

Swelling and Free-Volume Characteristics of TEMPO-Oxidized Cellulose Nanofibril Films

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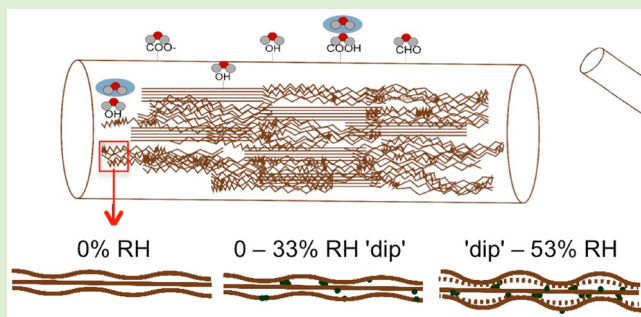
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ABSTRACT: Cellulose nanofibrils (CNFs) are becoming increasingly ubiquitous in diverse technologies requiring sustainable nanoscale species to form or modify films. The objective of the present study is to investigate the swelling behavior and accompanying free volume of self-standing TEMPO-oxidized (TO) CNF films in the presence of water vapor. For this purpose, we have performed time-resolved swelling experiments on films, prepared according to different experimental protocols, at 90% relative humidity (RH) and ambient temperature. Corresponding free-volume characteristics are elucidated by positron annihilation lifetime spectroscopy (PALS) conducted at ambient temperature and several RH levels. Increasing the drying temperature of the films (from ambient to 50 °C) is observed to promote an increase in film density, which serves to reduce bulk swelling. These elevated drying temperatures likewise cause the free-volume pore size measured by PALS to decrease, while the corresponding total free-volume fraction remains nearly constant. Similarly, dispersion of TO-CNF into aqueous suspensions by ultrasonication prior to film formation increases both the total free-volume fraction and pore size but reduces the size of individual nanofibrils with little net change in bulk swelling. The swelling and concurrent free-volume measurements reported here generally reveal an increase in the free volume of TO-CNF films with increasing RH.



INTRODUCTION

Cellulose nanofibrils (CNFs) surface-modified in the presence of 2,2,6,6-tetramethyl-piperidin-1-yl)oxyl, referred to as TEMPO (TO), have become increasingly popular since their inception in 2006.¹ Since that time, TO-CNF has been considered for use in a wide variety of technologies as, for instance, a functional paper additive,² an air-filter precursor,³ a biomedical device component,⁴ and a polymer composite additive to improve mechanical properties.⁵ Production of TO-CNF from sustainable wood resources is commonly achieved in a two-step process: TEMPO-mediated oxidation, followed by mechanical agitation to isolate the CNFs.⁶ While the starting material is routinely cellulose pulp originating from a broad range of sources, the final material consists of nano/microfibrils possessing a net negative surface charge due to the presence of carboxyl groups, which promote hydrophilicity. TEMPO treatment selectively oxidizes these hydroxyl groups to carboxylic moieties, resulting in a stronger negative surface charge and enhanced hydrophilicity. Depending on the oxidation conditions, the carboxylic acid content can be as high as ~1.6 mmol/g.⁷ Mechanical agitation introduces

disruptive shear forces through the use of homogenization, sonication, or microfluidization and generates CNFs typically possessing the same crystallinity as the starting pulp⁷ and measuring 1.6–3.8 nm in width^{8,9} and ranging from several hundred nanometers¹⁰ up to several micrometers in length.¹¹ Due to their high aspect ratio and to facilitate analysis, TO-CNFs are often schematically portrayed as rods (cf. Figure 1). A molecular-level study^{12,13} of CNF dissolution reveals that nanofibrils possessing a square cross section measuring 5.2 nm across contain 36 chains, thereby yielding ~1.3 chains/nm². Atomic force microscopy (AFM) images of the present TO-CNF (an example of which is included in Figure 1) indicate that the nanofibrils measure 2.37 ± 0.65 nm in diameter (measured in triplicate on 15 TO-CNFs) or, alternatively, 4.4 nm² on average in cross-sectional area. The corresponding number of chains/nanofibrils is therefore estimated to be ~6.

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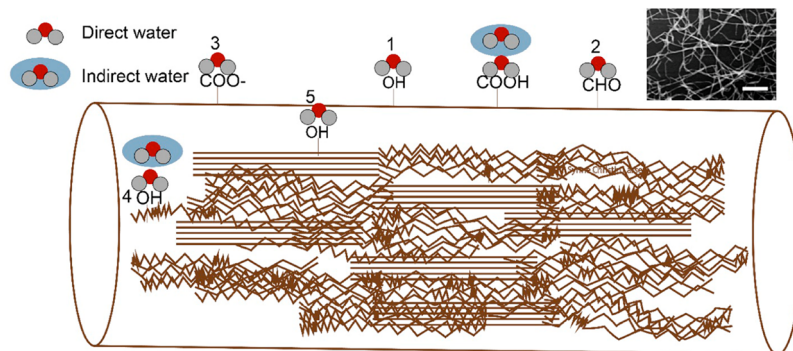


Figure 1. Schematic illustration of water sorption in a TEMPO-oxidized cellulose nanofibril (TO-CNF) film. Sorption occurs as water binds primarily to external hydroxyl, aldehyde, and carboxylic groups (1–3) in internal amorphous regions (4) and along crystalline facets (5). A representative AFM image of the TO-CNF is provided in the inset (scale bar = 200 nm).

66 We recognize that hierarchical structures, including (partial)
67 plant cells, might also be present.^{14,15}

68 Films composed of TO-CNF are relatively straightforward to
69 prepare but nontrivial to analyze. Of particular interest in the
70 present work is the swelling behavior of such hygroscopic films
71 in the presence of water vapor, since they are likely to be used
72 in environments varying in relative humidity (RH). As
73 described in more detail below, bulk swelling under these
74 conditions has been extensively studied. In this work, however,
75 we focus on the accompanying changes in TO-CNF free
76 volume. Free volume generally refers to the unoccupied space
77 between polymer chains¹⁶ and, when modeled as spherical
78 pores, is expected to measure on the order of a few angstroms
79 from equation-of-state considerations of small molecules.¹⁷
80 This concept, introduced to explain the glassy behavior of
81 amorphous polymers, is rooted in the entropic contribution¹⁸
82 to statistical thermodynamics.¹⁹ Free volume is associated with
83 the amorphous regions of a polymer,²⁰ in which case an
84 increase in polymer crystallinity proportionately decreases the
85 available free volume, which governs bulk properties such as the
86 glass transition temperature (T_g) and melt viscosity, as well as
87 the diffusive transport of molecular penetrants.¹⁷ While free
88 volume can be indirectly measured from PVT (pressure–
89 volume–temperature) or viscosity data, it can also be accurately
90 estimated from various group contribution methods. Since the
91 1990s, however, positron annihilation lifetime spectroscopy
92 (PALS) has gained increasing popularity as an analytical tool
93 that is capable of directly measuring free volume in polymer
94 films and composites.^{17,21} General observations to date indicate
95 that the free-volume pores (possessing a presumed spherical
96 shape) present in many polymers measure 0.4–1.0 nm in
97 diameter.^{22–25}

98 Of particular interest in the present study is the use of PALS
99 to explore the free volume of TO-CNF films.²⁴ In this case, the
100 diameter of spherical free-volume pores has been reported to be
101 0.47 nm, irrespective of position across film thickness under
102 vacuum. The use of evacuated conditions, however, altogether
103 precludes exposure to water vapor, which has been investigated
104 by PALS in conjunction with a variety of hydro/amphiphilic
105 polymeric materials, including poly(vinyl alcohol),²⁶ ep-
106 oxies,^{27,28} cellulose acetate,²⁵ and copolymers.²⁵ At low RH
107 levels, some of these systems are observed to experience a
108 decrease in average free-volume size that is lower than that
109 measured at 0% RH. This initial reduction has been attributed
110 to a molecular-level mechanism wherein water molecules fill
111 existing free-volume pores before the pores are forced to

112 expand at higher RH levels. In this study, TO-CNF films
113 produced by different fabrication methods are characterized to
114 reveal differences in film structure, macroscopic swelling, and
115 free volume due to the presence of water vapor. Here, we have
116 developed an experimental protocol that permitted direct
117 measurement of free volume at systematically varied RH levels,
118 which has not been previously reported to the best of our
119 knowledge with regard to TO-CNF films. The molecular-level
120 role of water discerned from resultant PALS data under these
121 conditions is interpreted to explain the mechanism of TO-CNF
122 film swelling.

Water Sorption Kinetics. Cellulose, as well as TO-CNF, is
123 a hygroscopic material, indicating that it naturally remains in
124 thermodynamic equilibrium with surrounding water vapor or
125 liquid.²⁹ This equilibrium is complex and thoroughly
126 investigated for different types of cellulose,³⁰ as well as for
127 cellulose in composites.³¹ Moreover, cellulosic materials are
128 prone to water-sorption hysteresis,³² which relates the
129 measured moisture content to the method by which the
130 material is conditioned in a moist or liquid environment prior
131 to measurement. When water uptake of hygroscopic cellulosic
132 materials is under consideration, it necessarily includes both
133 absorption and adsorption. Nanocellulose, of which TO-CNF
134 constitutes an example, does not possess a clearly defined
135 exterior surface, implying that both sorption mechanisms occur
136 simultaneously. In addition, sorption can be classified as either
137 slow or fast,³³ depending on the chemical makeup and spatial
138 location of the binding species. For instance, sorption occurs
139 quickly in TO-CNF films as water binds to surface hydroxyl
140 and carboxylic groups. Conversely, water molecules bind more
141 slowly to internal amorphous moieties and crystalline facets
142 (water cannot penetrate the crystallites³⁴). A distinction is also
143 made between direct sorption, which corresponds to water
144 molecules that hydrogen-bond directly to CNFs, and indirect
145 sorption, wherein water molecules bind to other, already bound
146 water molecules.
147

148 Other considerations that must be considered in water
149 sorption and TO-CNF film swelling⁶ are interfibril interactions,
150 which can inhibit swelling under the following conditions: (i)
151 the volume increase of a hydrated nanofibril is physically
152 hindered by other nanofibrils in the network, and (ii) water
153 binding competes with interfibril binding for available hydrogen
154 bonds. Since water sorption in TO-CNF films follows a
155 bimechanistic process (e.g., fast vs slow, direct vs indirect, and
156 water binding vs interfibril binding), a model that is commonly
157 used to describe the water sorption kinetics for cellulosic films

158 is the parallel exponential kinetics (PEK) model.³⁵ This model
 159 derives from the presumption that the mass gain at a given time
 160 (t) during swelling at a specific RH (M_t) is the sum of two
 161 parallel swelling processes, each with its own time constant (α_1
 162 or α_2), as indicated by

$$163 \quad M_t = M_{\infty 1}(1 - e^{-t/\alpha_1}) + M_{\infty 2}(1 - e^{-t/\alpha_2}) \quad (1)$$

164 Equation 1 can be interpreted as the occurrence of two
 165 concurrent sorption processes with each capable of reaching its
 166 equilibrium water solubility ($M_{\infty 1}$ and $M_{\infty 2}$). The processes in
 167 the PEK model possess nominally fast (α_1) and slow (α_2)
 168 characteristic times, and the corresponding sorption sites are
 169 schematically depicted in Figure 1. While precise categorization
 170 of different sorption sites as fast or slow has yet to be
 171 conclusively established³⁶ in terms of their relation to specific
 172 terms in eq 1,^{30,33,35} the time constants are material-specific and
 173 can be extracted from sorption experiments performed at
 174 different RH levels.³⁰

175 **Positron Annihilation Lifetime Spectroscopy.** Since
 176 PALS remains a largely specialized characterization technique,
 177 we briefly describe its underlying principle and analysis
 178 methodology in this section. The technique uses positrons
 179 (electron antiparticles) as a probe by which to characterize
 180 vacancy defects and free-volume voids in a wide range of
 181 inorganic and organic materials. Detailed reviews of PALS are
 182 available elsewhere.^{12,32} When positrons are implanted into a
 183 subject material, they tend to diffuse and become trapped in
 184 such vacancies and voids, and eventually annihilate with
 185 surrounding electrons. In a typical bulk system subjected to
 186 PALS analysis that employs ²²Na as the positron source, the
 187 birth of a positron is marked as the start signal by detection of
 188 coincident γ radiation of 1274 keV, whereas the decay of the
 189 positron is signaled by detection of annihilation γ rays of 511
 190 keV, which effectively serves as the stop signal. By recording the
 191 temporal intensity (I) histogram between the start and stop
 192 signals, a PALS spectrum is obtained. Such spectra frequently
 193 consist of several exponential decay components of the form

$$194 \quad I(t) = \sum_{j=1}^n I_j \exp\left(-\frac{t}{\tau_j}\right) \quad (2)$$

195 where I_j and τ_j denote the intensity (or number-density
 196 population) and lifetime of the j th annihilation lifetime
 197 component, respectively.

198 In polymers and semiconductors, positrons might also form
 199 positronium (Ps) species (a metastable bound state of a
 200 positron and an electron) that also undergo annihilation. The
 201 alignment of the electron and positron spins dictates whether
 202 Ps is in a singlet state (*para*-positronium or p-Ps) or a triplet
 203 state (*ortho*-positronium or o-Ps). Since the first two lifetimes
 204 (τ_1 and τ_2) in eq 2 correspond to positron and/or p-Ps
 205 annihilation and are only related to fast processes that are
 206 relatively short, their analysis is not relevant in the present
 207 study. Of particular importance in this study is the o-Ps lifetime,
 208 which can be correlated with the free-volume pore size of
 209 polymers, thereby revealing the morphological features of
 210 polymeric media at atomic length scales. Although o-Ps ions
 211 possess a lifetime of 142 ns in vacuum, this lifetime can be
 212 significantly shortened within materials through a pick-off
 213 process wherein the positron annihilates an environmental
 214 electron (from the polymer) instead of its original pair. This
 215 shortened o-Ps lifetime (τ_3) can be directly related to the free-

216 volume pore size through such correlations as the Tao–Eldrup
 217 model,^{37,38} expressed as

$$218 \quad \tau_3 = 0.5 \left(1 - \frac{r}{r + \Delta r} + \frac{1}{2\pi} \sin\left(\frac{2\pi r}{r + \Delta r}\right) \right)^{-1}$$

219 where r represents the radius of a spherical free-volume pore
 220 and Δr (=0.166 nm) accounts for the electron shell extending
 221 into the free-volume pore.³⁹ While this model is commonly
 222 used to extract free-volume characteristics from polymers, other
 223 models, including those not restricted to specific shapes, have
 224 been proposed. In the present work, a shape-free model⁴⁰ that
 225 cannot be expressed in closed form due to its quantum-
 226 mechanical derivation has been utilized for analysis purposes.
 227 Moreover, the associated intensity of the o-Ps component (I_3)
 228 in eq 2 provides the relative number density of free-volume
 229 elements⁴¹ among only similar material systems, since this
 230 quantity is sensitive to material chemistry. When comparing
 231 PALS results acquired from the same material series, however,
 232 the total volume (L) of the free-volume elements, approxi-
 233 mated as spherical in shape, can be obtained from

$$234 \quad L = I_3 \times \frac{4}{3} \pi r^3 \quad (4)$$

235 ■ EXPERIMENTAL SECTION

236 **Materials.** Softwood (fully bleached) pulp was supplied by Södra
 237 (Växjö, Sweden) and was oxidized at ambient temperature with 98%
 238 TEMPO (Sigma-Aldrich, St. Louis, MO, USA), sodium bromide
 239 (NaBr; > 99%, Sigma-Aldrich), and sodium hypochlorite (NaClO;
 240 Roth Chemicals, Karlsruhe, Germany). Sodium hydroxide (NaOH)
 241 and hydrochloric acid (HCl) were purchased from Fisher Scientific
 242 (Hampton, NH, USA).

243 **Methods. 1. Pulp Oxidation.** A fresh solution consisting of 1.25%
 244 w/w TEMPO/pulp and 12.5% w/w NaBr/pulp was prepared in
 245 deionized (DI) water on the day of use. The solution was added to
 246 220 g of pulp stirred in DI water for 10 min, and the resulting
 247 suspension was diluted to 1.33 wt % prior to oxidation with 1.1 mol of
 248 NaClO gradually added in eight allotments over the course of 50 min.
 249 This corresponds to 5 mmol of NaClO/g of pulp. During this
 250 oxidation process, the pH was maintained at 10.5 by gradually adding
 251 0.5 M NaOH. When the pH remained constant over a period of 10
 252 min, the reaction was presumed to be completed, and the suspension
 253 was neutralized (pH 7.0) with 0.5 M HCl. The oxidized pulp was
 254 vacuum-filtered using filter paper with DI water until a conductivity of
 255 <5 μ S/cm was measured and subsequently stored at 4 °C. Suspensions
 256 at 0.8 wt % concentration were homogenized with a Rannie 15 type
 257 12.56X instrument (APV, SPX Flow Technology, Silkeborg, Den-
 258 mark) operated at 600 bar for one pass, followed by a second pass at
 259 1000 bar. The final carboxylic acid content was measured by
 260 conductometric titration⁴² as the average of three parallel trials. The
 261 acid-soluble lignin fraction was determined according to the TAPPI
 262 Useful Method 250 standard with an absorption of 205 nm and lignin
 263 extinction coefficient of 110 g/L cm. The remaining cellulose and
 264 hemicellulose contents were discerned by subjecting the suspension to
 265 acid hydrolysis, followed by high-pressure liquid chromatography
 266 (HPLC).

267 **2. Film Preparation.** A suspension composed of 400 g of 0.3 wt %
 268 TO-CNF in DI water was prepared by stirring at 1100 rpm for 60 min,
 269 followed by either direct casting or probe sonication prior to casting.
 270 Probe sonication, conducted with a 3.2 mm Qsonica probe for 10 min
 271 at 80% intensity, was performed while concurrently stirring at 1100
 272 rpm. Specimens are hereafter designated as S (for sonicated
 273 suspension) or NS (for not sonicated suspension). Films were
 274 produced by casting 60 mL suspensions into polyethylene Petri dishes
 275 (measuring 15 mm in depth and 95 mm in diameter) and then drying
 276 at ambient temperature (20–22 °C), 35, or 50 °C in a gravity

277 convection oven. Specimens are further denoted as Amb (dried at 20–
278 22 °C), 35 (dried at 35 °C), or 50 (dried at 50 °C), as listed in Table
279 1. All films were finally dried at 80 °C for 3 h at 0.33 bar.

Table 1. Specimen Designations Employed in This Study

specimen designation	drying temperature (°C)	sonication status
Amb-NS	ambient	NS
Amb-S	ambient	S
35-NS	35	NS
35-S	35	S
50-S	50	S

280 **3. Sample Characterization.** Dynamic light scattering (DLS) was
281 performed on TO-CNF suspensions diluted to 0.01 wt % and gently
282 stirred for 5 min with a Malvern Nano Zetasizer. The viscosity was
283 taken as that of water (0.89 cP), and CNF suspensions were measured
284 for 30 runs of 30 s each for a total of 30 such measurements. These
285 measurements were averaged to yield the reported particle size
286 distribution. Scanning electron microscopy (SEM) of specimens
287 cryofractured in liquid nitrogen to yield cross sections that were
288 subsequently sputter-coated with 5 nm Au/Pd was conducted on an
289 ultrahigh-resolution FEI Verios 460L instrument at 1.0–1.5 kV
290 without stage bias. Samples were prepared and investigated the same
291 day. Films intended to investigate swelling kinetics were stored in a
292 desiccator at 0% RH for at least 16 h until the swelling tests were
293 performed. These tests were conducted gravimetrically by exposing
294 films to water vapor at $90 \pm 2\%$ RH and 23.9 ± 0.1 °C in a Percival
295 climate chamber. Specimens were weighed every hour for the first 12 h
296 and then at preselected times thereafter. All swelling experiment
297 results reflect four replicates, and equilibrium values correspond to 96
298 h. Film swelling (S) was characterized by the film weights prior to
299 swelling (W_0) and after a given time in the humidity chamber (W_t)
300 according to

$$S = \left(\left(\frac{W_t}{W_0} \right) - 1 \right) \times 100\% \quad (5)$$

301
302 The PALS measurements were performed in the presence of an
303 ~ 10 μCi ^{22}Na source sealed between Kapton films measuring 7.6 μm
304 thick. Two Hamamatsu 3378-50 photomultiplier tubes with plastic
305 scintillators (25.4 mm diameter) were used to detect the start and stop
306 signals of the experiments. While a more detailed description of the
307 PALS system has been provided elsewhere,^{43,44} a photograph of the
308 setup along with a schematic illustration is presented in Figure 2.

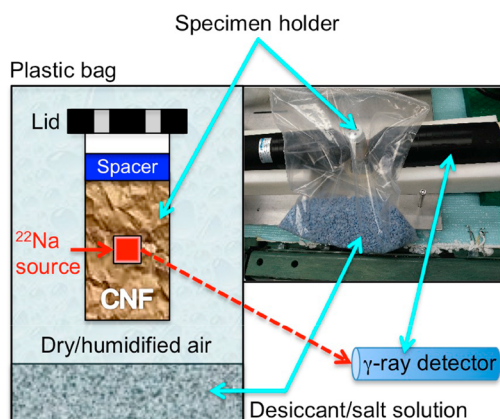


Figure 2. Schematic illustration of the PALS setup employed in this study. A sealed plastic bag contains a glass vial (into which is placed a TO-CNF specimen and the ^{22}Na positron source) and a desiccant or salt solution to maintain a specific RH level within an enclosed plastic bag.

Specimens to be examined were prepared by breaking about eight
films into pieces (totaling ~ 1.6 g) that were subsequently inserted into
a vial. The positron source was positioned among the sample pieces,
which were pressed to minimize spurious signals from air, and the vial
was capped. Multiple holes were drilled into the cap of each vial to
allow equilibration with the surrounding environment. Sample-filled
vials were placed in plastic bags containing either desiccants or ~ 150
mL of a suitable salt solution to generate the RH levels listed in Table
2. The plastic bags were then sealed, and the samples were allowed to

Table 2. Relative Humidity Levels Achieved in the PALS Analyses^a

water-vapor source	RH levels (%)
desiccant	0
saturated MgCl_2	33 ± 0.2
saturated $\text{Mg}(\text{NO}_3)_2$	53 ± 0.2
saturated NaCl	75 ± 0.1
deionized water	100

^aReported RH levels are extracted from ref 6, and the error corresponds to the standard deviation.

equilibrate. The desiccant remained blue at 0% RH, ensuring that the
sample environment was at 0% RH (otherwise the desiccant would
turn red). To monitor/confirm the equilibration process, PALS
spectra, recording at least 1.5×10^6 events, were acquired at ambient
temperature every 1–2 days. Resultant spectra were primarily fitted
with three annihilation lifetimes using the POSFIT software package,
and the regressed value of τ_3 was related to the size of free-volume
elements via a shape-free quantum-mechanical model.⁴⁰

RESULTS AND DISCUSSION

TO-CNF Film Characterization. Conductometric titration
reveals that the carboxylic acid content [$-\text{COO}- + \text{COOH}$] of
the TO-CNF generated here is 1343 ± 7.2 $\mu\text{mol/g}$. Acid
hydrolysis confirms that the cellulose content is ~ 79 wt %,
whereas the acid-soluble lignin is ~ 0.6 wt % (with no
detectable acid-insoluble lignin). Galacto-(glucomannan) and
xylan account for ~ 9 and ~ 7 wt %, respectively. Films
fabricated from the procedure described here measure ~ 20 μm
thick and appear highly transparent, as generally evidenced by
the photographs presented in Figure 3. While the films appear
visually similar when cast from sonicated or nonsonicated
suspensions, their appearance is noticeably affected by the
drying temperature. Films dried at ambient temperature for
about 23 days are virtually defect-free (Figure 3A). Films dried
at 35 °C over the course of 6 days display visible bubbles
(Figure 3C), and those dried at 50 °C for 1–2 days exhibit a
large population of bubbles (Figure 3E). This observation
clearly indicates that, as the specimens are subjected to higher
temperatures and expedited film drying, the resulting TO-CNF
films more readily entrap liquid water, which produces bubbles
upon vaporization. This observation is consistent with SEM
images of cross-fractured films, displayed in Figure 3. An
increase in the drying temperature and corresponding rate of
film formation is accompanied by enhanced film densification
in regions away from bubbles. The mechanical strength of the
films is likewise sensitive to bubble formation (and, hence,
drying time), since films dried quickly are fragile and tear easily.
The specimens displayed in Figure 3B, D, and F are all
sonicated to permit discrimination by SEM (this is not required
at the length scale associated with visual examination).
According to the DLS results provided in Figure 4, sonication

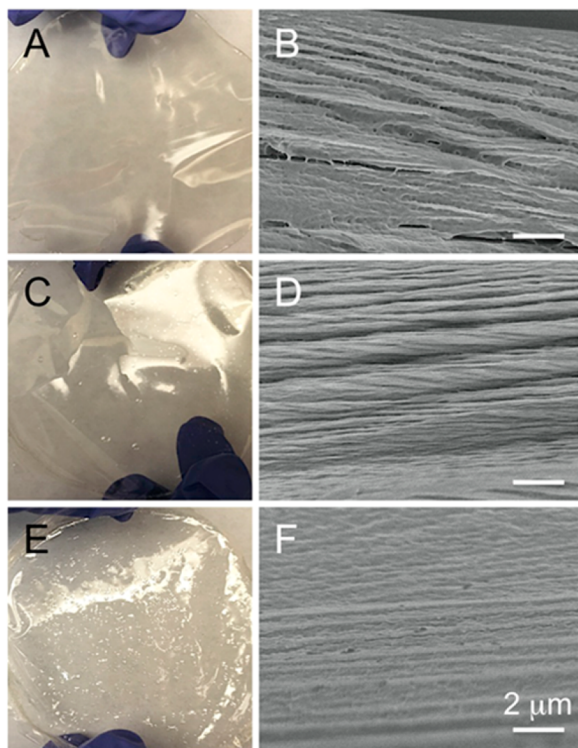


Figure 3. Photographs (left) and cross-sectional SEM images (right) of sonicated TO-CNF films dried at different temperatures: (A, B) ambient temperature, (C, D) 35 °C, and (E, F) 50 °C.

TO-CNF Film Swelling. Swelling tests performed in a Percival climate chamber on four replicates establish that S lies between ~ 24 – 33% after exposure to $90 \pm 2\%$ RH and 23.9 °C for 96 h. Figure 5A displays the time dependence of S for films

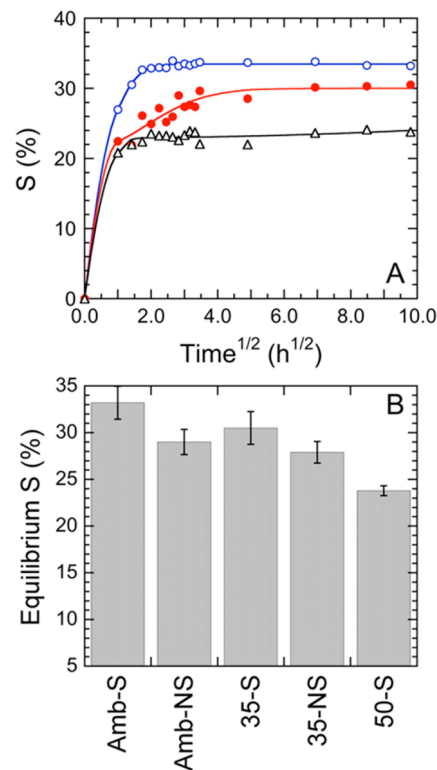


Figure 5. (A) Time-dependent swelling (S) curves determined from eq 5 at 90% RH for TO-CNF films prepared by sonication and dried at three different temperatures: ambient (○), 35 °C (●), and 50 °C (△). The solid lines are regressions of eq 1 to the data, and the error bars denote standard errors. (B) Equilibrium S values obtained from swelling curves at 90% RH for the five different specimen designations examined in this study (cf. Table 1).

serves to reduce the mean TO-CNF length and shift the size distribution to shorter nanofibrils.

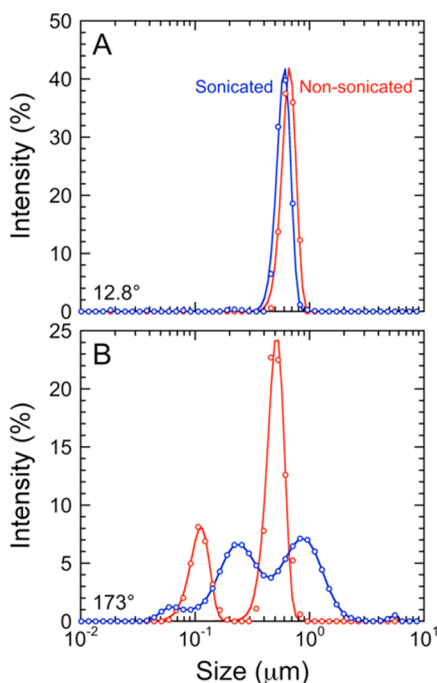


Figure 4. Nanofibril dimensions obtained from DLS at (A) 12.8° and (B) 173° on TO-CNF suspensions with (blue) and without (red) sonication. The solid lines represent Gaussian fits to the peaks in the data.

that have been sonicated as suspensions and dried at three different temperatures. Application of eq 1 to these data results in the following values of α_1 and α_2 , respectively (in h): 0.036 and 1.177 (ambient temperature), 0.008 and 6.553 (35 °C), and 0.430 and 9.341×10^4 (50 °C). After exposure to water vapor for 1 h, the TO-CNF films dried at ambient temperature swell by 27%, whereas those dried at 35 and 50 °C swell to 22 and 21%, respectively. After 12/24 h, these S levels increase to 34/34% (ambient), 30/29% (35 °C), and 22/22% (50 °C). Corresponding values of S at 96 h are 33, 31, and 24%, respectively, revealing that swelling is negligible after 12 h. In fact, these results indicate that at least $\sim 70\%$ of the equilibrium swelling in these TO-CNF films occurs within the first hour of exposure. In addition, the swelling results displayed in Figure 5A likewise demonstrate that an increase in drying temperature promotes a reduction in equilibrium swelling. This specimen preparation issue, as well as suspension sonication, is explicitly addressed in Figure 5B, which shows that, while sonication tends to improve swelling modestly (by 14% at ambient temperature and 9% at 35 °C), increased drying time lowers swelling to a greater extent (by 28% from ambient temperature to 50 °C). This latter observation, which is consistent with the SEM images provided in Figure 3, implies that densified TO-

Table 3. PALS Lifetimes Measured at 0% RH for TO-CNF Films Prepared from Sonicated and Nonsonicated Suspensions

specimen	τ_1 (ns)	I_1 (%)	τ_2 (ns)	I_2 (%)	τ_3 (ns)	I_3 (%)	τ_4 (ns)	I_4 (%)
Three-Lifetime Fitting								
Amb-NS	0.11 ± 0.05	11.81 ± 5.54	0.38 ± 0.01	73.28 ± 5.34	1.27 ± 0.01	14.91 ± 0.32		
Amb-S	0.19 ± 0.02	17.83 ± 2.14	0.40 ± 0.01	68.19 ± 1.99	1.36 ± 0.01	13.98 ± 0.21		
Four-Lifetime Fitting								
Amb-NS	0.08 ± 0.01	14.28 ± 1.03	0.37 ± 0.01	70.43 ± 0.84	1.24 ± 0.02	15.18 ± 0.32	7.47 ± 5.76	0.11 ± 0.04
Amb-S	0.09 ± 0.01	16.01 ± 1.81	0.37 ± 0.01	67.88 ± 1.62	1.19 ± 0.04	15.11 ± 0.34	2.55 ± 0.46	1.01 ± 0.58

387 CNF films dried at elevated temperatures are more likely to
 388 hinder the sorption of water molecules and thus swell. Another
 389 important consideration here is that hornification might be at
 390 least partially responsible for this apparent result. Hornification,
 391 an irreversible stiffening of cellulose chains that naturally
 392 occurs upon drying as glucose units undergo chemical cross-
 393 linking,⁴⁵ would also serve to inhibit swelling. This process has
 394 been reported to increase with increasing carboxylate content⁴⁶
 395 and drying temperature.⁴⁷

396 **Free Volume: Dry TO-CNF Films.** In this section, we
 397 explore the effects of sonication and drying temperature on the
 398 free volume of dry TO-CNF films. To begin, PALS spectra
 399 collected from films at 0% RH prepared from both sonicated
 400 and nonsonicated suspensions dried at ambient temperature
 401 have been well-fitted with three (and, for comparison, four)
 402 characteristic lifetimes in eq 2. Multiple spectra have been
 403 acquired to confirm that the specimens have reached their
 404 equilibrium state, and results from the last series are listed as
 405 typical values in Table 3. The regressed value of the o-Ps
 406 lifetime (τ_3) ascertained from the sonicated specimen after
 407 regression with three characteristic lifetimes (1.36 ns) is
 408 noticeably larger than that of the nonsonicated one (1.27 ns),
 409 which suggests that the sonication process affects the radius (r)
 410 of the free-volume pores. Calculated values of r — 0.206 ± 0.002
 411 and 0.218 ± 0.001 nm for the Amb-NS and Amb-S specimens,
 412 respectively—are lower than results previously reported²⁴ for
 413 dry TO-CNF films ($r = 0.235$ nm). Corresponding values of L
 414 from eq 4 are $(5.26 \pm 0.08) \times 10^{-3}$ nm³ (Amb-NS) and $(5.78$
 415 $\pm 0.06) \times 10^{-3}$ nm³ (Amb-S), indicating that the sonicated
 416 specimen possesses more free volume (with modestly larger
 417 free-volume pores). This result might help to explain the
 418 sonication-induced increase in equilibrium swelling evident in
 419 Figure 5B. To discern the extent to which the values of τ_3 and L
 420 are robust, the PALS spectra have also been fitted to four
 421 characteristic lifetimes (cf. Table 3). While τ_4 and I_4 are also
 422 metrics of free volume in polymeric media and suggest the
 423 presence of larger free-volume pores, the values of I_4 are
 424 substantially smaller than the other intensity values ($\sim 1\%$ for
 425 the sonicated specimen and $\sim 0.1\%$ for the nonsonicated one),
 426 in which case these contributions to the free-volume analysis
 427 are probably negligible and are not considered further in this
 428 work. Another indicator that these additional lifetimes are most
 429 likely inconsequential is the surprisingly large uncertainty in τ_4
 430 relative to all the other extracted τ_j values in Table 3 (<0.05 ns).

431 In addition to TO-CNF suspension sonication, another
 432 consideration of importance in this section is the drying
 433 temperature. Three-lifetime analysis of relevant PALS spectra as
 434 above for only the sonicated specimens yields free-volume pore
 435 radii of 0.215 ± 0.001 and 0.212 ± 0.001 nm at drying
 436 temperatures of 35 and 50 °C, respectively. Values of r and L
 437 extracted from these data are displayed as a function of drying
 438 temperature (where ambient is taken as 21 °C) in Figure 6 and
 439 reveal that, while r systematically decreases, L decreases to a

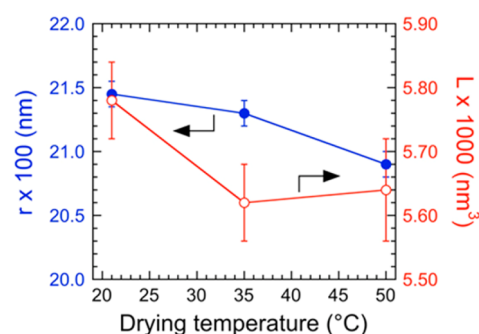


Figure 6. Free-volume pore radius (r) and total free volume (L) calculated from PALS spectra collected from TO-CNF films that were sonicated as suspensions and dried at different temperatures (color-coded). The solid lines serve to connect the data, and the error bars denote standard errors.

plateau, with increasing drying temperature. That is, increasing
 440 the drying temperature from 35 to 50 °C has no discernible
 441 effect on L . These changes in free-volume characteristics with
 442 drying temperature can be explained in terms of the hierarchical
 443 structure of TO-CNF.⁶ We have established that an increase in
 444 drying temperature promotes film densification, as independ-
 445 ently determined from SEM (Figure 3) and equilibrium
 446 swelling (Figure 5). On the basis of these experimental
 447 observations, such drying-induced densification is presumed to
 448 influence nanofibrillar packing. If this is the case, then it follows
 449 that either interfibrillar pores or possibly the nanofibrils
 450 themselves are most likely compressed.
 451

452 Since the crystalline regions of the chains are expected to
 453 remain largely unaffected, intrachain, as well as interchain,
 454 amorphous regions responsible for the free volume within CNF
 455 densify at the nanoscale, resulting in smaller free-volume pores.
 456 Faster drying times at elevated temperatures most likely
 457 prevents the chains from adopting their lowest energy
 458 conformations, and thus, the population of free-volume pores
 459 increases (apparently to a limit) at higher drying temperatures.
 460 Corresponding values of I_3 under these conditions are
 461 calculated to be $(14.0 \pm 0.2)\%$ for Amb-S and $(14.8 \pm 0.4)\%$
 462 for 50-S.

463 We anticipate that not all of the free-volume pores in this
 464 scenario are accessible to penetrant water molecules, in which
 465 case equilibrium swelling decreases. In marked contrast,
 466 sonication loosens the nanofibrils⁴⁸ and disrupts their
 467 crystalline regions⁴⁹ prior to drying. Doing so simultaneously
 468 increases both r and L , as reported above.

469 **Free Volume: Swollen TO-CNF Films.** Thus far, we have
 470 explored the effects of suspension sonication and drying
 471 temperature on the free-volume and swelling characteristics of
 472 TO-CNF films at 0% RH. In this section, the response of TO-
 473 CNF films exposed to sonication and dried at ambient
 474 temperature (designated as Amb-S in Table 1) is monitored
 475 at several different RH levels. The tests performed here utilize

476 the setup described with regard to Figure 2. Compared to the
 477 static swelling performed in the climate chamber (cf. Figure 5),
 478 the films investigated by PALS require significantly longer
 479 equilibration times (several days vs ~ 12 h) due to the
 480 experimental configuration. Representative peak-normalized
 481 PALS spectra obtained from Amb-S specimens at different
 482 RH levels and ambient temperature after an exposure time of
 483 5–11 days (spectra shown correspond to equilibrium
 484 conditions at each RH level) are presented in Figure 7 and

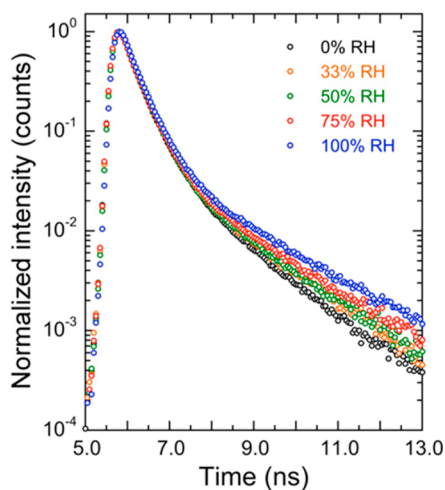


Figure 7. Peak-normalized PALS spectra collected from TO-CNF films prepared from sonicated aqueous TO-CNF suspensions dried at ambient temperature for 23 days and measured at different RH levels (see legend) at ambient temperature.

485 qualitatively indicate that the o-Ps lifetimes are systematically
 486 shifted to higher values as the RH level is increased.
 487 Corresponding values of r and I_3 extracted by analyzing such
 488 spectra according to eq 2 and the model provided elsewhere⁴⁰
 489 with three characteristic lifetimes are provided in Figure 8 and

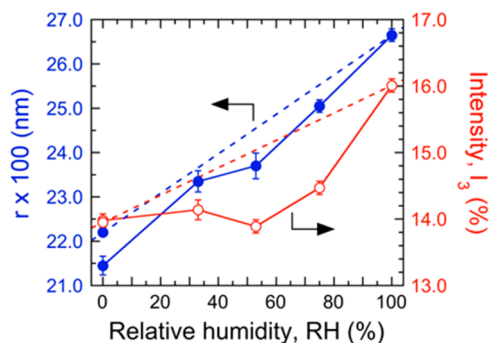


Figure 8. Long-time values of r (blue) and I_3 (red) as functions of relative humidity (RH) at ambient temperature. The solid lines serve to connect the data in the increasing RH direction, the dashed lines connect the data upon redrying, and the error bars denote standard errors.

490 reveal that r (which exhibits the same RH dependence as τ_3 , not
 491 included for that reason) increases monotonically with
 492 increasing RH level from 0.215 ± 0.001 nm at 0% to 0.267
 493 ± 0.001 nm at 100%, an increase of about 24%. The
 494 dependence of I_3 on RH differs slightly: it remains relatively
 495 constant up to 53% RH and then increases by about 15%. To
 496 discern the extent of hysteresis upon deswelling, the specimen

at 100% RH was redried to 0% RH and remeasured. As seen in
 Figure 8, the initial free-volume intensity is recovered, but the
 free-volume pore size is not. The original pore radius measures
 0.215 nm, while the radius after redrying is 0.222 nm. At
 macroscopic length scales, pulp drying results in lower
 porosity.^{45,50} Free volume that exists at nanoscopic length
 scales is not, however, affected by surface tension in the same
 way as macropores when water evaporates.

To determine the time dependence of RH-induced free-
 volume changes, we have performed PALS at different times at
 each RH level to follow the evolution of free-volume pore size
 and o-Ps intensity during the course of the experiment. The
 results are shown for both r and I_3 in Figure 9 and generally

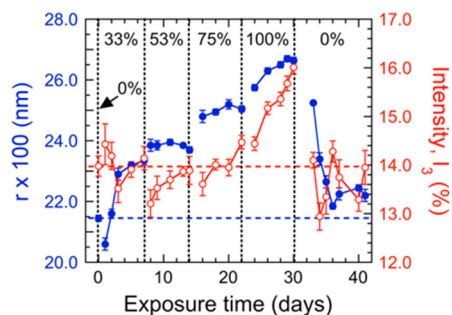


Figure 9. Free-volume pore radius and intensity of the Amb-S specimen in the presence of different RH levels (labeled) as a function of exposure time. Dotted lines represent the times at which the long-time value is recorded for a given RH level (these values are displayed in Figure 8).

demonstrate that, except during the initial swelling and final
 redrying, r is not as sensitive to exposure time at each RH as
 I_3 . An interesting observation on the first day at 33% RH provides
 evidence of atypical pore filling, which is also identified in other
 polymer systems.⁵¹ This phenomenon is manifested as a slight,
 yet statistically significant, initial decrease in average pore radius
 from 0.215 to 0.206 nm (a reduction of $\sim 4\%$) before r
 increases or levels off at longer times. Values of r and I_3
 recorded at the longest time examined at each RH level are
 presumed to be representative of near-equilibrium and are used
 to construct Figure 8. While r behaves nearly as anticipated
 (even during the transient periods of initial swelling and final
 redrying), the free-volume intensity does not during the
 transient periods. Upon initial swelling, for instance, I_3
 decreases over the course of the first 3 days as r systematically
 increases. As the specimen is redried, r expectedly decreases
 to a relatively constant value. While we do not have a conclusive
 explanation for these results, we posit that the presence of
 water molecules might be a contributing factor. Free water
 possesses an o-Ps lifetime of 1.86 ns,⁵² which might impact the
 measured free-volume characteristics here if water in the TO-
 CNF film elicits its own o-Ps signal. Free water molecules can
 potentially contribute to the extracted values of r , since its own
 “free-volume” (Ps bubble) radius is 0.27 nm (cf. Table 4),⁵²
 which is comparable to the pore size ascertained here and in
 cellulose acetate.²⁵ Below, we address the issue of how water
 contributes to PALS spectra at different RH levels.

The Kelvin equation describes the thermodynamics govern-
 ing pore filling and assumes that pores < 10 nm are not filled
 at 90% RH under ambient conditions. While this relationship can
 elucidate the size distribution of pores through the use of
 adsorption porosimetry, it is only valid for relatively large pores

Table 4. o-Ps Characteristics and Pore Radii for Dry TO-CNF Film Dried from Sonicated Suspension and Pure Water

system	τ_3 (ns)	I_3 (%)	r (nm)
TO-CNF film	1.36 ± 0.01	13.98 ± 0.21	0.215 (this work)
pure water ⁵²	1.86	26.9 ± 0.5	0.27

and does not account for site-specific interactions such as hydrogen bonding in its surface tension term. It nonetheless provides an experimental tactic, as detailed in the following section. Our results reported here, as well as others,^{25,51} indicate that water initially fills free-volume pores measuring $\ll 10$ nm. The PALS results provided in Figure 8 establish that the o-Ps intensity is relatively constant, whereas r systematically increases, up to $\sim 53\%$ RH, implying that the free-volume pores expand in this interval but that their number density does not change appreciably. Since evidence of pore filling is observed after just 1 d at 33% RH in Figure 9, water molecules most likely adsorb to the surface of the free-volume pores and partially fill them. Pore filling is subsequently overtaken by concomitant pore swelling, which serves to push neighboring TO-CNF chains away from each other. Since these water molecules are bound by hydrogen bonds, they should not be treated as free water with the free-volume characteristics listed in Table 4. At RH levels beyond 53%, both free-volume pore size and intensity increase with increasing exposure time in Figure 8. This observation is explained by water opening the amorphous regions of TO-CNF, thus creating more and larger free-volume pores. Under these conditions, however, the existence of free water can generate a PALS signal with the o-Ps intensity and pore size included in Table 4. As seen in Figure 3B, the TO-CNF films exhibit interfibrillar pores that measure in the micrometer range. While these are not to be confused with the free-volume pores, they could be filled with free water at high RH levels, thereby eliciting a pure water signal in PALS. This possibility is supported by oxygen permeability data,²⁴ which increases as TO-CNF films are exposed to water vapor, signifying that water penetrates into interfibrillar spaces and thus reduces film density.

To further determine the role of water in the PALS analysis, the o-Ps intensity data in Figure 8 have been correlated with RH level by an empirical exponential function to interpolate the o-Ps intensity at 90% RH. With a regression coefficient (R^2) of 0.98, this correlation yields an intensity of 15.1%, which can be used to ascertain if I_3 corresponds to the mass-weighted average of o-Ps intensities from free water and cellulose. The swelling data included in Figure 5A indicate that the mean equilibrium swelling for Amb-S at 90% RH is $33.2 \pm 1.8\%$, which is equivalent to 24.9 ± 1.0 wt % water relative to the total swollen mass. To test the hypothesis above, the total signal intensity is expressed in terms of the pure water intensity (I_{3w}), the contributing water fraction (f_w), the TO-CNF intensity (I_{3c}), and the contributing cellulose fraction (f_c) in a linear fashion to yield

$$I_3 = f_w I_{3w} + f_c I_{3c} \quad (6)$$

By utilizing the values listed in Table 5, it becomes clear that not all of the free water contributes to the measured o-Ps signal, since this leads to a higher value of I_3 than interpolated from measurements ($\sim 17\% > \sim 15\%$). Solving eq 6 for f_w with the requirement that $f_w + f_c = 1$ and $I_3 = 0.151$ yields 8.7 wt % water, thereby verifying that a nontrivial fraction of water in the system is bound to the TO-CNF chains and incapable of contributing an o-Ps signal to PALS spectra. A similar comparison performed by applying a simple linear rule of mixtures to the o-Ps lifetimes for pure water and cellulose results in 7.9 wt % water, which is in fair agreement with the water content from the o-Ps intensity calculations.

The free water fractions calculated above from the o-Ps intensities of the pure and swollen TO-CNF systems reveals that 35% of the water in the Amb-S specimen is free (and contributes to the measured value of I_3) and the remaining 65% is bound to the cellulose and is responsible for the measured free-volume characteristics of the nanofibrils. A related issue to consider is the possibility that bound water alters the formation of o-Ps in such a fashion that a reduction in o-Ps intensity due to the presence of water-containing free-volume pores is compensated by the intensity contribution from free water. For this effect to be realized, the correct o-Ps intensities and characteristic lifetimes would have to be concurrently maintained over the entire RH range, which is not very likely. On the basis of the experimental results reported here, water initially fills existing free-volume pores (as evidenced by a decrease in r after 1 day in Figure 9) but is strongly bound to the cellulose (without contributing to the o-Ps intensity) in general agreement with the classification of “nonfreezing” water during initial cellulose swelling.^{53,54} After the initial pore size reduction, water at longer exposure times and higher RH levels expands the existing free volume pores. In this regime, the o-Ps intensity remains relatively steady, suggesting that most of the water remains tightly bound to the cellulose without contributing to the o-Ps intensity and consists of both “nonfreezing” and “freezing bound” water.⁵⁴ The latter consists of water molecules that are tightly bound to cellulose and exhibit a significantly depressed freezing point compared to free water. At RH levels above $\sim 53\%$ RH, however, the presence of free water becomes non-negligible. Since the concurrent increases in o-Ps characteristic lifetime (which follows the RH dependence of r) and intensity in Figure 9 are much more pronounced than those attributable to free water, we propose that bonded water opens new free-volume pores in amorphous interchain regions in addition to swelling existing pores. A schematic illustration of these free-volume changes is depicted in Figure 10.

CONCLUSIONS

In this study, we have investigated the macroscopic swelling and accompanying free-volume changes of TO-CNF films. While these films swell according to expectation when subjected to 90% RH water vapor, swelling is reduced when the film drying temperature is increased. This observation is attributed to higher film density, as verified by SEM and

Table 5. Weighted PALS Intensities of Water and TO-CNF Films Swollen at 90% RH

equilibrium swelling (%)	f_w	$f_w \times I_{3w}$	$f_c \times I_{3c}$	I_3 (%) from eq 6
31.4	0.239	$0.239 \times 26.9 = 6.43$	$0.761 \times 13.98 = 10.64$	17.07 ± 0.2
35.0	0.259	$0.259 \times 26.9 = 6.97$	$0.741 \times 13.98 = 10.36$	17.33 ± 0.2

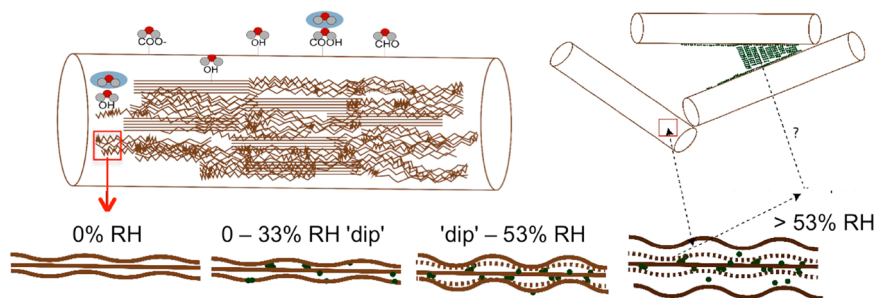


Figure 10. Proposed model for water–cellulose interactions in the amorphous regions of cellulose chains within CNF (rod). During the initial transient swelling between 0 and ~33% RH, water fills intra/interchain free-volume pores and reduces the measured free volume. Beyond this transient and up to ~53% RH, the free-volume pores expand without the creation of new pores. Beyond ~53% RH, water generates the formation of new pores by displacing chains, thus leading to simultaneous increases in *o*-Ps intensity and pore radius.

645 suggesting enhanced hornification. In marked contrast, the total
 646 free volume of fully dried films is not affected by the drying
 647 temperature even though the pore size decreases and the pore
 648 number density increases with increasing drying temperature.
 649 Suspension sonication prior to casting serves to increase the
 650 free-volume pore size due to a combination of disruption and
 651 size reduction of the TO-CNF fibrils. Subjecting TO-CNF films
 652 to various RH levels reveals free-volume pore filling at low
 653 levels (<33% RH). As the RH is increased, free-volume pores
 654 expand. At RH levels beyond ~53%, water likely creates new
 655 free-volume pores and behaves as free water inside the film,
 656 thereby eliciting a free-water PALS signal. This conclusion is
 657 supported by comparing PALS spectra with free-volume
 658 intensity and lifetime calculations. Lastly, swollen films that
 659 are subjected to redrying exhibit slight free-volume pore size
 660 hysteresis. This study establishes a direct relationship between
 661 macroscale and nanoscale water sorption in TO-CNF films
 662 prepared under several different conditions.

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669 Notes

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