# Nutritional correlates of spatio-temporal variations in the marine habitat use of brown trout, Salmo trutta, veteran migrants 

Xavier Bordeleau ${ }^{1^{*}}$, Jan G. Davidsen ${ }^{2}$, Sindre H. Eld $\varnothing y^{2}$, Aslak D. Sjursen ${ }^{2}$, Fred G. Whoriskey ${ }^{3}$, Glenn T. Crossin ${ }^{1}$<br>${ }^{1}$ Department of Biology, Dalhousie University, 1355 Oxford Street, P.O. Box 15000, Halifax, Nova Scotia, B3H 4R2, Canada / xavier.bordeleau@dal.ca, glenn.crossin@dal.ca<br>${ }^{2}$ NTNU University Museum, Norwegian University of Science and Technology, NO-7491 Trondheim, Norway / jan.davidsen@ntnu.no, sindre.eldoy@ntnu.no, aslak.sjursen@ntnu.no<br>${ }^{3}$ Ocean Tracking Network, Dalhousie University, 1355 Oxford Street, P.O. Box 15000, Halifax, Nova Scotia, B3H 4R2, Canada / fwhoriskey@dal.ca<br>* Corresponding author: Xavier Bordeleau / Mobile: 1-819-238-8762 / Department of Biology, Dalhousie University, 1355 Oxford Street, P.O. Box 15000, Halifax, Nova Scotia, B3H 4R2, Canada / Fax: 1-902-494-3736 / xavier.bordeleau@dal.ca


#### Abstract

The brown trout (Salmo trutta) is an iteroparous, anadromous salmonid that exhibits a complex continuum of feeding migration tactics, ranging from freshwater residency, to potamodromy, to estuarine migration, as well as short-to-long distance coastal migrations. While anadromous migrants are believed to play an important role in the species' population dynamics, little is known about the factors driving differences in the extent of individual marine habitat use. In this study, 32 brown trout veteran migrants were acoustically tagged prior to their seaward migration and sampled for indices of their nutritional state. Our findings suggest that: i. body condition factor differed amongst fish adopting different migratory tactics, with outer-fjord migrant being in poorer condition; and ii. within migratory groups, plasma triglyceride concentration was negatively correlated with the duration of marine residency. Results support the idea of condition-dependent migration in veteran migrants, with individual variation in nutritional state influencing the spatio-temporal aspects of marine habitat use. Furthermore, overall marine minimum survival during the summer feeding migration was $86 \%$, the highest reported estimate for this life-stage.


Keywords: alternative migratory tactics, marine migrations, marine survival, plasma triglycerides, sea trout

## Introduction

From an evolutionary point of view, migration can be regarded as an individual adaptation to the changing requirements of life-histories and resource availabilities (Dingle 2006). As an individually expressed trait, migration can also be viewed as a syndrome, shaped by natural selection, wherein correlated behavioural, physiological, morphological and other traits combine to maximize fitness within particular life-history contexts (Dingle 2006, e.g.; Peiman et al. 2017). As such, partial migration has been described in a variety of taxa, in which variable proportions of a population are either migratory or non-migratory (Chapman et al. 2011). While differences in migratory strategies can have both environmental and genetic links, the degree to which these contribute to an expressed migratory strategy can vary widely (Pulido 2011). In anadromous brown trout (Salmo trutta) for example, little genetic variation seems to exist among migrant and resident individuals living sympatrically (Hindar et al. 1991). According to the conditional strategy concept, different tactics regulated by the same genotype can be maintained within populations. This can occur when the migratory decisions and fitness gained from alternative tactics depend primarily on individual phenotype, e.g., age, size, sex, energetic state, etc. (Repka and Gross 1995).

The brown trout is a facultative anadromous, iteroparous salmonid species native to the cold waters of Eurasia and North Africa. The species is socially and economically important, and because of its high adaptability and phenotypic plasticity, it has been successfully introduced to every continent except Antarctica (MacCrimmon and Marshall 1968). Within a population, the species shows complex variation in its feeding migratory tactics, a continuum ranging from freshwater residency, to potamodromy, and estuarine migration (Cucherousset et al. 2005;

Boel et al. 2014), as well as beyond, to short- and long-distance coastal migration (del VillarGuerra et al. 2013; Eldøy et al. 2015; Flaten et al. 2016). This migratory continuum has been observed in both first-time migrants (i.e., parr and smolts; del Villar-Guerra et al. 2013; Boel et al. 2014; Flaten et al. 2016), as well as veteran migrants (i.e., either mature or immature individuals that have previously completed a first summer feeding migration; Eldøy et al. 2015). While the drivers of the different migratory tactics remain obscure, especially for the marine migrations of veteran migrants (Drenner et al. 2012; Thorstad et al. 2016), previous studies of first-time migrants suggest that the choice of migratory tactic is a plastic response to individual physiological state, metabolic rate, and food availability (Wysujack et al. 2009). In experimental feeding studies, brown trout exposed to low food availability had lower pre-migratory body condition factors, which has been shown to influence the decision of first-time migrants to adopt anadromy, whereas higher-condition fish tended towards non-anadromy (Davidsen et al. 2014; Olsson et al. 2006; Wysujack et al. 2008). Wysujack et al. (2008) similarly concluded that energetic state (i.e., body lipid content) close to the time of actual migration was likely responsible for the choice of migratory tactic. A recent telemetry study of the marine migrations of brown trout veteran migrants concluded that migratory decisions were likely influenced by body condition (Eldøy et al. 2015). However, the link between individual premigratory nutritional state and the spatio-temporal extent of subsequent marine habitat use by brown trout has not been well-defined (Aldvén and Davidsen 2017).

Following marine feeding migrations, mostly occurring in summer, anadromous brown trout usually return to freshwater to spawn and/or over-winter. During the winter, feeding opportunities in freshwater can be sparse, and so brown trout depend on somatic lipid stores
for survival, especially in post-spawned individuals (summarized in Jonsson and Jonsson 2011). Lipids stored in adipose and muscle tissues are catabolized and released into circulation as triglycerides. Triglycerides are delivered to target tissues, where hydrolysis (lipolysis) produces glycerol and free fatty acids, the major source of metabolic energy production in fish (Sargent et al. 2002). During starvation, plasma triglyceride concentration diminishes in an effort to sustain basal metabolic processes (Kakizawa et al. 1995). As such, plasma triglyceride concentration can be used as an indicator of nutritional status in many taxa including wild brown trout (Boel et al. 2014; Gauthey et al. 2015), and is therefore a good candidate parameter for testing hypotheses about condition-dependent models of migratory behaviour in wild animals. However, virtually nothing is known about how natural variation in pre-migratory levels of plasma triglycerides might affect the migratory decisions of brown trout veteran migrants.

By providing enhanced growth opportunities, the marine environment is believed to support higher abundances and more productive brown trout populations (Thorstad et al. 2016). Marine foraging, especially in areas where nutrient rich feeding opportunities are numerous (e.g., pelagic fish species, Davidsen et al. 2017), allows anadromous individuals to attain larger sizes than their freshwater resident counterparts, which for females translates to a higher fecundity-at-age (Jonsson and Jonsson 1993). Coupled with the higher prevalence of anadromy in females (as increased size represents a more direct fitness gain for females), it is likely that anadromous brown trout make important contributions to the species' population dynamics (Thorstad et al. 2016), although this has yet to be fully investigated. With the recent increase in marine mortality and decreased growth of anadromous brown trout due to
anthropogenic impacts on marine habitats (Thorstad et al. 2015), researchers have speculated that a reduction in the benefits of anadromy might favour selection for freshwater residency (Hendry et al. 2004; Thorstad et al. 2016). Better knowledge about the whereabouts of brown trout veteran migrants at sea and the endogenous factors affecting the extent of the marine migrations (e.g., distance and duration) will contribute to a fuller understanding of the drivers of marine habitat use of this important life-history stage. Such information is currently lacking but needed for improving current management and conservation actions.

To address these knowledge gaps, we combined acoustic telemetry and physiological sampling techniques within a fjord system in Northern Norway to quantify variation in migratory tactics and in the extent of marine habitat use of brown trout veteran migrants (e.g., duration of marine residency, distance travelled), as well as the influence of pre-migratory nutritional state (i.e., body condition factor, plasma triglyceride concentration). Utilizing this approach, we addressed the over-arching hypothesis that nutritional state underlies interindividual differences in migratory behaviour. We tested the following predictions: i. individuals in poorer nutritional state (i.e., low body condition factor, low plasma triglyceride concentration) have a stronger tendency towards marine migration, and this tendency would be emphasized in females; ii. within marine migrants, fish in poorer nutritional state migrate further out; iii. fish in poorer nutritional state require more time in the marine environment to recondition; and iv. marine survival (i.e., to completion of the summer feeding migration) is lower for fish migrating to the outer reaches of the fjord.

## Materials and methods

## Study area and acoustic receiver array

The study was conducted in the inter-connected, marine, Tosenfjord and Bindalsfjord $\left(65.13^{\circ} \mathrm{N}, 12.13^{\circ} \mathrm{E}\right)$ in Nordland country, Norway (Fig. 1). Brown trout were captured from two different watercourses (i.e., freshwater systems): Åbjøra and Urvold. Both watercourses, situated $\sim 14 \mathrm{~km}$ apart, drain into the Tosenfjord ( $\sim 97 \mathrm{~km}^{2}$, maximum depth of $\sim 550 \mathrm{~m}$ ) which then leads to the Bindalsfjord ( $\sim 91 \mathrm{~km}^{2}$, maximum depth of $\sim 700 \mathrm{~m}$ ) and finally to the Atlantic Ocean, located at $\sim 33 \mathrm{~km}$ from Åbjøra and Urvold estuaries (defined as the interface between river mouth and fjord). During the study period, from April 8 - September 5, 2015 the surface salinity level ( $\sim 1 \mathrm{~m}$ depth) of the Åbjøra estuary (i.e., Floet, station 69 in Fig. 1) and the outerfjord (station 1) varied respectively from $0.0-28.6 \mathrm{ppt}($ mean $=9.4 \mathrm{ppt})$ and $4.8-33.5 \mathrm{ppt}$ (mean = 20.0 ppt ). Surface water temperature ( $\sim 1 \mathrm{~m}$ depth) for those two sections of the fjord varied from $3.0-17.6^{\circ} \mathrm{C}$ and $4.5-17.0^{\circ} \mathrm{C}$, respectively. The eastern section of Tosenfjord is similar to other inner-fjord areas in terms of depth, temperature, and surface salinity level that varies between $\sim 0-34$ ppt depending on areas, freshwater inputs, and tidal cycles.

In Åbjøra, the tagging site was located 2.7 km upstream of the river mouth (around station 65, Fig. 1). This area is situated in the tidal zone of the river with a surface salinity level that varied from $7.4-22.3 \mathrm{ppt}($ mean $=10.1 \mathrm{ppt})$ and water temperature from $2.7-16.7^{\circ} \mathrm{C}$ during the study period, depending on tidal cycle and freshwater discharge. Spawning in the Åbjøra watercourse occurs in River Åbjøra and its tributaries (> 2 km upstream of the tagging area), while the lower part of the system includes deep pools and slow currents, and is consequently considered suitable overwintering habitat. In Urvold, the tagging site was situated in a freshwater lake, near the outlet (around station 61, Fig. 1). Lake Urvold has a surface area of $\sim 0.6 \mathrm{~km}^{2}$ and is $\sim 80 \mathrm{~m}$ deep, which provides good overwintering habitat for brown trout. The
lake then drains into the fjord through a 0.2 km riverine stretch. Unlike Åbjøra, the transition from freshwater to the fjord is much more direct in Urvold due to the steepness of the short river section. In this watercourse, the main spawning site is located upstream of the lake.

## Capture, blood sampling and tagging

A total of 32 brown trout veteran migrants were captured, sampled (i.e., for blood, measurements and scales), and tagged with an internal acoustic transmitter between April 912, 2015 prior to their seaward feeding migration; 20 individuals from the tidal zone in river Åbjøra and 12 from lake Urvold (Table 1). At capture, veteran migrants were distinguished from smolts and parr based on size, which was later confirmed by scale readings. Fish were captured using a combination of angling ( $n=26$ ) and gill netting ( $n=6 ; 35-42 \mathrm{~mm}$ mesh size), and efforts were made to reduce stress and the risk of injury by minimizing fight times and monitoring the gill net continuously (e.g., the net was pulled-in as soon as vibrations were felt). The average time-to-blood-sampling, as calculated from the first encounter with fishing gear, was similar for both methods; 7:51 $\pm$ 5:15 min for angling and 10:32 $\pm 5: 20 \mathrm{~min}$ for gill netting ( $p$-value $=0.29$; Welch Two Sample t-test). Shortly after landing and prior to anaesthesia, $\sim 2 \mathrm{ml}$ blood samples were collected via caudal venipuncture from all fish (as described in Huston 1990). Samples were then placed in an ice-water slurry until processing. Following blood sampling, fish were kept for no more than 2 hours in a net cage situated in the river to allow recovery from capture and sampling before the tagging procedure. Prior to tagging, brown trout were anesthetized using 2-phenoxyethanol at a concentration of $0.5 \mathrm{ml} \mathrm{L}^{-1}$ water (SIGMA Chemical Co., USA). Depending on fish size, 69 kHz MP-9 (for fish between 310-390 mm in total length) or 69 kHz MP-13 acoustic transmitter (for fish between 400 to 720 mm in total length) were inserted in
the abdominal cavity (nominal delay of $30-90$ seconds, Thelma Biotel AS, Norway), according to standard operating procedures (as described in Cooke et al. 2011). MP-9 tags ( 5.2 g in air; estimated battery life of $\sim 15$ months) were 1.0-1.8\% of fish total body mass, and MP-13 (11.8 g in air; estimated battery life of $\sim 24$ months) were $0.4-2.3 \%$ of fish total body mass. Immediately after surgical transmitter implant, fish were weighed and measured (i.e., total length), $\sim 5-10$ scales were sampled and stored for later aging, and a small adipose fin clip was collected for genetic sexual determination. Following tagging, the brown trout were released back at their respective capture site, and subsequently detected on an acoustic receiver array composed of 54 VR2W-69 kHz receivers (Vemco Ltd., Canada) deployed throughout the lakes, rivers, and marine fjords (Fig. 1). The array was established prior to the start of tagging and was in place until fall 2017 (although we are focusing on 2015 detections in the current study). The experimental procedures followed national ethical requirements and were approved by the Norwegian National Animal Research Authority (permit number: 7277).

## Blood processing and triglycerides assay

Within three hours of blood sampling, samples were centrifuged at 3300 g for 10 minutes and the resultant plasma was collected and flash-frozen in dry ice before being transferred to a $-80^{\circ} \mathrm{C}$ freezer. Plasma triglyceride levels were assayed in duplicate using a commercially available colorimetric kit (Cayman Chemical Company, USA) and read at 530 nm with a BioTek Synergy HTX microplate reader (BioTek Instruments, Inc., USA), according to the manufacturer's standard procedure. The mean coefficient of variation between duplicates was 3.9\%.

## DNA sex determination

DNA was extracted from ethanol preserved fin clips with the QuickExtract kit (Epigen), according to manufacturer's protocol with the exception for the extraction volume, which was reduced to $150 \mu$ l. Sex was determined by PCR amplification of a 200 pb fragment situated in the first intron of the male specific SDY gene, using the Salmo-sdY-F and Salmo sdY-R primers (Quéméré et al. 2014). The PCR was performed in $10 \mu \mathrm{l}$ reactions using the Qiagen Multiplex PCR kit. The following PCR profile was used: $95^{\circ} \mathrm{C}$ for $15 \mathrm{~min}, 11$ cycles of touchdown $\mathrm{PCR}, 94^{\circ} \mathrm{C}$ for 30 seconds, $63-52^{\circ} \mathrm{C}$ for 30 seconds, $72^{\circ} \mathrm{C}$ for 1 minute, followed by 25 cycles of $94^{\circ} \mathrm{C}$ for 30 seconds, $52^{\circ} \mathrm{C}$ for 30 seconds, $72^{\circ} \mathrm{C}$ for 1 minute, with a final extension at $72^{\circ} \mathrm{C}$ for 10 minutes. Sex was scored by running the PCR products on 1\% Agarose gels. Sex could be determined with confidence for all but one fish, so this individual had to be excluded from final models including sex as an explanatory variable.

## Data analysis

Individual migratory behaviours can be classified into distinct tactics. First, brown trout are either riverine/lacustrine resident or marine migrant. Then, marine migrants could be further divided into groups depending on where and how far they migrated. To test the overarching hypothesis that nutritional state underlies inter-individual differences in migratory behaviour, statistical tests were performed to assess how individuals' characteristics influenced the different spatial and temporal aspects of migration, as well as survival (specific predictions are presented in the sections below). All individuals' morphological (i.e., length, mass, body condition factor) and physiological (i.e., plasma triglyceride concentration) characteristics, as well as sex and population of origin were considered as explanatory variables. However, due to limited sample sizes, final statistical models were limited to the inclusion of a maximum of two
explanatory variables. Selection of the best model for each aspect of individuals' migratory behaviour was determined using a forward stepwise approach (based on AIC values and comparison with a null model; Anderson et al. 2001). Body condition factor (i.e., Fulton's K) was calculated from the formula: $K=100 \times$ Mass $[\mathrm{g}] \times$ Total length $[\mathrm{cm}]^{-3}$, following the observation that the regression coefficient of the mass-length relationship was 2.99 for tagged individuals, thus confirming the assumption of isometric growth. For each statistical model, we first verified that underlying assumptions were met. Out of 32 acoustically tagged individuals, four were excluded from further analyses due to abnormal patterns of detections: one transmitter was continuously detected on a single station (no. 63 in Fig. 1) for about five months before disappearing, two transmitters were not detected anywhere in the array after April $22^{\text {nd }}$, and one transmitter seems to have experienced technical failure as soon after release not a single detection was registered.

Potential false detections were first flagged using the "falseDetectionFilter" function of the "glatos" package in R , requiring a minimum of two detections from a given transmitter in a time span of 30 minutes at given receiver. Then, flagged detections that were legitimate were kept (i.e., subsequent detections on other receivers located nearby) and others were deleted. To evaluate the detection efficiencies of various portions of the acoustic array, we examined 11 receiver gates, where spacing between receivers was 400 m and the distance from shore to the nearest receivers was 200 m . We calculated the percentage of events in which an individual was detected on a gate when crossing from one side to the other (as reveled from detections on other receivers). For example, in Fig. 1, if a fish was first detected on station 43 and then on station 16 , we controlled if that fish was properly detected on the gate formed by stations 40 ,

41 and 42 , as it had to cross that gate. Overall, out of a total of 203 gate-crossing events that occurred during the entire study period, individuals were detected on the gate $92 \%$ of the time. The detection efficiency (and location) of individual gates were as follow: $100 \%$ in Åbjøra estuary (stations $67-68$, Fig. 1); 100\% on Osane opening gate (stations 31-32); 91\% on Terråk gate (stations $33-35$ ); 100\% on Sørfjord gate (stations $28-30$ ); 100\% on Bindalsfjord west gate (stations $11-13$ ); 95\% on Bindalsfjord east gate (stations $16-17$ ); 86\% on Tosenfjord outer gate (stations 40 - 42); 81\% in Urvold estuary (stations 43 - 44); 100\% on Tosenfjord central gate (stations $45-47$ ); 100\% on Tosenfjord east gate (stations $18-20$ ); and $100 \%$ on Tosbotn gate (stations 26, 52, 53).

## i. Riverine/lacustrine residency versus marine migration

In Åbjøra, the tidal zone of the river offers a brackish environment in which some individuals may decide to stay into to feed. While these fish are not true freshwater resident, they were regrouped as riverine "resident" to be distinguish from marine migrants that ventured into the fjord system. In Urvold, however, the transition from freshwater to the brackish waters of the fjord is much more direct and fish can either decide to reside within the freshwater lake or to move into the fjord, and were unlikely to reside in the short river section. For this reason, models were developed separately for each population. To test the prediction that individuals in poorer nutritional state (i.e., low body condition factor and depleted plasma triglyceride level) have a stronger tendency towards marine migration, binomial logistic regression models using the logit link were performed ("glm" function in R; two outcomes: staying in the river/lake, or migrating to the fjord/marine environment).
ii. Marine migratory tactics

Once individuals were detected entering the fjord, fish from both populations were faced with similar habitat choices. Based on migratory distances and the areas of the fjord that were utilized, three different marine migratory tactics were identified (Fig. 1); i. short-distance migration (furthest detection <2 km from the river mouth; Fig. 2a); ii. long-distance/inner-fjord migration (furthest detection from $\sim 13$ to 28 km of the river mouth, without any detection on the outer receiver line, Fig. 2b); and iii. long-distance/outer-fjord migration (furthest detection $>21 \mathrm{~km}$ from the river mouth and detected on the outer receiver line; Fig. 2c). To test the prediction that, within marine migrants, fish in poorer nutritional state (i.e., low body condition factor and depleted plasma triglyceride level) migrate further out in the marine environment, multinomial logistic regression models were performed ("multinom" package in R), suitable for nominal categorical response variable with three outcomes, here: short-distance, long-distance/inner-fjord, or long-distance/outer-fjord migration (Kwak and Clayton-Matthews 2002; Zuur et al. 2007a). The p -values of the regression coefficients were computed using Wald ztests. Furthermore, to facilitate the interpretation of the model's coefficients, changes in the predicted probabilities of adopting a given migratory tactic as a function of explanatory variables were calculated for each tactic using the "predict" function in R (Kwak and ClaytonMatthews 2002).

## iii. Marine residency period

The marine residency period could be calculated for a total 14 individuals (5 fish from Åbjøra and 9 from Urvold) that were detected entering the fjord in the spring and returning to the rivers later in the summer. For those fish, the marine residency period started at the time of first detection at a river mouth/estuarine receiver (stations 67-68 in Åbjøra, and 43-44 in

Urvold, Fig. 1) and ended at the time of last detection at a river mouth/estuarine receiver (conditional to later detection on upstream receiver, station 63 in Åbjøra, and 61 in Urvold). To test the prediction that fish with low body condition factor and depleted plasma triglyceride level would remain in the marine environment for longer periods of time, general linear regression models were performed ("Im" function in R).

## iv. Marine survival

The marine minimum survival was estimated from the initial number of fish that were detected entering the fjord system and the proportion of them that came back to the rivers, completing their summer marine migration (as indicated by detection at a river mouth/estuarine receiver conditional to later detection on upstream receivers). In addition to those fish, marine migrants that were detected returning to Åbjøra or Urvold estuary (stations 67-69 in Åbjøra, and 43-44 in Urvold, Fig. 1) in late summer and fall were also deemed to have survived their marine summer feeding migration, despite not re-entering freshwater systems. These fish were assumed to be over-wintering in estuaries, as they were detected there through the fall and early winter. Marine migrants that presumably died or disappeared from the acoustic array through the summer might be the result of tag lost, technical failure of the tag, exhaustion of tag battery life, residency in uncovered areas, or animal mortality. To test the prediction that survival to completion of the summer feeding migration is lower for long-distance/outer-fjord migrants in comparison with other marine migratory tactics, binomial logistic regression models using the logit link were performed ("glm" function in R).

## Results

## i. Riverine/lacustrine residency versus marine migration

Out of the 20 brown trout tagged in Åbjøra, four individuals had to be removed from the analysis as their migratory decision could not be identified with confidence (as described previously). From those remaining ( $n=16$ ), 7 individuals (44\%) opted to reside in the tidal zone in the river for the summer, moving in and out of Floet but never entering the fjord (Table 1). In contrast, 9 individuals (56\%) moved out of the river between April $10^{\text {th }}$ and May $29^{\text {th }}$ (median of May $2^{\text {nd }}$ ) migrating into the fjord. The prediction that individuals in poorer nutritional state (i.e., low body condition factor and depleted plasma triglyceride level) would have a stronger tendency towards marine migration was not supported by our results, as the null model had a lower AIC value than alternative models. In addition, no statistically significant morphological differences (i.e., length and mass) were found between riverine resident and marine migrants (p-values > 0.31; Welch Two Sample t-tests). However, as expected, females tended to migrate into the fjord in higher proportion than males, as 8 out of 11 females ( $73 \%$ ) left the river compared to only 1 out of 5 males ( $20 \%$ ) ( $p=0.070$ for sex; binomial logistic regression), although marginally non-significant - which, considering the effect size, is likely due to small sample size. In contrast to Åbjøra, all 12 brown trout tagged in freshwater, near the outlet of Lake Urvold, migrated into the fjord to feed in the marine environment irrespective of sex, morphology and physiology. Those individuals migrated into the fjord between April $25^{\text {th }}$ and May $15^{\text {th }}$ (median of May $10^{\text {th }}$ ). As all fish from this population migrated, we could not test the hypothesis that individuals in poorer nutritional state would have a stronger tendency towards anadromy.

## ii. Marine migratory tactics

For marine migrants (pooled sample of 9 fish from Åbjøra and 12 from Urvold), 3
individuals (14\%; 2 females and 1 male) were classified as short-distance migrants, 7 individuals (33\%; 2 females and 5 males) were classified as long-distance/inner-fjord migrants, and 11 individuals (52\%; 6 females, 4 males, one unknown sex) were classified as long-distance/outerfjord migrants (as summarized in Table 1). In assessing which factors might influence marine migratory decisions, the best fitting multinomial logistic model included body condition factor and sex as explanatory variables (Table 2). The other explanatory variables considered, i.e., population, plasma triglycerides, length and mass, were not retained in the model during the stepwise process. Because population of origin was not retained as an influential explanatory variable, and in combination with the observation that a mix of fish from both populations were present in all three migratory tactics (Table 1), we pooled fish from both populations in a single model. In support of our second prediction, for long-distance migrants, the probability of outerfjord migration decreased significantly with increasing body condition factor (-20.7 in the log odds of migrating to the outer-fjord following a one unit increase in body condition, $p=0.027$, Table 2). Thus, fish migrating to the outer part of the fjord were generally in the poorest condition prior to migration. Comparing sexes, for long-distance migrants, the log odds of migrating to the outer section of the fjord versus staying in the inner part increased by 3.15 for females, in which $75 \%$ (6/8) migrated to the outer section versus $44 \%(4 / 9)$ in males (although marginally non-significant at $p=0.058$, Table 2 ). To facilitate interpretation, the predicted probabilities of adopting a given marine migratory tactic as a function of body condition factor and sex were calculated using the regression coefficients (Fig. 3). Body condition factor and sex mostly affected the probabilities of adopting the long-distance/outer-fjord versus long-distance/inner-fjord tactic, with no significant influence on the decision to undertake a short-
distance marine migration (Table 2). However, we interpret the latter result with caution due to the low number of short-distance marine migrants. Moreover, following the observation that no significant morphological and physiological differences (i.e., in length, mass, body condition factor, and plasma triglyceride level) existed between long-distance/inner-fjord and shortdistance migrants ( $p$-values $>0.68$; Welch Two Sample t-tests), the two tactics were pooled to take a closer look at what might be driving the decision to migrate to the outer section of the fjord (reducing the tests to only two marine migratory outcomes: outer-fjord versus inner-fjord migrants). In this analysis, female outer-fjord migrants had significantly lower body condition factor prior to migration compared to inner-fjord migrants ( $0.79 \pm 0.05$ vs $0.92 \pm 0.05, p=$ 0.006, Welch Two Sample t-test, Fig. 4). A similar, but non-significant difference in body condition was observed in males. Pooling both sexes, outer-fjord migrants had significantly lower body condition $(0.75 \pm 0.09)$ than inner-fjord migrants $(0.87 \pm 0.10, p=0.005$, nonparametric Mann-Whitney $U$ test). While male outer-fjord migrants were significantly smaller than inner-fjord migrants ( $433 \pm 59 \mathrm{~mm}$ versus $576 \pm 100 \mathrm{~mm}, \mathrm{p}=0.022$, Welch Two Sample ttest), females showed the opposite tendency with outer-fjord migrants being generally larger (range: 400-640 mm) than inner-fjord migrants (range: 310-445 mm, although the difference in mean length was not significant, Fig. 4). Supporting the assumption that plasma triglyceride concentration is informative of individuals' post-winter/pre-migratory nutritional state, body condition factor and plasma triglycerides were correlated at $r=0.45$ ( $p=0.009$ ). Interestingly, females displayed a negative correlation between length and pre-migratory body condition factor ( $r=-0.57, p=0.02$ ), and similarly between length and plasma triglyceride concentration $(r=-0.48, p=0.06)$, so that larger females were generally found in poorer pre-migratory
condition. This negative relationship between body condition factor and length was not observed in males.

## iii. Marine residency period

Of the individuals that were detected entering the fjord in the spring ( $N=21$ ), a total of 14 individuals (5 from Åbjøra and 9 from Urvold) came back to the river/lake later in the spring or during the summer (May $23^{\text {rd }}$ to July $16^{\text {th }}$; median of July $4^{\text {th }}$ ). Individuals' marine residency period varied between 32 and 83 days (average of 59 days). Looking at the factors that might explain inter-individual variance in the marine residency period, the best fitting linear regression model included plasma triglyceride concentration and marine migratory tactic (Table 3 ; multiple $R^{2}=0.60$ ). All other potential explanatory variables (population of origin, body condition factor, length, mass, and simple interactions) were not retained in the model during the stepwise process. This, combined with the fact that no significant differences existed in the marine residency period between fish from the two study populations ( $p>0.20$; Welch Two Sample t-test), allowed us to pool fish from the two populations. Out of the 14 individuals that came back to freshwater, 7 were long-distance/outer-fjord migrants, 6 were long-distance/inner-fjord migrants, and only one was a short-distance migrant. In order to meet the assumptions of general linear regression models (i.e., independence, normality of residuals, homoscedasticity, and balanced influence of individual observations), the short-distance migratory tactic $(N=1)$ had to be removed from the final model as it had high leverage (hat value 3.5 -fold greater than the average, "hatvalue" function in R) which led to a deviation from normality (Zuur et al. 2007b). In support of our third prediction, within migratory groups, the duration of marine residency was negatively correlated with pre-migration plasma triglyceride
concentration (Table 3), such that depleted individuals generally spent more time in the marine environment ( $\sim 24$ more days for a decrease of $1 \mathrm{mmol} \mathrm{L}^{-1}$ in triglyceride level). In addition, long-distance/inner-fjord migrants spent on average $69.2 \pm 11$ days in the marine environment, $\sim 15$ days more than long-distance/outer-fjord migrants ( $54.6 \pm 16$ days), a significant difference controlling for the effect of plasma triglyceride level (Table 3; Fig. 5). The only short-distance migrant for which marine residency could be calculated spent only 35 days there (Table 1).

## iv. Marine survival

Out of a total of 21 veteran migrants that were detected entering the fjord in the spring, seven were not detected coming back to freshwater. Of those, four individuals (i.e., three females and one male) were presumed to have survived but opted for over-wintering in the marine environment as they were detected in estuaries in the fall and winter (summarized in Table 1), apparently deciding to skip spawning. Those three females were generally smaller (range: $310-420 \mathrm{~mm}$ ) than the five females that had migrated back to freshwater (range: 405640 mm ), and were possibly still immature (Klemetsen et al. 2003). For the remaining three individuals that neither returned to freshwater nor over-wintered in estuaries, two disappeared from the array (as they were last detected in late May/early June in the outer section of the fjord, and were never heard from again despite the array remaining in place until fall 2017) and one was continuously detected on a single receiver for $>6$ months starting in mid-June. Minimum marine survival through the summer feeding migration was thus estimated at $86 \%$, as at least 18 out of 21 individuals that migrated out into the fjord survived to return to freshwater, or were detected in estuaries in the fall and winter. In partial support of our fourth prediction, all three fish that presumably died or disappeared at in the marine environment
were long-distance migrants, with significantly lower body condition factor ( $0.75 \pm 0.02$ ) than the average marine migrant ( $0.82 \pm 0.12, p=0.048$; Welch Two Sample t-test), implying a $73 \%$ minimum survival for this marine migratory tactic ( $N=11$ ) versus $100 \%$ survival for the other two tactics $(\mathrm{N}=10)$. However, no morphological or physiological individual characteristics, nor sex, population or marine migratory tactics statistically influenced the probability of marine survival, as the null model had lower AIC value than alternative binomial models.

## Discussion

Our findings support the over-arching hypothesis that pre-migratory nutritional state, as indicated by body condition factor and plasma triglyceride concentration, is correlated with spatio-temporal variations in the marine habitat use of brown trout veteran migrants. They also bring some support to the general belief that females are more inclined to migrate to the marine environment.

We found that, after a winter in freshwater, female trout from the Åbjøra watershed showed a higher tendency than males to leave the river and migrate into the fjord in spring. This higher tendency of females towards anadromy is commonly observed in facultative anadromous salmonid populations, and is believed to be driven by the strong relationship between body size and fecundity, with the productivity of the marine environment sustaining faster growth (reviewed in Jonsson and Jonsson 1993). Previous work on the migratory behaviour of brown trout post-smolts suggest a condition-dependent migration, with low body condition factor generally promoting anadromy (Davidsen et al. 2014; Olsson et al. 2006; Wysujack et al. 2008), but not always (del Villar-Guerra et al. 2013; Boel et al. 2014). Despite our first prediction that poorer nutritional state would promote marine migration, no
significant differences in body condition factor and plasma triglyceride concentration were observed between riverine/lacustrine residents and marine migrants.

However, among brown trout that initiated marine migrations, body condition factor differed between migratory tactics. As predicted, brown trout in the poorest relative body condition were those most likely to migrate to the outer section of the fjord, where nutrient rich foraging opportunities are presumably more abundant (e.g., increased dependence upon pelagic fishes, Davidsen et al. 2017), and a greater proportion of female (77\%), than male longdistance migrants (44\%), opted for this tactic then the alternative inner-fjord tactic. While fish of various sizes migrated to the outer reaches of the fjord, female long-distance/outer-fjord migrants were generally larger than female inner-fjord migrants. Interestingly, females displayed negative correlations between length and pre-migratory body condition factor as well as triglyceride concentration, so that larger females were generally found in poorer premigratory condition and were more likely to migrate to the outer-fjord. This negative relationship between body condition factor and length was not observed in males. By comparing immature brown trout, with first-time and repeat-spawners of both sexes, Berg et al. (1998) showed that lipid and protein depletion through the spawning season increased with size in female, but not male. Accordingly, while the energy content of immature individuals increased, the specific energy content of spawning females decreased with length due to increased reproductive investments (Berg et al. 1998). While it was shown that anadromous female brown tout invest more into reproduction than resident counterparts (Elliott 1988; Jonsson and Jonsson 1997), which affects post-spawning nutritional state, our findings suggest that the costs of reproduction might influence future migratory decision by affecting the extent
of individual marine habitat use the following year. Larger, depleted females might be migrating further to sea in an attempt to recondition themselves more effectively and offset the costs of reproduction. However, while the variation in weight and plasma metabolites through spawning is reflective of energy investment to reproduction (Gauthey et al. 2015), postspawning nutritional state alone, as measured in the current study, is not necessarily directly representative of reproductive investment. As such, the link between reproductive investment, post-spawning condition, and subsequent migratory decision is an area requiring further investigation.

In contrast to those initiating outer-fjord migrations, brown trout remaining within the inner-fjord tended to be in better overall condition. Interestingly, these inner-fjord migrants spent significantly more time in the marine environment than outer-fjord migrants (Table 3; Fig. 5). Anadromous trout (brown trout and Arctic charr), both first-time and veteran migrants, generally experience a rapid initial growth phase in the marine environment, which is probably a result of energetic reconditioning after winter-depletion in freshwater (e.g., compensatory growth; Berg and Berg 1987, 1989; Rikardsen et al. 2004). Fish then migrate back to safer freshwater habitats once marine growth potential diminishes later in the season in Northern Norway (Berg and Berg 1987, 1989; Rikardsen et al. 2004). If migration to the outer-fjord was expressly for the purpose of energetic reconditioning for those incurring the greatest costs of reproduction, then it might seem counter-intuitive that they should spend less time there foraging compared to trout in the inner-fjord. This might, however, simply indicate that trout in the outer-fjord need less time to recondition because they are in an area where energy-rich pelagic fish prey are presumably more abundant (see also Eldøy et al. 2015; Davidsen et al.
2017), although there could also be risks associated with migration to the outer-fjord that might limit their time there, such as predation (Lyse et al. 1998) and sea lice parasitism (Thorstad et al. 2015). Within migratory groups that travelled to similar areas of the fjords, marine residency time was negatively correlated with plasma triglyceride levels so that fish with depleted circulating lipid levels spent more time in the marine environment, perhaps reflecting higher nutritional requirements (Table 3; Fig. 5). The duration of the marine residency period is thus probably influenced by individual lipid depletion (or nutritional requirements), and by the relative productivity of the marine habitat to which the fish migrate. As revealed from the significant positive correlation with body condition, plasma triglyceride concentration is believed to reflect post-winter, pre-migratory nutritional condition in these fish that were captured in early spring. Collectively, our findings suggest a condition-dependent migratory tactic in brown trout veteran migrants, in which an individual's pre-migratory nutritional state influences its spatio-temporal use of the marine environment.

In the current study, the survival of marine migrants was estimated at $86 \%$ (18 of 21). This is slightly higher than previously reported marine survival estimates of 29-85\%, for this lifestage (Jensen 1968; Jonsson and Jonsson 2009; Jensen et al. 2014; only including post-spawners in Aarestrup et al. 2015). Interestingly, mortality only occurred among those fish that migrated to the outer fjord. Although sample sizes are small, this might reflect the idea that long-distance migration to more pelagic habitats might be a riskier tactic undertaken by fish in poorer premigratory nutritional condition. This tendency for fish in poorer post-spawning/pre-migratory nutritional condition to opt for a riskier migratory tactic has also been observed in Atlantic salmon, in the form of differential migratory timing. Halttunen et al. (2013) showed that salmon
from the Alta River with lower body condition factors following spawning initiated their seaward migration earlier, and likely encountered unfavorable environmental condition at sea.

Anadromous migrants, and especially females, are believed to play an important role in brown trout population dynamics due to their increased growth potential at in the marine environment and higher reproductive investments (Thorstad et al. 2016). However, large interindividual variation exists in the spatial and temporal extent of the marine habitat use of anadromous migrants. Our results showed that the pre-migratory nutritional state of veteran migrants differed among marine migratory tactics and was associated with the duration of marine residency period. Future research efforts investigating the benefits and costs of different marine migratory tactics in veteran migrants (in terms of survival, growth and fecundity) would contribute to a better understanding of the evolution of the brown trout migratory continuum. Assessing how much intra-individual variability exists in the migratory decisions of individuals tracked through multiple consecutive marine feeding migrations, as it relate to differences in environmental conditions, would also provide novel and highly useful information in the face of rapidly changing environmental conditions.

## Acknowledgments

This study was financed or supported by contributions from Sinkaberg-Hansen AS, the County Governor of Nordland, Nordland County Authority, the Norwegian Environment Agency, the River Åbjøra landowners' association, Plathes Eiendommer and the NTNU University Museum. Additional support was provided by the Ocean Tracking Network via a Travel Grant to XB and via a loan of acoustic receivers to JGD, and by a Natural Sciences and Engineering Research Council of Canada Discovery Grant to GTC. The crew of RV Gunnerus, Lars Rønning,

Jan Ivar Koksvik, Embla $\emptyset$ stebrøt and Hilde Dørum are all thanked for their extensive help during the field work. Marc Daverdin at the NTNU University Museum is thanked for assistance with making the maps.

## References

Aarestrup, K., Baktoft, H., Thorstad, E.B., Svendsen, J.C., Höjesjö, J., and Koed, A. 2015. Survival and progression rates of anadromous brown trout kelts Salmo trutta during downstream migration in freshwater and at sea. Mar. Ecol. Prog. Ser. 535(July): 185-195. doi:10.3354/meps11407.

Aldvén, D., and Davidsen, J.G. 2017. Marine migrations of sea trout ( Salmo trutta ). In Sea Trout: Science \& Management: Proceedings of the 2nd International Sea Trout Symposium. Edited by G. Harris. Troubador Publishing Ltd. pp. 288-297.

Anderson, D.R., Link, W. a, Johnson, D.H., Burnham, K.P., and Press, A. 2001. Suggestions for presenting the results of data analyses. J. Wildl. Manage. 65(3): 373-378. doi:10.2307/3803088.

Berg, O.K., and Berg, M. 1987. The seasonal pattern of growth of the sea trout (Salmo trutta L.) from the Vardnes river in northern Norway. Aquaculture 62(2): 143-152. doi:10.1016/0044-8486(87)90318-8.

Berg, O.K., and Berg, M. 1989. Sea Growth and Time of Migration of Anadromous Arctic Char ( Salvelinus alpinus ) from the Vardnes River, in Northern Norway. Can. J. Fish. Aquat. Sci. 46(6): 955-960. doi:10.1139/f89-123.

Berg, O.K., Thronæs, E., and Bremset, G. 1998. Energetics and survival of virgin and repeat spawning brown trout (Salmo trutta). Can. J. Fish. Aquat. Sci. 55(1): 47-53.
doi:10.1139/cjfas-55-1-47.

Boel, M., Aarestrup, K., Baktoft, H., Larsen, T., Søndergaard Madsen, S., Malte, H., Skov, C., Svendsen, J.C., and Koed, A. 2014. The physiological basis of the migration continuum in brown trout (Salmo trutta). Physiol. Biochem. Zool. 87(2): 334-45. doi:10.1086/674869.

Chapman, B.B., Brönmark, C., Nilsson, J.Å., and Hansson, L.A. 2011. The ecology and evolution of partial migration. Oikos 120(12): 1764-1775. doi:10.1111/j.1600-0706.2011.20131.x.

Cooke, S.J., Murchie, S., Mc Connachie, S., and Goldberg, T. 2011. Standardized surgical procedures for the implantation of electronic tags in key Great Lakes fishes. Technical Report. Great Lakes Fishery Commission, Ann Arbor, MI.

Cucherousset, J., Ombredane, D., Charles, K., Marchand, F., and Bagliniere, J.L. 2005. A Continuum of Life History Tactics in a Brown Trout Salmo Trutta Population. Can. J. Aquat. Sci. 1610: 1600-1610. doi:10.1139/f05-057.

Davidsen, J.G., Daverdin, M., Sjursen, A.D., Rønning, L., Arnekleiv, J. V, and Koksvik, J.I. 2014. Does reduced feeding prior to release improve the marine migration of hatchery brown trout Salmo trutta smolts? J. Fish Biol. 85(6): 1992-2002. doi:10.1111/jfb.12485.

Davidsen, J.G., Knudsen, R., Power, M., Sjursen, A.D., Rønning, L., Hårsaker, K., Næsje, T.F., and Arnekleiv, J. V. 2017. Trophic niche similarity among sea trout Salmo trutta in central Norway investigated using different time-integrated trophic tracers. 26: 217-227.

Dingle, H. 2006. Animal migration: Is there a common migratory syndrome? J. Ornithol. 147(2): 212-220. doi:10.1007/s10336-005-0052-2.

Drenner, S.M., Clark, T.D., Whitney, C.K., Martins, E.G., Cooke, S.J., and Hinch, S.G. 2012. A Synthesis of Tagging Studies Examining the Behaviour and Survival of Anadromous Salmonids in Marine Environments. 7(3): 1-13. doi:10.1371/journal.pone.0031311.

Eldøy, S.H., Davidsen, J.G., Thorstad, E.B., Whoriskey, F., Aarestrup, K., Næsje, T.F., Rønning, L., Sjursen, A.D., Rikardsen, A.H., Arnekleiv, J.V., and Jonsson, B. 2015. Marine migration and habitat use of anadromous brown trout ( Salmo trutta ). Can. J. Fish. Aquat. Sci. 72(9): 1366-1378. doi:10.1139/cjfas-2014-0560.

Elliott, J.M. 1988. Growth, size, biomass and production in contrasting populations of trout Salmo trutta in two Lake District streams. J. Anim. Ecol. 57(1): 49-60. doi:10.2307/4762. Flaten, A.C., Davidsen, J.G., Thorstad, E.B., Whoriskey, F., Rønning, L., Sjursen, A.D., Rikardsen, A.H., and Arnekleiv, J. V. 2016. The first months at sea: marine migration and habitat use of sea trout Salmo trutta post-smolts. J. Fish Biol. 89(3): 1624-1640. doi:10.1111/jfb. 13065.

Gauthey, Z., Freychet, M., Manicki, A., Herman, A., Lepais, O., Panserat, S., Elosegi, A., Tentelier, C., and Labonne, J. 2015. The concentration of plasma metabolites varies throughout reproduction and affects offspring number in wild brown trout (Salmo trutta). Comp. Biochem. Physiol. Part A 184: 90-96. doi:10.1016/j.cbpa.2015.01.025.

Halttunen, E., Lovisa, J., Jensen, A., Næsje, T.F., Davidsen, J.G., Thorstad, E.B., Chittenden, C.M., Hamel, S., Primicerio, R., and Rikardsen, A.H. 2013. State-dependent migratory timing of postspawned Atlantic salmon. Can. J. Aquat. Sci. 1071(9037): 1063-1071.

Hendry, A.P., Bohlin, T., Jonsson, B., and Berg, O.K. 2004. To sea or not to sea? In Evolution illuminated: Salmon and their relatives. Edited by A.P. Hendry and S.C. Stearns. Oxford University Press. Oxford, UK. pp. 92-125.

Hindar, K., Jonsson, B., Ryman, N., and Ståhl, G. 1991. Genetic relationships among landlocked, resident, and anadromous Brown Trout, Salmo trutta L. Heredity (Edinb). 66(1): 83-91. doi:10.1038/hdy.1991.11.

Huston, A.H. 1990. Blood and circulation. In Methods for Fish Biology. Edited by C.B. Schreck and P.B. Moyle. American Fisheries Society. pp. 273-343.

Jensen, J.L.A., Rikardsen, A.H., Thorstad, E.B., Suhr, A.H., Davidsen, J.G., and Primicerio, R. 2014. Water temperatures influence the marine area use of Salvelinus alpinus and Salmo trutta. J. Fish Biol. 84(6): 1640-1653. doi:10.1111/jfb.12366.

Jensen, K.W. 1968. Seatrout (Salmo trutta L.) of the River Istra, western Norway. Rep. Inst. Freshw. Res. (48): 1187-1213.

Jonsson, B., and Jonsson, N. 1993. Partial migration: niche shift versus sexual maturation in fishes. Rev. Fish Biol. Fish. 3(4): 348-365. doi:10.1007/BF00043384.

Jonsson, B., and Jonsson, N. 2009. Migratory timing, marine survival and growth of anadromous brown trout Salmo trutta in the River Imsa, Norway. J. Fish Biol. 74(3): 621-638. doi:10.1111/j.1095-8649.2008.02152.x.

Jonsson, B., and Jonsson, N. 2011. Maturation and spawning. In Ecology of Atlantic Salmon and Brown Trout. Springer. pp. 327-414. doi:10.1007/978-94-007-1189-1.

Jonsson, N., and Jonsson, B. 1997. Energy allocation in polymorphic Brown Trout. Funct. Ecol. 11(3): 310-317. doi:10.1046/j.1365-2435.1997.00083.x.

Kakizawa, S., Kaneko, T., Hasegawa, S., and Hirano, T. 1995. Effects of feeding, fasting, background adaptation, acute stress, and exhaustive exercise on the plasma somatolactin concentrations in rainbow trout. doi:10.1006/gcen.1995.1054.

Klemetsen, a, P-a, A., Jb, D., Jonsson, B., Jonsson, N., O'Connell, M., and Mortensen, E. 2003. Atlantic salmon Salmo salar L ., brown trout Salmo trutta L . and Arctic charr Salvelinus alpinus (L.): a review of aspects of their life histories. Ecol. Freshw. Fish 12: 1-59.

Kwak, C., and Clayton-Matthews, A. 2002. Multinomial Logistic Regression. Nurs. Res. 51(6):

404-410. doi:10.1097/00006199-200211000-00009.
Lyse, A.A., Stefansson, S.O., and Fernö, A. 1998. Behaviour and diet of sea trout post-smolts in a Norwegian fjord system. J. Fish Biol. 52(923): 923-936. doi:10.1006/jfbi.1998.0641.

MacCrimmon, H.R., and Marshall, T.L. 1968. World distribution of brown trout Salmo trutta. J. Fish. Res. Board Canada 25(12): 2527-2548.

Olsson, I.C., Greenberg, L.A., Bergman, E., and Wysujack, K. 2006. Environmentally induced migration: The importance of food. Ecol. Lett. 9(6): 645-651. doi:10.1111/j.14610248.2006.00909.x.

Peiman, K.S., Birnie-Gauvin, K., Larsen, M.H., Colborne, S.F., Gilmour, K.M., Aarestrup, K., Willmore, W.G., and Cooke, S.J. 2017. Morphological, physiological and dietary covariation in migratory and resident adult brown trout (Salmo trutta). Zoology. doi:10.1016/j.zool.2017.07.002.

Pulido, F. 2011. Evolutionary genetics of partial migration - the threshold model of migration revis(it)ed. Oikos 120(12): 1776-1783. doi:10.1111/j.1600-0706.2011.19844.x.

Quéméré, E., Perrier, C., Besnard, A.-L., Evanno, G., Baglinière, J.-L., Guiguen, Y., and Launey, S. 2014. An improved PCR-based method for faster sex determination in brown trout (Salmo trutta) and Atlantic salmon (Salmo salar). Conserv. Genet. Resour. 6(4): 825-827. doi:10.1007/s12686-014-0259-8.

Repka, J., and Gross, M.R. 1995. The evolutionarily stable strategy under individual condition and tactic frequency. J. Theor. Biol. 176(1): 27-31. doi:10.1006/jtbi.1995.0172.

Rikardsen, A.H., Thorpe, J.E., and Dempson, J.P. 2004. Modeling the life history variation of Arctic charr. Ecol. Freshw. Fish 13: 305-311. doi:10.1111/j.1600-0633.2004.00070.x.

Sargent, J., Tocher, D., and Bell, J. 2002. The lipids. In Fish nutrition. Edited by R.W. Halver, J.E.,

Hardy. Academic Press. pp. 181-257.
Thorstad, E.B., Todd, C.D., Uglem, I., Bjørn, P.A., Gargan, P.G., Vollset, K.W., Halttunen, E., Kålås, S., Berg, M., and Finstad, B. 2015. Effects of salmon lice Lepeophtheirus salmonis on wild sea trout salmo trutta - A literature review. Aquac. Environ. Interact. 7(2): 91-113. doi:10.3354/aei00142.

Thorstad, E.B., Todd, C.D., Uglem, I., Bjørn, P.A., Gargan, P.G., Vollset, K.W., Halttunen, E., Kålås, S., Berg, M., and Finstad, B. 2016. Marine life of the sea trout. Mar. Biol. 163(3): 47. Springer Berlin Heidelberg. doi:10.1007/s00227-016-2820-3.
del Villar-Guerra, D., Aarestrup, K., Skov, C., and Koed, A. 2013. Marine migrations in anadromous brown trout (Salmo trutta). Fjord residency as a possible alternative in the continuum of migration to the open sea. Ecol. Freshw. Fish: 594-603. doi:10.1111/eff. 12110.

Wysujack, K., Greenberg, L.A., Bergman, E., and Olsson, I.C. 2009. The role of the environment in partial migration: Food availability affects the adoption of a migratory tactic in brown trout Salmo trutta. Ecol. Freshw. Fish 18(1): 52-59. doi:10.1111/j.1600-0633.2008.00322.x.

Zuur, A.F., leno, E.N., and Smith, G.M. 2007a. Univariate tree models. In Analysing Ecological Data. Springer. pp. 143-162. doi:10.1016/B978-0-12-387667-6.00013-0.

Zuur, A.F., leno, E.N., and Smith, G.M. 2007b. Linear regression. In Analysing Ecological Data. Springer. pp. 49-78. doi:10.1016/B978-0-12-387667-6.00013-0.

## 654

655

## Tables

Table 1. Summary of the migratory behaviour of tagged fish ( $\mathrm{N}=32$ ). The Information is filtered by migratory tactic: riverine/lacustrine residents $(\mathrm{N}=7)$, short-distance migrants $(\mathrm{N}=3)$, long-distance/inner-fjord migrants ( $N=7$ ), long-distance/outer-fjord migrants ( $N=11$ ), and 4 NA's.

| Tag_ID | Pop. | TL | Sex | Marine | Migratory | Marine res. | Marine | Over- |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | (mm) |  | migration | tactic | (days) | survival | winter |
| A69-1303-13 | Åbjøra | 540 | M | 0 | Resident | 0 | NA | NA |
| A69-1303-01 | Åbjøra | 390 | F | 0 | Resident | 0 | NA | NA |
| A69-1303-14 | Åbjøra | 430 | M | 0 | Resident | 0 | NA | NA |
| A69-1303-15 | Åbjøra | 430 | F | 0 | Resident | 0 | NA | NA |
| A69-1303-16 | Åbjøra | 485 | M | 0 | Resident | 0 | NA | NA |
| A69-1303-33 | Åbjøra | 490 | M | 0 | Resident | 0 | NA | NA |
| A69-1303-35 | Åbjøra | 530 | F | 0 | Resident | 0 | NA | NA |
| A69-1303-23 | Åbjøra | 420 | F | 1 | Short mig. | NA | 1 | Estuary |
| A69-1303-19 | Åbjøra | 405 | F | 1 | Short mig. | 35 | 1 | River/lake |
| A69-1303-29 | Urvold | 590 | M | 1 | Short mig. | NA | 1 | Estuary |
| A69-1303-11 | Åbjøra | 445 | F | 1 | Long/in mig. | 83 | 1 | River/lake |
| A69-1303-02 | Åbjøra | 310 | F | 1 | Long/in mig. | NA | 1 | Estuary |
| A69-1303-31 | Urvold | 500 | M | 1 | Long/in mig. | 54 | 1 | River/lake |
| A69-1303-30 | Urvold | 720 | M | 1 | Long/in mig. | 66 | 1 | River/lake |
| A69-1303-28 | Urvold | 628 | M | 1 | Long/in mig. | 80 | 1 | River/lake |
| A69-1303-27 | Urvold | 430 | M | 1 | Long/in mig. | 64 | 1 | River/lake |
| A603-25 | Urvold | 590 | M | 1 | Long/in mig. | 68 | 1 | River/lake |


| A69-1303-17 | Åbjøra | 430 | F | 1 | Long/out mig. | 32 | 1 | River/lake |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| A69-1303-20 | Åbjøra | 490 | M | 1 | Long/out mig. | 43 | 1 | River/lake |
| A69-1303-21 | Åbjøra | 600 | F | 1 | Long/out mig. | 58 | 1 | River/lake |
| A69-1303-22 | Åbjøra | 420 | F | 1 | Long/out mig. | NA | 0 | Dead at sea |
| A69-1303-34 | Åbjøra | 400 | F | 1 | Long/out mig. | NA | 1 | Estuary |
| A69-1303-05 | Urvold | 350 | M | 1 | Long/out mig. | 81 | 1 | River/lake |
| A69-1303-32 | Urvold | 640 | F | 1 | Long/out mig. | 54 | 1 | River/lake |
| A69-1303-22498 | Urvold | 480 | F | 1 | Long/out mig. | NA | 0 | Dead at sea |
| A69-1303-22497 | Urvold | 560 | NA | 1 | Long/out mig. | NA | 0 | Dead at sea |
| A69-1303-26 | Urvold | 450 | M | 1 | Long/out mig. | 50 | 1 | River/lake |
| A69-1303-24 | Urvold | 440 | M | 1 | Long/out mig. | 64 | 1 | River/lake |
| A69-1303-12 | Åbjøra | 490 | F | NA | NA | NA | NA | NA |
| A69-1303-03 | Åbjøra | 310 | F | NA | NA | NA | NA | NA |
| A69-1303-04 | Åbjøra | 320 | M | NA | NA | NA | NA | NA |
| A69-1303-18 | Åbjøra | 430 | F | NA | NA | NA | NA | NA |

Table 2. Output of the best fitting multinomial logistic regression model of the log odds of adopting a given marine migratory tactic versus an alternative tactic (3 outcomes; shortdistance, long-distance/inner-fjord, and long-distance/outer-fjord migrants) as a function of body condition factor ( $K$ ) and sex.

| Migratory tactics | Coefficient | SE | p-value |
| :---: | :---: | :---: | :---: |
| comparison |  |  |  |
| Long/Out vs Long/ln |  |  |  |


| $K$ | -20.7 | 9.4 | 0.027 |
| :--- | :--- | :--- | :--- |


| Sex (F) | 3.2 | 1.7 | 0.058 |
| :--- | :--- | :--- | :--- |

Long/Out vs Short

Long/In vs Short

| $K$ | -15.6 | 9.7 | 0.108 |
| :--- | :--- | :--- | :--- |

Sex (F)
1.3
$1.8 \quad 0.460$

| K | 5.0 | 9.5 | 0.592 |
| :---: | :---: | :---: | :---: |
| (F) | -1.8 | 1.6 | 0.247 |

Table 3. Output of the best fitting general linear regression model of individuals' marine residency period (in days) as a function of plasma triglyceride concentration and marine migratory tactics, 2 outcomes: long-distance/inner-fjord, and long-distance/outer-fjord migrants (baseline outcome).

| Explanatory variable | Coefficient | SE | t-statistic | p -value |
| :---: | :---: | :---: | :---: | :---: |
| Intercept | 67.5 | 5.9 | 11.5 | $4.5 \times 10^{-7}$ |
| Migratory tactic |  |  |  |  |
| Long-dist./in | 15.3 | 5.8 | 2.6 | 0.025 |
| Triglycerides (mmol L-1) | -23.7 | 8.0 | -3.0 | 0.015 |

Multiple $\mathrm{R}^{2}=0.60$

## Figures

Fig. 1. Map of the acoustic array deployed ( $\mathrm{N}=54$ receivers). Receivers deployed in freshwater are represented by squares and those deployed in brackish or saltwater are represented by pentagons. The different coloured areas of the fjords represent the different marine migratory strategies: short-distance in pale grey (two areas; for Åbjøra and Urvold fish), long-distance/inner-fjord in dark grey, and long-distance/outer-fjord in black. Temperature and salinity loggers (T/S) were deployed on stations 1, 65 and 69 . The tagging site was located around station 65 in Åbjøra, and in the lake around station 61 in Urvold. Map data: Norge Digitalt - Statens kartverk.

Fig. 2. Example of observed individuals' marine migratory tactics: a) short-distance migrant; b) long-distance/inner-fjord migrant; and c) long-distance/outer-fjord migrant. The dots represent the position of each acoustic receivers: white crossed dots for receivers on which the individual was not detected; and from yellow to red for receivers on which it was detected (the colour reflecting the number of detections that were registered for that individual on each receivers). Map data: Norge Digitalt - Statens kartverk.

Fig. 3. Predicted probabilities of adopting a given marine migratory tactics as a function of body condition factor and sex, as calculated from the regression coefficients (information presented in Table 2). The shaded areas represent the $95 \%$ confidence intervals.

Fig. 4. Comparisons of body condition factor, plasma triglyceride concentration, and total length in brown trout adopting an outer-fjord (long-distance) in black versus inner-fjord marine
migration tactic in grey (short and long-distance combined). The boxplots show median (black lines) and mean values (white dots), as well as the interquartile ranges (boxes) and the $5^{\text {th }}$ and $95^{\text {th }}$ percentiles (whiskers). Comparisons were made using Welch Two Sample t-tests.

Fig. 5. Marine residency period as a function of pre-migration plasma triglyceride level and marine migratory tactics: long-distance/outer-fjord migrants in black; and long-distance/innerfjord migrants in grey (regression coefficients and p-values can be found in Table 3). The hollow symbol represents the only short-distance migrant for which marine residency period could be calculated. However, this individual was excluded from the analysis for reasons described previously.


Figure 1:


Figure 2:


Figure 3:


Figure 4:


Figure 5:

