathrm{mg}, 0.24 \mathrm{mmol}$ ), $\mathrm{Pd}(\mathrm{dppf}) \mathrm{Cl}_{2}(1.75 \mathrm{mg}, 0.01 \mathrm{mmol}), 1,4$-dioxane ( 2.00 mL ) and water $(2.00 \mathrm{~mL})$ were stirred for 1 hour at $80^{\circ} \mathrm{C}$. The color changed from purple to dark blue-purple during the reaction. The reaction was quenched with water $(10 \mathrm{~mL})$ and extracted with EtOAc $(3.40 \mathrm{~mL})$. The organic phase was washed with water ( 40 mL ) and dried with anhydrous $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was filtered and removed in vacuo. TLC showed seven spots and following eluent systems were used in silica gel column chromatography; chloroform, $\mathrm{Et}_{2} \mathrm{O}, \mathrm{EtOAc}, \mathrm{EtOAc}: H e x a n e$ (4:1). After the four different silica gel chromatography columns, TLC showed three different spots ( $\mathrm{R}_{f}=0,0.31,0.57, \mathrm{Et}_{2} \mathrm{O}$ ). The exact same reaction was done with a reaction time of 26 hours, which resulted in the same amounts of spots on TLC.

### 5.2.13 4-(5-(2,5-Dihexyl-3,6-dioxo-4-(thiophen-2-yl)-2,3,5,6-tetrahydropyrrolo[3,4]pyrrol-1-yl)thiophen-2-yl) benzaldehyde (13)




Compound 9 ( $150 \mathrm{mg}, 0.32 \mathrm{mmol}$ ), 4-bromobenzaldehyde ( $42.1 \mathrm{mg}, 0.27 \mathrm{mmol}$ ), $\mathrm{Pd}_{2}(\mathrm{dba})_{3}(24.4 \mathrm{mg}, 0.03 \mathrm{mmol})$, tris(o-anisyl)phosphine ( $18.8 \mathrm{mg}, 0.05 \mathrm{mmol}$ ), $\mathrm{K}_{2} \mathrm{CO}_{3}(95.6 \mathrm{mg}, 0.29 \mathrm{mmol})$, pivalic acid ( $5.45 \mathrm{mg}, 0.05 \mathrm{mmol}$ ) and dry toluene $(2.50 \mathrm{~mL})$ were stirred for 14 hours at $120^{\circ} \mathrm{C}$. The reaction was then cooled with water ( 40 mL ) and extracted with chloroform $(3.80 \mathrm{~mL})$. The organic phase was washed with water $(40 \mathrm{~mL})$ and dried with anhydrous $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was filtered and removed in vacuo. In total four silica gel chromatography columns were used. First one column ( 39 g silica, gradient hexane:EtOAc, 9:1, - hexane:EtOAc:DCM, 130:20:50) followed by three more ( 39 g silica, chloroform, $\mathrm{R}_{f}$ 0.80 ) isolated some product $\mathbf{1 3}$ after each column. In total $118.7 \mathrm{mg}(0.18 \mathrm{mmol}$, $68 \%$ ) was collected of product $\mathbf{1 3}$ as a dark purple solid, mp. 211.5-212.9 ${ }^{\circ} \mathrm{C}$.
${ }^{1} \mathrm{H} \operatorname{NMR}\left(600 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta: 10.03(\mathrm{~s}, 1 \mathrm{H}), 8.97(\mathrm{dd}, \mathrm{J}=3.8, \mathrm{~J}=1.0,1 \mathrm{H}), 8.94$ $(\mathrm{d}, \mathrm{J}=4.1,1 \mathrm{H}), 7.94(\mathrm{~d}, \mathrm{~J}=8.3,2 \mathrm{H}), 7.84(\mathrm{~d}, \mathrm{~J}=8.1,2 \mathrm{H}), 7.66(\mathrm{dd}, \mathrm{J}=5.0, \mathrm{~J}=$ $1.3,1 \mathrm{H}), 7.60(\mathrm{~d}, \mathrm{~J}=4.1,1 \mathrm{H}), 7.31-7.29(\mathrm{~m}, 1 \mathrm{H}), 4.13-4.07(\mathrm{~m}, 4 \mathrm{H}), 1.81-1.73$ $(\mathrm{m}, 4 \mathrm{H}), 1.49-1.41(\mathrm{~m}, 4 \mathrm{H}), 1.36-1.32(\mathrm{~m}, 8 \mathrm{H}), 0.91-0.88(\mathrm{~m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(150 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta: 191.2,161.4$ (2C), 147.3, 140.5, 139.0, 138.7, 136.3, 136.0, 135.6, 131.1, 130.7, 130.6, 129.7, 128.7, 126.4, 126.3, 108.3 (2C), 42.3 (2C), 31.4 (2C), 30.0 (2C), 26.6 (2C), 22.6 (2C), 14.0 (2C); IR (neat, $\mathrm{cm}^{-1}$ ) $\nu: 2953$ (w), 2854 (w), 1704 (m), 1653 (s), 1599 (m), 1556 (m/s), 1460 (w), 1401(m), 1336 (w), 1211 (m), 1170 (w), 1101 (w), $834(\mathrm{~m}), 730(\mathrm{~m}), 465(\mathrm{w})$; HRMS (APCI/ASAP+, m/z): found $573.2246\left([\mathrm{M}+\mathrm{H}]^{+}\right.$, calculated $\left.\mathrm{C}_{33} \mathrm{H}_{36} \mathrm{~N}_{2} \mathrm{O}_{3} \mathrm{~S}_{2}, 572.7791\right)$.

### 5.2.14 4-(5-(2,5-Dihexyl-3,6-dioxo-4-(thiophen-2-yl)-2,3,5,6-tetrahydropyrrolo[3,4]pyrrol-1-yl)thiophen-2-yl) benzaldehyde (15)



Following the reaction described in Section 5.2.13, compound $\mathbf{1 5}$ was also performed. Silica gel column chromatography ( 39 g silica, chloroform, $\mathrm{R}_{f}=0.68$ ) gave product 15 as a blue solid, $20.0 \mathrm{mg}(0.03 \mathrm{mmol}, 11 \%), \mathrm{mp} .290 .7-292.0^{\circ} \mathrm{C}$.
${ }^{1}{ }^{H}$ NMR ( $600 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta: 10.04(\mathrm{~s}, 2 \mathrm{H}), 8.98(\mathrm{~d}, \mathrm{~J}=4.1,2 \mathrm{H}), 7.95(\mathrm{~d}, \mathrm{~J}=$ $8.2,4 \mathrm{H}), 7.85(\mathrm{~d}, \mathrm{~J}=8.2,4 \mathrm{H}), 7.62(\mathrm{~d}, \mathrm{~J}=4.1,2 \mathrm{H}), 4.13(\mathrm{t}, \mathrm{J}=7.8,4 \mathrm{H}), 1.83-1.78$ $(\mathrm{m}, 4 \mathrm{H}), 1.49-1.46(\mathrm{~m}, 4 \mathrm{H}), 1.38-1.33(\mathrm{~m}, 8 \mathrm{H}), 0.90(\mathrm{t}, \mathrm{J}=7.0,6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $150 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta: 191.2$ (2C), 161.3 (2C), 147.8 (2C), 139.4 (2C), 138.6 (2C), 136.6 (2C), 136.0 (2C), 130.6 (2C), 130.5 (2C), 126.5 (2C), 126.4 (2C), 108.8 (2C), 42.4 (2C), 31.4 (2C), 30.0 (2C), 26.6 (2C), 22.6 (2C), 14.0 (2C); IR (neat, $\mathrm{cm}^{-1}$ ) $\nu: 2953$ (w), 2727 (w), 1701 (m), 1651 ( s$), 1551$ (m/s), 1401 (m), 1305 (w), $1238(\mathrm{~m}), 1023(\mathrm{w}), 805(\mathrm{~m} / \mathrm{s}), 728(\mathrm{~m}), 468(\mathrm{w}) ;$ HRMS (APCI/ASAP+, m/z): found $677.2508\left([\mathrm{M}+\mathrm{H}]^{+}\right.$, calculated $\left.\mathrm{C}_{40} \mathrm{H}_{40} \mathrm{~N}_{2} \mathrm{O}_{4} \mathrm{~S}_{2}, 676.8850\right)$.

### 5.2.15 4-(5-(2,5-Dihexyl-4-(5-(10-(4-(2-(2-(2-methoxyethoxy) ethoxy)ethoxy)phenyl)-7-(pyren-1-yl)-10H-phenothiazin-3-yl)thiophen-2-yl)-3,6-dioxo-2,3,5,6-tetrahydropyrrolo [3,4]pyrrol-1-yl)thiophen-2-yl)benzaldehyde (18)





Compound 2 ( $44.0 \mathrm{mg}, 0.08 \mathrm{mmol}$ ), compound $13(45.9 \mathrm{mg}, 0.06 \mathrm{mmol}), \mathrm{Pd}_{2}(\mathrm{dba})_{3}$ ( $5.86 \mathrm{mg}, 0.01 \mathrm{mmol}$ ), Tris(o-anisyl)phosphine ( $4.51 \mathrm{mg}, 0.01 \mathrm{mmol}$ ), $\mathrm{K}_{2} \mathrm{CO}_{3}$ $(22.9 \mathrm{mg}, 0.07 \mathrm{mmol})$, pivalic acid $(1.31 \mathrm{mg}, 0.01 \mathrm{mmol})$ and dry toluene $(1.00$ mL ) were stirred for 12 hours at $120^{\circ} \mathrm{C}$. The reaction was then quenched with water $(10 \mathrm{~mL})$ and extracted with chloroform $(3.25 \mathrm{~mL})$. The organic phase was washed with water ( 40 mL ) and dried with anhydrous $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was filtered and removed in vacuo. Silica gel column chromatography (two times 39 g silica, $\mathrm{CHCl}_{3}, 38 \mathrm{~g}$ silica, $\left.\mathrm{Et}_{2} \mathrm{O}\right)$ yielded product 18 as a blue solid, $56.8 \mathrm{mg}(0.05$ $\mathrm{mmol}, 73 \%, 79 \%$ pure $) . \mathrm{mp} .98 .1-100.0^{\circ} \mathrm{C}$.
${ }^{1} \mathrm{H}$ NMR ( $600 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta: 10.03(\mathrm{~s}, 1 \mathrm{H}), 9.01(\mathrm{~d}, \mathrm{~J}=4.2,1 \mathrm{H}), 8.91(\mathrm{~d}, \mathrm{~J}$ $=4.2,1 \mathrm{H}), 8.20-8.28(\mathrm{~m}, 3 \mathrm{H}), 8.16(\mathrm{~d}, \mathrm{~J}=7.5,1 \mathrm{H}), 8.08(\mathrm{~s}, 2 \mathrm{H}), 8.03(\mathrm{~d}, \mathrm{~J}=$ $9.2,1 \mathrm{H}), 8.01(\mathrm{t}, \mathrm{J}=7.6,1 \mathrm{H}), 7.93(\mathrm{~d}, \mathrm{~J}=8.4,2 \mathrm{H}), 7.91(\mathrm{~d}, \mathrm{~J}=7.8,1 \mathrm{H}), 7.83$ $(\mathrm{d}, \mathrm{J}=8.3,2 \mathrm{H}), 7.60(\mathrm{~d}, \mathrm{~J}=4.2,1 \mathrm{H}), 7.41(\mathrm{~d}, \mathrm{~J}=8.8,2 \mathrm{H}), 7.35(\mathrm{~d}, \mathrm{~J}=4.2,1 \mathrm{H})$, $7.33(\mathrm{~d}, \mathrm{~J}=2.1,1 \mathrm{H}), 7.29(\mathrm{~d}, \mathrm{~J}=2.0,1 \mathrm{H}), 7.21(\mathrm{~d}, \mathrm{~J}=8.9,2 \mathrm{H}), 7.19(\mathrm{dd}, \mathrm{J}=$ $8.7, \mathrm{~J}=2.1,1 \mathrm{H}), 7.12(\mathrm{dd}, \mathrm{J}=8.4, \mathrm{~J}=2.0,1 \mathrm{H}), 6.36(\mathrm{~d}, \mathrm{~J}=8.4,1 \mathrm{H}), 6.25(\mathrm{~d}, \mathrm{~J}$ $=8.7,1 \mathrm{H}), 4.26(\mathrm{t}, \mathrm{J}=4.8,2 \mathrm{H}), 4.14-4.09(\mathrm{~m}, 4 \mathrm{H}), 3.95(\mathrm{t}, \mathrm{J}=4.8,2 \mathrm{H}), 3.81$ $-3.79(\mathrm{~m}, 2 \mathrm{H}), 3.74-3.72(\mathrm{~m}, 2 \mathrm{H}), 3.70-3.68(\mathrm{~m}, 2 \mathrm{H}), 3.59-3.57(\mathrm{~m}, 2 \mathrm{H})$, $3.40(\mathrm{~s}, 3 \mathrm{H}), 1.82-1.75(\mathrm{~m}, 4 \mathrm{H}), 1.48-1.42(\mathrm{~m}, 4 \mathrm{H}), 1.38-1.29(\mathrm{~m}, 8 \mathrm{H}), 0.91-$ $0.88(\mathrm{~m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $150 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta: 191.4,161.5,161.2,158.9,149.8$, 147.0 (2C), 145.1, 143.0, 140.5, 138.8, 138.0, 137.5, 136.2, 135.9 (2C), 132.9, 132.0 (2C), 131.5, 131.0, 130.9, 130.6 (2C), 129.3, 128.8, 128.5, 127.6, 127.5, 127.4 (2C), 127.3, 127.2, 126.4 (3C), 126.0, 125.5, 125.1, 125.0 (3C), 124.9 (2C), 124.7, 123.8, 123.7, 120.4, 119.0, 116.9 (2C), 115.9, 115.8, 72.0, 71.0, 70.7, 70.3, 69.8, 67.9, 59.1, 42.3 (2C), 31.4 (2C), 30.1 (2C), 29.8 (2C), 26.6 (2C), 14.1 (2C); IR (neat, $\mathrm{cm}^{-1}$ ) $\nu: 3037$ (m), 2955 (s), 2855 (m), 1658 (s), 1731 (w), 1600 (m), 1555 (m), 1431 (m), 1310 (m), 1214(m), 1105 (w), 833 (w), 730 (w), 479 (w).

### 5.2.16 (z)-3-(5-(4-(2-((2-Azaneylidene)-3-methyl)-3-hydro-peroxy-32-prop-1-en-1-yl)phenyl)thiophen-2-yl)-2,5-dihexyl-6-(5-(10-(4-(2-(2-(2-methoxyethoxy)ethoxy)ethoxy) phenyl)-7-(pyren-1-yl)-10H-phenothiazin-3-yl)thiophen-2-yl)-2,5-dihydropyrrolo[3,4-c]pyrrole-1,4-dione (MEL3)



Compound 18 ( $44.0 \mathrm{mg}, 0.04 \mathrm{mmol}$ ), 2-cyanoacetic acid ( $61.9 \mathrm{mg}, 0.73 \mathrm{mmol}$ ) and acetonitrile ( 6.50 mL ) were stirred for 1 hours at reflux $\left(84^{\circ} \mathrm{C}\right)$. Acetonitrile $(13.0 \mathrm{~mL}), \mathrm{CHCl}_{3}(6.50 \mathrm{~mL})$ and piperidine $(0.06 \mathrm{~mL}, 0.44 \mathrm{mmol})$ were added and the reaction mixture was stirred for 7 hours, then allowed to reach room temperature. The reaction was quenched with acidic water ( $120 \mathrm{~mL}, 2 \mathrm{M} \mathrm{HCl}$ ), and stirred for 30 minutes. The mixture was extracted with $\mathrm{CHCl}_{3}(3.30 \mathrm{~mL})$ and dried over anhydrous $\mathrm{Na}_{2} \mathrm{SO}_{4}$. The solvent was filtered and removed in vacuo. Silica gel column chromatography ( 38 g silica, gradient $\mathrm{DCM}-\mathrm{DCM}: \mathrm{MeOH}, 5: 1$ ) gave product MEL3 as a blue solid, $46.4 \mathrm{mg}(0.04 \mathrm{mmol}, 88 \%), \mathrm{mp} .133 .2-134.7^{\circ} \mathrm{C}$.

IR (neat, $\mathrm{cm}^{-1}$ ) $\nu: 3037$ (m), 2916 ( s$), 2850(\mathrm{~s}), 2214$ (w), 1701 ( s$), 1652(\mathrm{~m} / \mathrm{s})$, 1555 (m), 1423 (s), 1307 ( s ), 1242 ( s ), 1081 (m/s), 941 (m), 844 (m), 743 (s), $666(\mathrm{~m} / \mathrm{s}), 555(\mathrm{~m}), 428(\mathrm{w})$; HRMS (APCI/ASAP+, m/z): found $1231.4536([\mathrm{M}-$ $\mathrm{COO}+\mathrm{H}]^{+}$, calculated $\mathrm{C}_{76} \mathrm{H}_{71} \mathrm{~N}_{4} \mathrm{O}_{6} \mathrm{~S}_{3}$, 1231.5844).

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## Appendix

## A Spectroscopic Data - Compound 2



Figure 1: ${ }^{1} \mathrm{H}$ NMR spectrum of compound 2.

## B Spectroscopic Data - Compound 3



Figure 2: ${ }^{1} \mathrm{H}$ NMR spectra of compound 3.

## C Spectroscopic Data - Compound 4



Figure 3: ${ }^{1} \mathrm{H}$ NMR spectra of compound 4.


Figure 4: ${ }^{13} \mathrm{C}$ NMR spectra of compound 4.


Figure 5: COSY NMR spectra of compound 4.


Figure 6: HSQC NMR spectra of compound 4.


Figure 7: HMBC NMR spectra of compound 4.


Figure 8: IR spectra of compound 4.

## Elemental Composition Report

Page 1
Single Mass Analysis
Tolerance $=2.0$ PPM / DBE: $\min =-50.0, \max =50.0$
Element prediction: Off
Number of isotope peaks used for i-FIT = 3
Monoisotopic Mass, Even Electron Ions
15419 formula(e) evaluated with 15 results within limits (all results (up to 1000) for each mass)
Elements Used:
$\begin{array}{llllll}\mathrm{C}: 0-100 & \mathrm{H}: 0-150 & 11 B: 0-1 & \mathrm{~N}: 0-10 & \mathrm{O}: 0-10 & \mathrm{~S}: 0-4\end{array}$
2019-480 297 (5.790) AM2 (Ar,35000.0,0.00,0.00); Cm (280:300)
2019-480 297 (5.790)
1: TOF MS ASAP +
$4.83 \mathrm{e}+006$



Figure 9: Mass spectra of compound 4.

## D Spectroscopic Data - Compound 5



Figure 10: ${ }^{1} \mathrm{H}$ NMR spectra of compound 5.


Figure 11: ${ }^{13} \mathrm{C}$ NMR spectra of compound 5.




Figure 12: COSY NMR spectra of compound 5.


Figure 13: HSQC NMR spectra of compound 5.


Figure 14: HMBC NMR spectra of compound 5.


Figure 15: IR spectra of compound 5.

Single Mass Analysis
Tolerance $=2.0$ PPM / DBE: $\min =-50.0, \max =50.0$
Element prediction: Off
Number of isotope peaks used for i-FIT = 3
Monoisotopic Mass, Even Electron Ions
7663 formula(e) evaluated with 11 results within limits (all results (up to 1000) for each mass)
Elements Used:
$\begin{array}{lllll}\text { C: } 0-100 & \mathrm{H}: 0-150 & \mathrm{~N}: 0-10 & \mathrm{O}: 0-10 & \mathrm{~S}: 0-4\end{array}$
2019-483 297 (5.791) AM2 (Ar,35000.0,0.00,0.00); Cm (279:300)
1: TOF MS ASAP+


Figure 16: Mass spectra of compound 5.

## E Spectroscopic Data - Compound 6



Figure 17: ${ }^{1} \mathrm{H}$ spectrum of compound 6.


Figure 18: ${ }^{13} \mathrm{C}$ NMR spectra of compound 6.


Figure 19: COSY NMR spectra of compound 6.


Figure 20: HSQC NMR spectra of compound 6.


Figure 21: HMBC NMR spectra of compound 6.


Figure 22: IR spectra of compound 6.

Single Mass Analysis
Tolerance $=2.0$ PPM / DBE: $\min =-50.0, \max =50.0$
Element prediction: Off
Number of isotope peaks used for i-FIT = 3
Monoisotopic Mass, Even Electron Ions
7663 formula(e) evaluated with 10 results within limits (all results (up to 1000) for each mass)
Elements Used:
C: 0-100 $\quad \mathrm{H}: 0-150 \quad \mathrm{~N}: 0-10 \quad \mathrm{O}: 0-10 \quad \mathrm{~S}: 0-4$
2019-438 139 (1.549) AM2 (Ar,35000.0,0.00,0.00); Cm (135:139)
1:TOF MS ES +


Figure 23: Mass spectra of compound 6.

## F Spectroscopic Data - Compound MEL1



Figure 24: ${ }^{1} \mathrm{H}$ spectrum of compound MEL1.


Figure 25: ${ }^{13} \mathrm{C}$ NMR spectra of compound MEL1.


Figure 26: COSY NMR spectra of compound MEL1.


Figure 27: HSQC NMR spectra of compound MEL1.


Figure 28: HMBC NMR spectra of compound MEL1.


Figure 29: IR spectra of compound MEL1.

## Elemental Composition Report

Page 1
Single Mass Analysis
Tolerance $=1.0$ PPM / DBE: $\min =-50.0, \max =50.0$
Element prediction: Off
Number of isotope peaks used for i-FIT = 3
Monoisotopic Mass, Even Electron Ions
7773 formula(e) evaluated with 6 results within limits (all results (up to 1000) for each mass)
Elements Used:
$\begin{array}{lllll}\text { C: } 0-100 & \mathrm{H}: 0-150 & \mathrm{~N}: ~ 0-10 & \mathrm{O}: 0-10 & \mathrm{~S}: 0-4\end{array}$
2019-484 297 (5.791) AM2 (Ar,35000.0,0.00,0.00); Cm (281:302)


Minimum:

|  |  |  |
| :--- | :--- | :--- |
| 5.0 | 1.0 | 50.0 |

Mass Calc. Mass mDa PPM DBE i-FIT Norm Conf(\%) Formula
$\begin{array}{llllllllllll}771.2348 & 771.2351 & -0.3 & -0.4 & 30.5 & 659.2 & 1.706 & 18.16 & \text { C48 H39 N2 } 04\end{array}$

|  |  |  |  |  |  |  | S2 |  |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| 771.2343 | 0.5 | 0.6 | 31.5 | 657.9 | 0.462 | 62.99 | C47 H35 N2 09 |  |  |
| 771.2349 | -0.1 | -0.1 | 27.5 | 661.5 | 4.009 | 1.81 | C40 H35 N8 07 S |  |  |
| 771.2345 | 0.3 | 0.4 | 21.5 | 659.4 | 1.956 | 14.15 | C40 H43 N4 06 |  |  |
|  |  |  |  |  |  |  | S3 |  |  |
| 771.2352 | -0.4 | -0.5 | 17.5 | 661.1 | 3.647 | 2.61 | C33 H43 N10 04 |  |  |
|  |  |  |  |  |  |  | S4 |  |  |
| 771.2343 | 0.5 | 0.6 | 18.5 | 663.3 | 5.871 | 0.28 | C32 H39 N10 09 |  |  |

Figure 30: Mass spectra of compound MEL1.

## G Spectroscopic Data - Compound MEL2



Figure 31: ${ }^{1} \mathrm{H}$ spectrum of compound MEL2.


Figure 32: ${ }^{13} \mathrm{C}$ NMR spectra of compound MEL2.


Figure 33: COSY NMR spectra of compound MEL2.


Figure 34: HSQC NMR spectra of compound MEL2.


Figure 35: HMBC NMR spectra of compound MEL2.


Figure 36: IR spectra of compound MEL2.

## Elemental Composition Report

Page 1
Single Mass Analysis
Tolerance $=2.0$ PPM / DBE: $\min =-50.0, \max =50.0$
Element prediction: Off
Number of isotope peaks used for i-FIT = 3
Monoisotopic Mass, Even Electron Ions
7755 formula(e) evaluated with 13 results within limits (all results (up to 1000) for each mass)
Elements Used:
C: 0-100 $\quad \mathrm{H}: 0-150 \quad \mathrm{~N}: 0-10 \quad \mathrm{O}: 0-10 \quad \mathrm{~S}: 0-4$
2019-479 289 (5.635) AM2 (Ar,35000.0,0.00,0.00); Cm (281:301)


| Minimum: |  |  | -50.0 |
| :--- | :--- | :--- | :--- |
| Maximum: | 5.0 | 2.0 | 50.0 |


| Mass | Calc. Mass | mDa | PPM | DBE | i-FIT | Norm | Conf(\%) | Formula |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 853.2237 | 853.2235 | 0.2 | 0.2 | 42.5 | 445.3 | 2.160 | 11.53 | C60 | H37 | 02 | S2 |
|  | 853.2240 | -0.3 | -0.4 | 48.5 | 444.2 | 1.060 | 34.63 | C60 | H29 | N4 | 03 |
|  | 853.2226 | 1.1 | 1.3 | 43.5 | 444.6 | 1.468 | 23.03 | C59 | H33 | 07 |  |
|  | 853.2247 | -1.0 | -1.2 | 44.5 | 445.9 | 2.801 | 6.08 | C53 | H29 | N10 | 0 S |
|  | 853.2242 | -0.5 | -0.6 | 38.5 | 445.9 | 2.764 | 6.31 | C53 | H37 | N6 | S3 |
|  | 853.2228 | 0.9 | 1.1 | 33.5 | 446.2 | 3.132 | 4.36 | C52 | H41 | N2 | 04 |
|  |  |  |  |  |  |  |  | S3 |  |  |  |
|  | 853.2233 | 0.4 | 0.5 | 39.5 | 446.2 | 3.059 | 4.70 | C52 | H33 | N6 | 05 S |
|  | 853.2253 | -1.6 | -1.9 | 29.5 | 446.7 | 3.589 | 2.76 | C48 | H41 | N2 | 09 |
|  |  |  |  |  |  |  |  | S2 |  |  |  |
|  | 853.2235 | 0.2 | 0.2 | 29.5 | 446.7 | 3.609 | 2.71 | C45 | H41 | N8 | 02 |
|  |  |  |  |  |  |  |  | S4 |  |  |  |
|  | 853.2227 | 1.0 | 1.2 | 30.5 | 447.5 | 4.430 | 1.19 | C44 H37 N8S2 |  |  |  |
|  |  |  |  |  |  |  |  |  |  |  |  |
|  | 853.2222 | 1.5 | 1.8 | 24.5 | 447.2 | 4.051 | 1.74 | C44 H45 N4 06S 4 |  |  |  |
|  |  |  |  |  |  |  |  |  |  |  |  |
|  | 853.2220 | 1.7 | 2.0 | 21.5 | 448.6 | 5.465 | 0.42 | C36 H41 N10S3 |  |  |  |
|  |  |  |  |  |  |  |  |  |  |  |  |
|  | 853.2254 | -1.7 | -2.0 | 16.5 | 448.3 | 5.210 | 0.55 | C33 | H45 | N10 | 09 |
|  |  |  |  |  |  |  |  | S4 |  |  |  |

Figure 37: Mass spectra of compound MEL2.

## H Spectroscopic Data - Compound 9



Figure 38: ${ }^{1} \mathrm{H}$ NMR spectra of compound 9 .

## I Spectroscopic Data - Compound 10



Figure 39: ${ }^{1} \mathrm{H}$ NMR spectra of compound $\mathbf{1 0}$.

## J Spectroscopic Data - Compound 12



Figure 40: ${ }^{1} \mathrm{H}$ NMR spectra of compound 12.

## K Spectroscopic Data - Compound 16



Figure 41: ${ }^{1} \mathrm{H}$ NMR spectra of the impure compound 16.

## L Spectroscopic Data - Compound 13



Figure 42: ${ }^{1} \mathrm{H}$ spectrum of compound 13.


Figure 43: ${ }^{13} \mathrm{C}$ NMR spectra of compound 13 .


Figure 44: COSY NMR spectra of compound 13.


Figure 45: HSQC NMR spectra of compound 13.


Figure 46: HMBC NMR spectra of compound 13.


Figure 47: IR spectra of compound 13.

## Elemental Composition Report

Page 1
Single Mass Analysis
Tolerance $=2.0$ PPM / DBE: $\min =-50.0, \max =50.0$
Element prediction: Off
Number of isotope peaks used for i-FIT = 3
Monoisotopic Mass, Even Electron Ions
7466 formula(e) evaluated with 9 results within limits (all results (up to 1000) for each mass)
Elements Used:
$\begin{array}{lllll}\text { C: } 0-100 & \mathrm{H}: 0-150 & \mathrm{~N}: 0-10 & \mathrm{O}: 0-10 \quad \mathrm{~S}: 0-4\end{array}$
2019-481 239 (4.652) AM2 (Ar,35000.0,0.00,0.00); Cm (225:239)


| Minimum: <br> Maximum: |  | 5.0 | 2.0 | $\begin{aligned} & -50.0 \\ & 50.0 \end{aligned}$ |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Mass | Calc. Mass | mDa | PPM | DBE | i-FIT | Norm | Conf(\%) | Formula |  |
| 573.2240 | 573.2244 | -0.4 | -0.7 | 13.5 | 1112.8 | 0.087 | 91.71 | C25 H33 | N8 06 S |
|  | 573.2230 | 1.0 | 1.7 | 8.5 | 1115.5 | 2.824 | 5.94 | $\begin{aligned} & \mathrm{C} 24 \mathrm{H} 37 \\ & \mathrm{~S} \end{aligned}$ | N4 010 |
|  | 573.2246 | -0.6 | -1.0 | 16.5 | 1116.5 | 3.761 | 2.33 | C33 H37 | N2 03 |
|  |  |  |  |  |  |  |  | S2 |  |
|  | 573.2237 | 0.3 | 0.5 | 4.5 | 1121.5 | 8.850 | 0.01 | C17 H37 | N10 08 |
|  |  |  |  |  |  |  |  | S2 |  |
|  | 573.2239 | 0.1 | 0.2 | 7.5 | 1122.2 | 9.500 | 0.01 | C25 H41 | N4 05 |
|  |  |  |  |  |  |  |  | S3 |  |
|  | 573.2237 | 0.3 | 0.5 | 17.5 | 1123.4 | 10.667 | 0.00 | C32 H33 | N2 08 |
|  | 573.2250 | -1.0 | -1.7 | 22.5 | 1124.2 | 11.471 | 0.00 | C33 H29 | N6 04 |
|  | 573.2246 | -0.6 | -1.0 | 3.5 | 1125.2 | 12.515 | 0.00 | C18 H41 | N10 03 |
|  |  |  |  |  |  |  |  | S4 |  |
|  | 573.2233 | 0.7 | 1.2 | -1.5 | 1126.1 | 13.378 | 0.00 | C17 H45 | N6 07 |
|  |  |  |  |  |  |  |  | S4 |  |

Figure 48: Mass spectra of compound 13.

## M Spectroscopic Data - Compound 15



Figure 49: ${ }^{1} \mathrm{H}$ spectrum of compound 15.


Figure 50: ${ }^{13} \mathrm{C}$ NMR spectra of compound 15.


Figure 51: COSY NMR spectra of compound 15.


Figure 52: HSQC NMR spectra of compound 15.


Figure 53: HMBC NMR spectra of compound 15.


Figure 54: IR spectra of compound 15.

## Elemental Composition Report

Page 1
Single Mass Analysis
Tolerance $=2.0$ PPM / DBE: $\min =-50.0, \max =50.0$
Element prediction: Off
Number of isotope peaks used for i-FIT = 3
Monoisotopic Mass, Even Electron Ions
7730 formula(e) evaluated with 10 results within limits (all results (up to 1000) for each mass)
Elements Used:
C: 0-100 $\quad \mathrm{H}: 0-150 \quad \mathrm{~N}: 0-10 \quad \mathrm{O}: 0-10 \quad \mathrm{~S}: 0-4$
2019-482 300 (5.842) AM2 (Ar,35000.0,0.00,0.00); Cm (283:302)
$\begin{array}{ll}1: \text { TOF MS ASAP }+ & 1.57 \mathrm{e}+007\end{array}$


Figure 55: Mass spectra of compound 15.

## N Spectroscopic Data - Compound 18



Figure 56: ${ }^{1} \mathrm{H}$ spectrum of compound 18.


Figure 57: ${ }^{13} \mathrm{C}$ NMR spectra of compound 18.


Figure 58: COSY NMR spectra of compound 18.


Figure 59: HSQC NMR spectra of compound 18.


Figure 60: HMBC NMR spectra of compound 18.


Figure 61: IR spectra of compound 18.

## O Spectroscopic Data - Compound MEL3



Figure 62: IR spectra of compound MEL3.

## Elemental Composition Report

Page 1

## Single Mass Analysis

Tolerance $=2.0$ PPM / DBE: $\min =-50.0, \max =50.0$
Element prediction: Off
Number of isotope peaks used for i-FIT = 3
Monoisotopic Mass, Even Electron Ions
3956 formula(e) evaluated with 2 results within limits (all results (up to 1000) for each mass)
Elements Used:
$\begin{array}{lllll}\text { C: } 2-100 & \text { H: 0-150 } & \text { N: 0-6 } & \text { O: 0-10 } & \text { S: 0-3 }\end{array}$
2019_556_20190605re 28 ( 0.321 ) AM2 (Ar,35000.0,0.00,0.00); ABS; Cm (17:28)
1: TOF MS ES +
$4.34 \mathrm{e}+004$


Figure 63: Mass spectra of compound MEL3.

Kunnskap for en bedre verden


[^0]:    ${ }^{a}$ Starting material, 4-formylphenylboronic acid. Detected by ${ }^{1} \mathrm{H}$ NMR analysis.

