Hollow core-shell structured Cu<sub>2</sub>O@Cu<sub>1.8</sub>S spheres as novel

electrode for non-enzymatic glucose sensing

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Abstract

This study reports a novel hollow Cu<sub>2</sub>O@Cu<sub>1.8</sub>S material used as an electrode for

non-enzymatic glucose sensing. Cu<sub>2</sub>O@Cu<sub>1.8</sub>S was successfully synthesized by a

facile in-situ growth method. The obtained Cu<sub>2</sub>O@Cu<sub>1.8</sub>S exhibited a hollow structure

with a Cu<sub>1.8</sub>S rich surface. Electrochemical results revealed that the Cu<sub>2</sub>O@Cu<sub>1.8</sub>S

electrode exhibited a much higher electrocatalytic activity toward glucose oxidation

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than Cu<sub>2</sub>O spheres owing to the synergistic effect between Cu<sub>2</sub>O and Cu<sub>1.8</sub>S. The Cu<sub>2</sub>O@Cu<sub>1.8</sub>S based sensor showed a rapid sensing response of 5 s, a wide linear range in concentrations of 1–1,000  $\mu$ M, a high sensitivity of 3,630  $\mu$ A·mM<sup>-1</sup>cm<sup>-2</sup>, and a low detection limit of 0.0678  $\mu$ M, along with an excellent selectivity, thus leading to a promising candidate for non-enzymatic glucose detection.

**Keywords:** Copper sulfide; Copper oxide; Hollow structure; Glucose; Non-enzymatic sensor

### 1. Introduction

Considerable efforts have been devoted to the development of fast, reliable and simple methods for monitoring glucose in the fields of clinical diagnostics, food industry and biotechnology [1]. The first enzyme glucose biosensor was reported in 1962 [2], and since then, glucose oxidase has attracted much research interest in the development of enzyme glucose biosensors due to its high sensitivity and activity [3-5]. Unfortunately, due to the high cost, complicated immobilization procedure, the glucose oxidase instability and poor reproducibility (as the catalytic activity can be easily affected by, for example, pH, temperature and humidity), extensive efforts have focused on the development of non-enzymatic glucose sensors to replace glucose oxidase biosensors [4, 6, 7]. Among these non-enzymatic glucose sensors, electrochemical non-enzymatic glucose sensors are attractive due to their low cost, high efficiency and ease of operation. Various non-enzymatic electrode materials, such as noble metal [8, 9], metal oxides [10, 11], metal sulfide and their hybrids [12, 13], have been widely developed for glucose detection.

It is well-known that copper oxide is a *p*-type semi-conductor material with great potential for use as a non-enzymatic sensor for detecting glucose due to its low cost, environmental friendliness and high catalytic activity [14-18]. However, copper oxides cannot be used as non-invasive glucose detection in saliva for routine glucose monitoring because of their poor linear detection range [19, 20]. In order to improve the detection ability, intensive R&D efforts have been devoted to combine copper oxide nanoparticles with other materials, such as metal, carbon materials and metal oxides [19, 21-24]. It has been reported that the fast response, low detection limit and excellent sensitivity for glucose detection can be achieved by combining copper oxides with other materials, because the combination can significantly improve electrical conductivity or/and synergistic effect. Thus, developing copper oxide hybrid materials can be an effective way to improve detection performance.

Copper sulfide possesses a variety of crystal polymorphs and structures. Among these copper sulfides, Cu<sub>2</sub>S and CuS have been developed as non-enzymatic glucose sensors, which show fast response, low detection limit and good sensitivity for glucose detection. Thus, it is expected that the highly sensitive detection of non-enzymatic glucose sensor could be achieved by forming hybrid structures which comprise copper oxide and copper sulfide. A literature search revealed that there is no report demonstrating the use of hybrid copper sulfide as sensors for glucose detection. In this study, a facile method was developed to synthesize hollow Cu<sub>2</sub>O@Cu<sub>1.8</sub>S spheres with Cu<sub>1.8</sub>S rich surface via an in-situ growth method. The results indicated

that the as-prepared samples exhibited a much better performance for glucose detection in terms of sensitivity, detection range and selectivity than Cu<sub>2</sub>O.

## 2. Experimental

## 2.1 Preparation of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S

All chemicals used throughout the experiments were of analytical grade (AR) and used without further purification. The first step of preparation of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S was to synthesize Cu<sub>2</sub>O; the detailed procedure is as follows: 0.3993g of (CH<sub>3</sub>COO)<sub>2</sub>Cu·H<sub>2</sub>O were firstly dissolved in 25 ml of DMF containing 0.3 ml deionized water and magnetically stirred for 10 min, then ultrasonically treated for 3 min. After that, the solution was stirred at 85 °C for 10 min and then left at room temperature for 2 h. The obtained product was washed thoroughly with ethanol (95%), collected by centrifuge and dried in a vacuum oven for 12 h at 40°C. Subsequently, the obtained Cu<sub>2</sub>O nanospheres (25 mg) were added into 15 ml of ultrapure water and ultrasonically treated for 5 min. The Cu<sub>2</sub>O solution was introduced into a Na<sub>2</sub>S solution (Na<sub>2</sub>S·9H<sub>2</sub>O was dissolved into 10 ml distilled water, and a molar ratio of Na<sub>2</sub>S·9H<sub>2</sub>O:CuO was 1.5:1), and then treated in an ice cooled ultrasonic bath for 5 min. A black product was formed during the ultrasonic treatment. The obtained black product was washed with 95% ethanol, collected by centrifuge and then dried in a vacuum oven at 40 °C for 12 h. The final product was labelled as Cu<sub>2</sub>O@Cu<sub>1.8</sub>S.

#### 2.2 Characterization

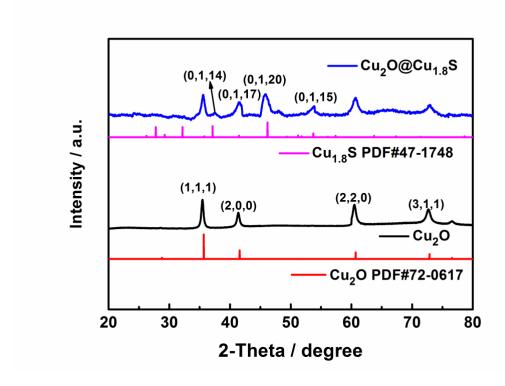
The crystalline structures of the samples were analyzed using X-ray diffraction (XRD, Shimadzu XD-3A (Japan) goniometer, using CuKα radiation operated at 40 kV and 35 mA). The morphology of the catalysts was observed by a Carl Zeiss Ultra Plus field emission scanning electron microscopy (SEM) and transmission electron microscopy (TEM) measurement which was carried out using a JEM-2010 Electron Microscope (Japan) with an acceleration voltage of 200 kV. Spectroscopy (XPS) tests were performed on a PHI-5702 (American). Binding energies were calibrated by referencing to the C1s peak at 285.0 eV.

### 2.3 Electrochemical measurements

Electrochemical measurements were carried out on an electrochemical work station (CHI 650D). A conventional three-electrode electrochemical cell was used comprising of a platinum wire as counter electrode, an Ag/AgCl (saturated KCl solution) as reference electrode, and a working electrode made of a thin film catalyst layer mounted on a 5-mm diameter glassy carbon disc (rotating disc electrode – *rde*). The thin film was prepared as follows: 2 mg of catalyst were dispersed ultrasonically in a 0.4 mL of Nafion®/ethanol solution (25 wt.% Nafion®). 8 μL of the above solution was transferred onto the glassy carbon and then dried in air. Cyclic voltammetry (CV) measurement was carried out in a three-electrode system using 0.1 M NaOH aqueous solution as electrolyte. The potential scan rate was 50 mV s<sup>-1</sup> in the potential range 0.20 - 0.80 V *vs.* Ag/AgCl. The electrode was firstly scanned in NaOH in the absence of glucose, subsequently scanned in NaOH with 50 μM, 100 μM, 500

μM, 1 mM, 2 mM, 3 mM, 4 mM and 5 mM of glucose respectively. Amperometric responses of the as-prepared electrodes with increasing glucose concentration was carried out in a 0.1 M NaOH solution and the rotation speed of the *rde* was set at 1,000 rpm. The selectivity of the electrodes was firstly tested in 0.1 M NaOH with 1,000 μM of glucose and then 0.1 mM of ascorbic acid (AA), uric acid (UA), NaCl and L-glucose was introduced into the NaOH/glucose solution respectively.

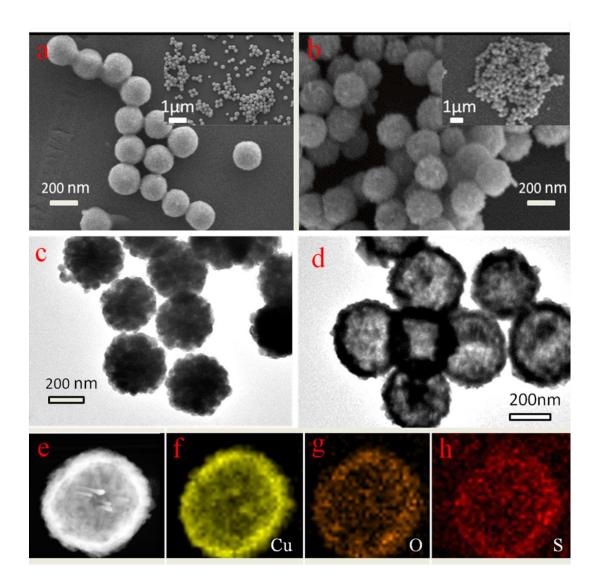
# 3. Results and discussion



**Figure 1.** XRD patterns of the Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S samples.

Figure 1 shows the XRD patterns of Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S. In the case of Cu<sub>2</sub>O, the diffraction peaks appear at  $2\theta = 35.4$ , 41.5, 60.5, 72.7 and  $76.6^{\circ}$ , corresponding to (110), (111), (200), (220), and (311) crystal planes of cubic Cu<sub>2</sub>O (PDF#72-0617), respectively. No diffraction peak of CuO was found, indicating that only a single

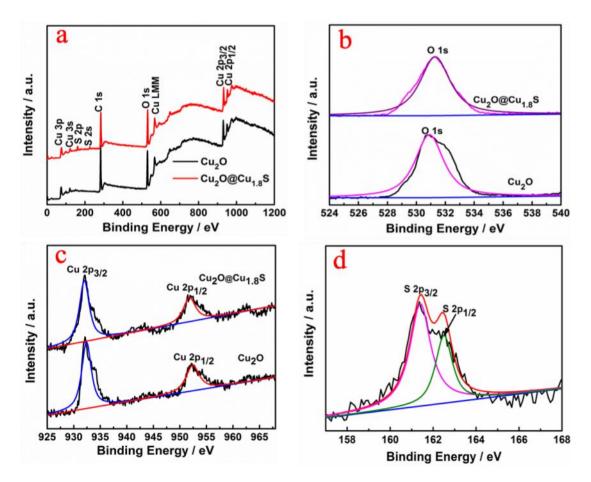
phase Cu<sub>2</sub>O was obtained. In the XRD pattern of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S, beside the diffraction peaks of Cu<sub>2</sub>O, four obvious diffraction peaks at 2θ degree of 37.6 45.8, 48.0 and 54.0° were observed. By carefully analyzing the data and critically reviewing the previous database, these diffraction peaks can be indexed as (0114), (0117), (0120), and (1115) planes of rhombohedral Cu<sub>1.8</sub>S (JCPDS No. 47-1748). The results indicated that Cu<sub>2</sub>O@Cu<sub>1.8</sub>S exhibited heterogeneous crystal structures.



**Figure 2.** SEM and TEM images and of the Cu<sub>2</sub>O (a,c) and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S (b,d); STEM (e) and corresponding EELS element maps (f-h) of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S.

The morphology of the as-prepared samples was studied by SEM and TEM. Figure

2(a) shows the overall morphology of the samples composed of large-scale uniform, sphere-like architectures of ca. 300 nm in diameter. After the in-situ formation of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S, the sphere shape was retained and the diameter increased to ca. 350 nm (Fig. 2(b)). The detail structure was investigated by TEM (Fig. 2(c)). The TEM image of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S clearly shows that the Cu<sub>2</sub>O spheres was made up of small particles. TEM images of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S, as represented in Figure 2(d), shows a hollow sphere-like structure assembled by small particles. According to He and Zhu et al.[25, 26], the formation of Cu<sub>1.8</sub>S could be ascribed to the exchange reaction between S<sup>2</sup>- and oxygen anions. Moreover, the mobility of oxygen anions was faster than that of the sulfur, leading to a continuous mass relocation of Cu<sub>2</sub>O crystallites from the inside out during the formation of Cu<sub>1.8</sub>S with a continuous supply of S<sup>2</sup>- for the exchange reaction, resulting in the hollow structure of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S. Elemental distribution in Cu<sub>2</sub>O@Cu<sub>1.8</sub>S was evaluated by HAADF-STEM (Fig. 2(e)) and corresponding EELS element concentration maps (Fig. 2(f-h)). Comparing the STEM and the elemental maps, the formation of a hollow structure is again confirmed. Moreover, the slight increase in the S diameter compared to that of O suggests the formation of Cu<sub>1.8</sub>S-rich surface.



**Figure 3.** (a) XPS survey spectra of Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S; (b) XPS spectra of Cu<sub>2</sub>O region in Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S; (c) XPS spectra of O 1s region in Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S; (d) XPS spectra of S2p region in Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S dendrite

XPS analyses were conducted to further investigate the chemical state and electronic state of the elements in Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S. A survey scan (Fig. 3(a)) indicates the presence of Cu and O in Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S, and the additional S signals in Cu<sub>2</sub>O@Cu<sub>1.8</sub>S. The detailed element contents were determined from XPS analyses. They revealed that the atomic ratio of Cu to O was 1.95:1 for Cu<sub>2</sub>O, close to 2, suggesting the formation of Cu<sub>2</sub>O, a finding which is in good agreement with the XRD results. In the case of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S, the S content was ca. 22.7 at.%. After

calculation, the molar ratio of Cu<sub>2</sub>O to Cu<sub>1.8</sub>S was found to be 1:2. Furthermore, as shown in Figure 3(b), the Cu 2*p* peak of Cu<sub>2</sub>O can be split into Cu 2*p*3/2 at 932.4 eV and Cu 2*p*1/2 at 952.4 eV, which is approximately consistent with the peak positions of Cu 2*p* spectra of reference Cu<sub>2</sub>O [27], further confirming the formation of Cu<sub>2</sub>O. A more detailed analysis in the spectra (Fig. 3(b)) indicates a slight shift to lower energy for Cu 2*p*3/2 (932.0 eV) and Cu 2*p*1/2 (951.8 eV) peaks of Cu<sub>1.8</sub>S compared to those of Cu<sub>2</sub>O, which are in good agreement with values found in the literature [28, 29]. It was also found that a shift in binding energy occurred as shown in the O 1*s* spectra (Fig. 3(c)); i.e. a binding energy of O 1*s* positively shifted from 530.9 eV in Cu<sub>2</sub>O to 531.3 eV in Cu<sub>2</sub>O@Cu<sub>1.8</sub>S, indicating the presence of S [28, 29]. In addition, the S 2*p* peak can be split into two doublets centered at 161.4 and 162.5 eV, corresponding to the sulfides of the multivalent copper [29].

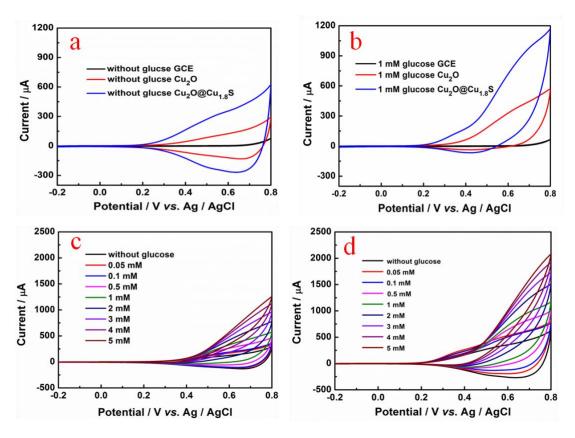


Figure 4. CV curves of Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S in 0.1 M NaOH (a) and 0.1 M NaOH + 0.1 mM glucose solution (b) at a scan rate of 50 mV s<sup>-1</sup>; CV curves of Cu<sub>2</sub>O (c) and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S (d) in 0.1 M NaOH + 0.05, 0.1, 0.5, 1, 2, 3, 4 and 5 mM glucose solution at a scan rate of 50 mV s<sup>-1</sup>.

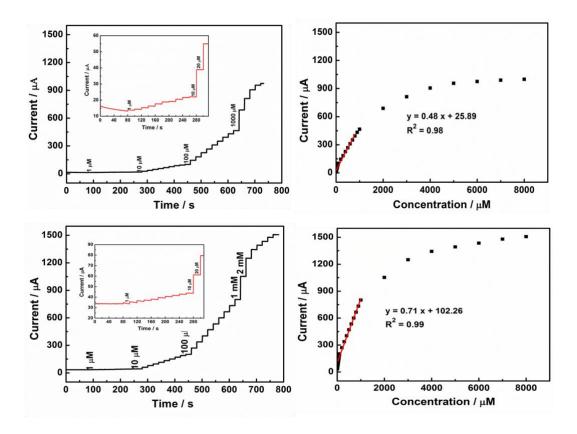
The electrochemical behaviour of Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S was firstly investigated by cyclic voltammetry (CV) in a 0.1 mol L<sup>-1</sup> KOH solution in the absence and presence of glucose (Fig. 4). As shown in Figure 4(a), the CVs on the glassy carbon disc electrode does not shows any redox peaks in the working potential range. The same observation is also found in Figure 4(b) when glucose is present, indicating that the bare glassy carbon disc electrode is not electrochemically active toward glucose. After Cu<sub>2</sub>O was loaded onto the electrode and immersed in 0.1 mol L<sup>-1</sup> NaOH, the CV showed an oxidation peak in the range +0.20 to 0.80 V vs. Ag/AgCl, corresponding to

the oxidation of Cu(I); the reverse reaction (reduction back to  $Cu_2O$ ) was also observed in the negative potential scan. The redox reactions can be described as follows [30-32]:

$$Cu2O + 2OH- \rightarrow 2CuO + H2O + 2e-$$
 (1)

$$CuO + OH^- \rightarrow CuOOH + e^-$$
 (2)

In our conditions, it is possible that the electron transfer between Cu (II) and Cu (III) could facilitate electron transfer during the oxidation of glucose. In the CVs of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S, it can be observed that the redox peaks shift negatively compared to Cu<sub>2</sub>O, suggesting that the electron transfer between Cu (I) and Cu (III) is easier. In the presence of glucose (Fig. 4(b)), the current increased compared to that in the absence of glucose, indicating that Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S have a catalytic activity toward glucose oxidation [33, 34]. It can also be observed that the current generated by the Cu<sub>2</sub>O@Cu<sub>1.8</sub>S electrode was larger than that of Cu<sub>2</sub>O, suggesting that the glucose oxidation occurred easily on Cu<sub>2</sub>O@Cu<sub>1.8</sub>S. Figure 4(c,d) shows that with increasing glucose concentration over a range of 0.05, 0.1, 0.5, 1, 2, 3, 4 and 5 mM, the activities of Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S increased linearly, indicating that specific electrocatalytic activity toward glucose oxidation had occurred. Moreover, it was found that all current responses on Cu<sub>2</sub>O@Cu<sub>1.8</sub>S were larger than those obtained on Cu<sub>2</sub>O, suggesting that Cu<sub>2</sub>O@Cu<sub>1.8</sub>S has good catalytic activity for glucose oxidation compared to Cu<sub>2</sub>O.



**Figure 5.** (a) Amperometric responses on Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S electrodes with successive addition of different amounts of glucose in 0.1 M NaOH at a potential of +0.5 V vs. Ag/AgCl; Insert: enlarged amperometric response of the electrode at low concentrations; (b) current–glucose concentration calibration curve obtained from Figure 4(b).

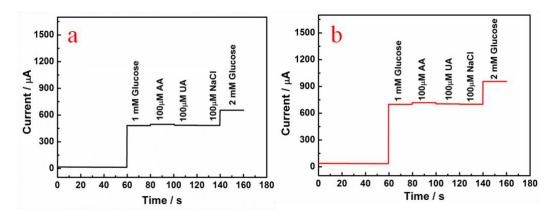
The amperometric method is a very sensitive method which can provide less signal-to-noise ratio and can result in rapid detection of an analyte. Therefore, in this study, the as-prepared Cu<sub>2</sub>O@Cu<sub>1.8</sub>S and Cu<sub>2</sub>O electrode was further used to investigate their amperometric responses at various glucose concentrations in a 0.1 M NaOH solution. Figure 5(a,c) shows the amperometric responses of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S and Cu<sub>2</sub>O electrode as an enzyme-free sensor for successive additions of glucose at different concentrations under the optimized detection potential of +0.50 V vs.

Ag/AgCl. The inserts in the figure show enlarged amperometric responses at low glucose concentrations. From the figures, it can be observed that the proposed sensor reached a steady-state current (95 % of the maximum) within 20 s (Fig. 7(a), inset), demonstrating a rapid amperometric response behaviour. As expected, the Cu<sub>2</sub>O@Cu<sub>1.8</sub>S displayed a stronger glucose response compared to the Cu<sub>2</sub>O modified glassy carbon disc electrode, which can be explained by its superior electrocatalytic activities for glucose oxidation and rapid charge-transfer behavior. Figure 5(b,d) show the electrochemical response calibration curve of Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S, which follows a linear response to glucose within the glucose concentration range of 0.1 to 800 mM ( $R^2 = 0.98$ ) for Cu<sub>2</sub>O and 0.1 to 1,000 mM ( $R^2 = 0.99$ ) for Cu<sub>2</sub>O@Cu<sub>1.8</sub>S. The upper-limit of the linear range is beyond the physiological level of 3–8 mM for practical use of glucose detection. From the slope of current vs. glucose concentration, a sensitivity of 2,444  $\mu$ A mM<sup>-1</sup> cm<sup>-2</sup> and a detection limit of 76.9 nM (S/N = 3) were found for the Cu<sub>2</sub>O electrode. In the case of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S electrode, a sensitivity of 3,630 µA mM<sup>-1</sup> cm<sup>-2</sup> was found, a much higher value than that of Cu<sub>2</sub>O, with a lower detection limit of 67.8 nM (S/N = 3). For comparison purposes, the performances of other copper oxide-based glucose sensors reported in literature are listed in Table 1. It can be seen that the sensor based on Cu<sub>2</sub>O@Cu<sub>1.8</sub>S exhibits better sensing performances in terms of sensitivity, detection potential and linear range than some of copper oxide/copper sulfide-based sensors [18, 35, 36], implying that the Cu<sub>2</sub>O@Cu<sub>1.8</sub>S material can be a promising and effective electrode for non-enzymatic glucose sensing. It is worth noting that the reported CuO film [34] and CuS modified Cu

electrodes [37] also showed an improved linearity and sensitivity to glucose compared to our Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S modified electrodes, which could be possibly attributed to an improved electronic conductivity. This finding suggests that there is still some room for further improvement in performances for Cu<sub>2</sub>O@Cu<sub>1.8</sub>S by using highly conductive support materials.

Electrode	Sensitivity/ μA cm <sup>-2</sup>	Linear	DoL/	Reference
	mM <sup>-1</sup>	Range/ mM	μM	
$Cu_2O@Cu_{1.8}S$	3630	0.001-1	0.0678	herein
Cu <sub>2</sub> O	2444	1-0.8	0.0769	herein
Cu/Cu <sub>2</sub> O	1434	0-40	1.6	[35]
Cu <sub>2</sub> O spindle	2828	0.001-1	0.3	[18]
Dandelion-like CuO films	s 5368	0.005-1.6	1.2	[34]
CuO nanoparticles	2762.5	0.05-18.45	0.5	[36]
$Cu_2S@Cu$	11750.8	0.0002-0.63	0.07	[37]
CuS nanotubes	7.842	0.05-5	10	[38]
Cu <sub>2</sub> SNP(EN)/GC	61.67	0.01-3.1	1.3	[39]

**Table 1.** Comparison of the key performance characteristics of some existing CuO-based electrodes for non-enzymatic electrooxidation of glucose.



**Figure 6.** Amperometric responses of the Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S electrodes with continuous injections of 1.0 mM glucose and 0.1 mM interferents of AA, UA, and NaCl into 0.1 M NaOH at a potential of +0.50 V vs. Ag/AgCl.

It is known that some easily oxidative species, such as ascorbic acid (AA), uric acid (UA), NaCl and other carbohydrate compounds, usually co-exist with glucose in biological samples and consequently may interfere with the detection of glucose. Therefore, selectivity is an important factor for the practical use of glucose sensors.

According to previous works, a normal physiological level of glucose is 3 – 8 mM, which is at least 10 times higher than that of the interfering species. The amperometric responses of the two electrodes by stepwise addition of 1.0 mM glucose in the presence of 0.1 mM of various interfering species at an applied potential of +0.50 V vs. Ag/AgCl is shown in Figure 6. From the figure, an obvious response of glucose oxidation can be observed, but there was no obvious current response from the interfering species, indicating that, in our conditions, a high selectivity against most of the important interfering species can be achieved with the Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S electrodes in an alkaline media. The excellent anti-interference

property of Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S electrodes for glucose against interfering species could be mainly ascribed to the intrinsic property of Cu<sub>2</sub>O, as well as its unique nanostructure. Under an alkaline environment (0.1 M NaOH), the surface of Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S may be negatively charged based on the value of isoelectric point for Cu<sub>2</sub>O (9.5); Moreover, interfering species like UA and AA may also be negatively charged due to the deprotonated effect in alkaline solution [18, 34, 36, 40, 41]. Consequently, the negatively charged UA and AA could be repelled on the negatively charged surface of Cu<sub>2</sub>O and Cu<sub>2</sub>O@Cu<sub>1.8</sub>S, thus resulting in corresponding weak signals.

### 4. Conclusions

The authors demonstrated for the first time the use of Cu<sub>2</sub>O@Cu<sub>1.8</sub>S as a posssible electrode material toward non-enzymatic glucose detection. Cu<sub>2</sub>O@Cu<sub>1.8</sub>S hybrid materials were successfully prepared by an in-situ growth method, in which out-diffusion of spherical Cu<sub>2</sub>O templates led to a hollow structure. Electrochemical measurements showed that Cu<sub>2</sub>O@Cu<sub>1.8</sub>S exhibited a better electrocatalytic activity toward glucose oxidation than Cu<sub>2</sub>O, which was attributed to the synergetic effects between Cu<sub>2</sub>O and Cu<sub>1.8</sub>S. The as-prepared Cu<sub>2</sub>O@Cu<sub>1.8</sub>S sensor exhibited a rapid response of 5 s, a wide linear range in glucose concentrations of 0.001–1 mM, a high selectivity of 3630 μA mM<sup>-1</sup> cm<sup>-2</sup>, and a low detection limit of 67.8 nM. Moreover, the sensor exhibited relatively excellent selectivity in 'real' sample analysis. The excellent sensing performances make Cu<sub>2</sub>O@Cu<sub>1.8</sub>S a promising candidate in the

development of non-enzymatic glucose sensors.

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