

1 To Carbohydrate Polymers
2 Effects of carboxylation of the side chains on the order-disorder transition in
3 aqueous solution of schizophyllan, a triple helical polysaccharide

4
5 Kazuto Yoshiba^{a*}, Satoka Okamoto^a, Toshiaki Dobashi^a, Hiroyuki Oku^a, Bjørn E.
6 Christensen^b, Takahiro Sato^c

7
8 *a. Division of Molecular Science, Graduate School of Science and Technology,*
9 *Gunma University, 1-5-1 Tenjin-cho, Kiryu, Gunma, 376-8515, Japan*

10 *b. NOBIPOL, Department of Biotechnology and Food Science, NTNU - Norwegian*
11 *University of Science and Technology, Trondheim NO-7491, Norway*

12 *c. Department of Macromolecular Science, Osaka University, 1-1 Machikaneyama-*
13 *cho, Toyonaka, Osaka 560-0043, Japan*

14

15 Corresponding author

16 * Kazuto Yoshiba,

17 *Division of Molecular Science, Graduate School of Science and Technology, Gunma*
18 *University, 1-5-1 Tenjin-cho, Kiryu, Gunma, 376-8515, Japan*

19 1-5-1 Tenjin-cho, Kiryu, Gunma, 376-8515, JAPAN

20 Tel: +81 277 30 1486 Fax: +81 277 30 1409

21 e-mail: yoshiba@gunma-u.ac.jp

22

23 **Abstract**

24 Schizophyllan and scleroglucan are water-soluble polysaccharides having
25 repeating units consisting of three β -1,3-linked glucose residues in the main
26 chain and a single β -1,6-linked glucose residue as the side chain. This
27 polysaccharide dissolves as a triple helix in an aqueous solution and shows a
28 cooperative order-disorder transition between the side chain and solvent
29 molecules while retaining the triple helical conformation. Periodate and
30 subsequent chlorite oxidations selectively modify the side chain glucose to
31 provide the corresponding dicarboxylate units. Optical rotation measurements
32 and differential scanning calorimetry were performed on carboxylated
33 schizophyllan/scleroglucan (sclerox) samples to investigate the effects of the
34 degree of carboxylation on the order-disorder transition in deuterium oxide with
35 0.1 M NaCl. The transition curves for the sclerox samples are strongly dependent
36 on the degree of carboxylation. The modified side chains cannot take the ordered
37 structure, resulting in a reduction of the transition enthalpy. The transition
38 temperature for carboxylated schizophyllan becomes lowered and the transition
39 curve broadens with increasing the degree of carboxylation. The permanent
40 disordered units are included in a trimer by the carboxylation to inhibit a long
41 sequence of the ordered units.

42

43 **Keywords:** schizophyllan, periodate oxidation, order-disorder transition,
44 differential scanning calorimetry, optical rotation

45

46 **Introduction**

47 Schizophyllan is a water-soluble polysaccharide produced by *Schizophyllum*
48 *commune*, which consists of tetrasaccharide repeating units containing three β -
49 1,3-glucoses and one branched β -1,6-glucose (Kikumoto, Miyajima, Yoshizumi,

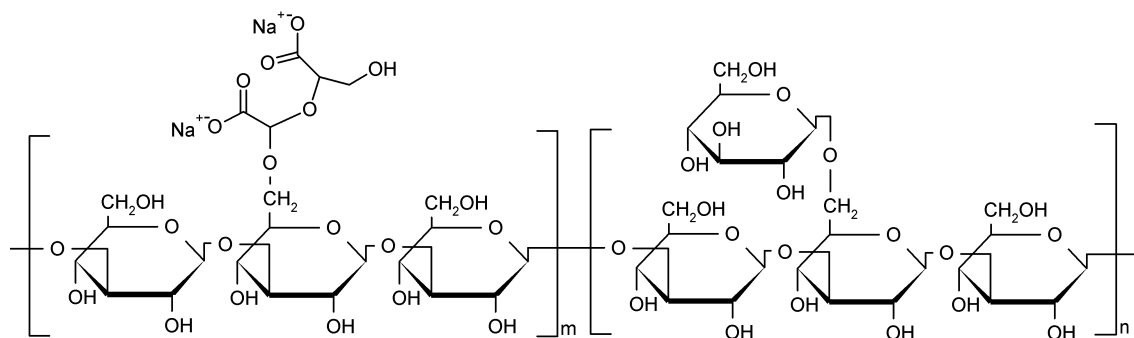
50 Fujimoto, Kimura, 1970; Kikumoto, Miyajima, Kimura, Okubo, Komatsu, 1971).
51 Scleroglucan is a structurally similar polysaccharide elaborated by a species of
52 the genus *Sclerotium* (Johnson et al, 1963). These polysaccharides dissolve in
53 water as triple helices, where three main chains form the triple helix and the side
54 chain glucoses are arranged outward the helical core to ensure high solubility to
55 water (Takahashi, Kobatake, Suzuki, 1984). The triple helix of schizophyllan and
56 scleroglucan shows strong molar mass dependencies of radius gyration
57 (Kashiwagi, Norisuye, Fujita, 1981; Yanaki & Norisuye, 1983) and intrinsic
58 viscosity (Norisuye, Yanaki, Fujita, 1980; Yanaki, Norisuye, Fujita, 1980) in
59 aqueous solutions. It was concluded on the analysis of the wormlike chain model
60 that the triple helix is a quite rigid rod represented with a large persistence
61 length above 150nm.

62 The schizophyllan triple helix has characteristic physical and biological
63 properties in the aqueous solutions, *e.g.*, high viscosity (Carriere, Amis, Schrag,
64 Ferry, 1985; Enomoto, Einaga, Teramoto, 1985; Fang & Nishinari, 2003), liquid
65 crystallinity (Van, Norisuye, Teramoto, 1980; Itou, Van, Teramoto, 1985),
66 anticancer activity (Norisuye, 1985; Okamura et al, 1986), and reassembly with
67 polynucleotides (Sakurai & Shinkai, 2000; Sakurai, Mizu, Shinkai, 2001;
68 Mochizuki & Sakurai, 2009). A cooperative order-disorder transition is one of the
69 characteristic features of the schizophyllan triple helix in aqueous solutions
70 (Asakawa, Van, Teramoto, 1984; Itou, Teramoto, Matsuo, Suga, 1986; Kitamura &
71 Kuge, 1989). The side chain forms an ordered structure along with hydrated
72 water molecules in the ordered triple helix state (Triple helix I). As a
73 temperature rises, the ordered structure is converted into the disordered triple
74 helix (Triple helix II). The transition has a strong molar mass dependency
75 because of its highly cooperative nature (Itou et al, 1986). In addition, the
76 transition temperature and the transition enthalpy depend largely on the solvent
77 conditions. The transition temperature shifts from 7°C in H₂O to 18°C in D₂O, and
78 the transition enthalpy of the D₂O solution is larger than in H₂O (Itou, Teramoto,
79 Matsuo, Suga, 1987). The same order-disorder transition was observed for other
80 branched β-1,3-glucans, and the number of branching per the main chain glucose
81 affects the order-disorder transitional behavior in DMSO-water mixtures
82 (Kitamura, Ozawa, Tokioka, Hara, Ukai, Kuge, 1990; Zhang, Xu, Zhang, 2008;
83 Wang, Zhang, Zhang, Ding, 2009), indicating that the hydration of the triple helix,
84 especially the side chains and solvent molecules, plays an important role in the
85 transition.

86 Chemical modifications generally alter the physicochemical and biological
87 properties of the parent polysaccharide. Periodate oxidation is well known as an
88 important reaction for the structural determination of polysaccharides (Perlin,
89 1980). This oxidation proceeds stoichiometrically with the glycol-cleavage
90 reaction at the C-C bond of the vicinal diols or successive hydroxyl groups.
91 Schizophyllan can be modified into ionic polysaccharide by periodate and
92 subsequently chlorite oxidations (Crescenzi, Gamini, Paradossi, Torri, 1983;
93 Gamini, Crescenzi, Abruzzese, 1984). The glucose ring of the side chain is
94 selectively modified into a linear dialdehyde chain, whereas the main chain of
95 schizophyllan is resistant to the periodate oxidation. Chlorite oxidation can
96 further alter the aldehyde groups into carboxyl groups. These oxidations allow
97 the preparation of different degrees of substitution (DS) of the carboxylated
98 derivative. Carboxylated schizophyllan (Sclerox, Figure 1) is soluble in water and

99 show different physical properties in aqueous solution from parent
100 schizophyllan, including polyelectrolyte properties (Coviello, Dentini, Crescenzi,
101 1995; Coviello, Dentini, Crescenzi, Vincenti, 1995), and related properties, such
102 as gelation with trivalent cations (Stokke, Elgsaeter, Smidrød, Christensen,
103 1995). The solubility to water is related to the hydration of this modified
104 polysaccharide, which may influence the order-disorder transition. However, the
105 effect of chemical modification of the side chains of schizophyllan has not been
106 investigated for the order-disorder transition.

107 In previous studies on the order-disorder transition of schizophyllan, the
108 molecular mechanism and the molecular motion were studied by optical
109 rotation, adiabatic calorimetry and dielectric relaxation (Teramoto, Gu, Miyazaki,
110 Sorai, Mashimo, 1995; Hayashi, Shinyashiki, Yagihara et al, 2001; Yoshiba, Ishino,
111 Teramoto Nakamura et al, 2002; Yoshiba, Teramoto, Nakamura, Kikuchi,
112 Miyazaki, Sorai, 2003; Yoshiba, Teramoto Nakamura et al, 2004). Chemical
113 modifications of polysaccharide side chains generally influence such transition
114 markers. For example, partial acid hydrolysis was used in a double helical
115 polysaccharide, xanthan, to investigate the influence of the removal of side
116 chains on the conformational transition of xanthan (Christensen, Knudsen,
117 Smidsrød, Kitamura, Takeo, 1993). Since the order-disorder transition of
118 schizophyllan is characteristic to the triple helical conformation (Kitamura &
119 Kuge, 1989), it is necessary to investigate both of the molecular conformation
120 and the transition behavior after the chemical modification. From
121 conformational aspects for sclerox, we showed that a sclerox at low DS remains a
122 trimer in 0.1 M NaCl (Yoshiba, Sato, Osumi, Ulset, Christensen, 2015). However,
123 size-exclusion chromatography combined with multi-angle light scattering
124 detection (SEC-MALS) indicated the global conformation of the trimer is different
125 from the triple helix of parent schizophyllan by becoming more flexible and
126 extended after the carboxylation. In the present study, we report effects of
127 carboxylation of the side chains on the cooperative order-disorder transition of
128 sclerox. The sclerox samples with different DS were investigated by differential
129 scanning calorimetry and optical rotation to characterize the transition behavior
130 of sclerox. **Since schizophyllan in aqueous solutions has a low transition**
131 **temperature of 7 °C, D₂O was used as solvent to extend the temperature range of**
132 **the triple helix I.** We added 0.1 M NaCl to essentially prevent the electrostatic
133 interaction between the trimers. Since the conformation of the sclerox trimer can
134 be determined from SEC-MALS measurements, the effects of chemical
135 modification on its specific hydration around the triple helix were analyzed.
136



138 **Figure 1** Chemical structure of the carboxylated schizophyllan/scleroglucan
139 (Sclerox) by partial periodate and chlorite oxidation.

140

141

142 **Experimental**

143

144 *Schizophyllan and carboxylated schizophyllan samples*

145 The schizophyllan sample (Taito Co., now Mitsui sugar Co., Japan) was sonicated
146 in an aqueous solution with a Branson Sonifier 250 (Branson Ultrasonics Co.,
147 Danbury, USA) to prepare two degraded schizophyllan samples. The sonicated
148 sample was purified by the fractional reprecipitation in the aqueous solution
149 with ethanol to obtain middle fraction of each sample (SPG-1, SPG-2). The
150 carboxylation of the sample was performed in an aqueous solution according to
151 previous study on sclerox (Yoshiba et al, 2015). In brief, 30 mM aqueous sodium
152 periodate (Wako Pure Chemical Industries Ltd., Osaka, Japan) was dropped into
153 120 ml of 0.1wt% aqueous schizophyllan containing 13.3 ml of 1-propanol to
154 prepare different degree of oxidation of the samples. After dialysis against
155 deionized water, sodium chlorite (Kanto Chemical Co. Inc., Tokyo, Japan) and
156 acetic acid (Wako Pure Chemical Industries Ltd., Osaka, Japan) were added into
157 the solution to be 0.5 M for each reagent. These oxidations were performed in
158 dark at 10 °C for 24 hours. After oxidations, the solutions were dialyzed and
159 neutralized with 0.01 M NaOH to obtain the sodium salt of the freeze-dried
160 samples. The sclerox samples were coded as SC-1-*X* or SC-2-*X*, where *X* is
161 denoted by the mole fraction of sodium periodate for the repeating unit of
162 schizophyllan at the oxidation. The degree of substitution, DS, was determined
163 by the potentiometric titration for the acid form of the sample with 0.01 M NaOH
164 in aqueous 0.1 M NaCl after the immersion with Amberlite-120B (Organo Co.,
165 Ltd., Tokyo, Japan) for 24 hours. The degree of substitution of the samples was
166 listed in Table 1.

167

168 *Size Exclusion Chromatography equipped with a Multi-Angle Light Scattering* 169 *detector (SEC-MALS)*

170 The weight average molar mass and the molar mass dispersity of the samples
171 were determined by the size exclusion chromatography equipped with a multi-
172 angle light scattering detector (SEC-MALS). The SEC columns (SHODEX OHPak
173 SB-G, two SHODEX OHPak SB806MHQ, Showa Denko KK, Kanagawa, Japan) were
174 serially connected in the SEC system (GPC-101, Showa Denko KK, Kanagawa,
175 Japan). The SEC system was connected with a DAWN HELEOS II MALS detector
176 (Wyatt Technology Co., Santa Barbara, USA) and differential refractometer.
177 Aqueous 0.1 M NaCl was used as an eluent. The flow rate was fixed at 1.0
178 cm³min⁻¹ and the columns were kept at 40 °C in the measurement. The solution
179 dissolved in 0.1 M NaCl with 0.01 M NaOH was injected (injection volume, 0.1
180 cm³) after the filtration with DISMIC-25AS (pore size 0.45 μm, Toyo Roshi Co.,
181 Ltd., Tokyo, Japan) and the measurement was operated by ASTRA® ver. 5.3
182 software (Wyatt Technology Co., Santa Barbara, USA). The refractive index
183 increment, $(\partial n/\partial c)_\mu=0.141$ cm³g⁻¹, was used to calculate the weight average
184 molar mass, M_w , and molar mass dispersity M_w/M_n . Table 1 summarizes M_w , and
185 M_w/M_n of the samples used in following measurements determined from the
186 SEC-MALS measurements.

187
188
189

Table 1 The degree of substitution and SEC-MALS results for carboxylated schizophyllan and schizophyllan samples in aqueous 0.1 M NaCl.

Sample	DS	$M_w / \text{g mol}^{-1}$	M_w/M_n
SPG-1	0	34.1×10^4	1.4
SC-1-0.2	0.16	32.4×10^4	1.4
SC-1-0.4	0.20	32.4×10^4	1.4
SPG-2	0	16.0×10^4	1.4
SC-2-0.4	0.21	13.2×10^4	1.4

190

191 *Optical rotation (OR) measurement*

192 Optical rotation measurements were performed with a P-1000 digital
193 polarimeter (JASCO Co., Tokyo, Japan) at the wavelength of 589 nm. 1 wt%
194 solution was prepared with D₂O (Sigma-Aldrich Co., USA) with 0.1 M NaCl. The
195 solution was filled in a cylindrical cell of the optical pass length of 5.0 cm with a
196 water jacket. The cell was thermo-stated by circulating water and the
197 temperature was recorded by thermocouples attached on the cell wall. The mass
198 concentration was calculated with the solution density to determine the specific
199 rotation, $[\alpha]_{589}$.

200

201 *Differential scanning calorimetry (DSC)*

202 Differential scanning calorimetry for schizophyllan and carboxylated
203 schizophyllan solutions was made on a Pyris-1 differential scanning calorimeter
204 (Perkin Elmer Co., Massachusetts, USA). D₂O containing 0.1 M NaCl were injected
205 into dried sample in a large volume stainless steel pan (Perkin Elmer Co.,
206 Massachusetts, USA). The measurements were performed at the scanning rate of
207 2.0 K min⁻¹ with the solvent as a reference at nitrogen gas flow of 10 cm³ min⁻¹.
208 The temperature scale of the calorimeter was calibrated with the melting
209 temperature of cyclohexane and indium and the heat flow was corrected with
210 the enthalpy of fusion of indium. The measurement was operated by a Pyris®
211 software (Perkin Elmer Co., Massachusetts, USA). The measurements were
212 performed repeatedly to confirm the heat flow curves to reproduce each
213 measurement within the experimental error.

214

215

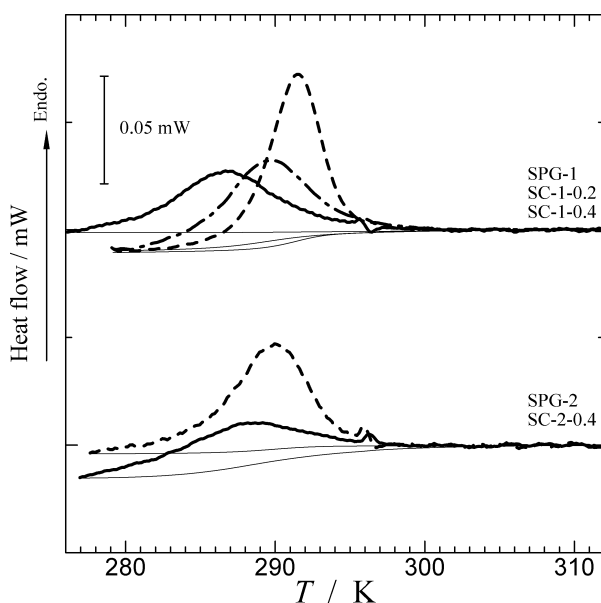
216 **RESULTS and DISCUSSION**

217

218 *DSC curves and optical rotation*

219 Figure 2 shows DSC curves for 10 wt% solutions of SPG-1, SC-1-0.2, SC-1-0.4
220 and SPG-2, SC-2-0.4, respectively, in D₂O containing 0.1 M NaCl. The heating rate
221 was 2.0 K min⁻¹. The DSC curves were reproduced by repeated heating/cooling
222 cycles. Hence, the transition is thermally reversible in this temperature range. All
223 DSC curves show a thermal peak corresponding to the order-disorder transition.
224 The transition temperature, T_r , was taken from the peak of the DSC curve (Table
225 2). By comparing SPG-1 and SPG-2 it is observed that both T_r and the sharpness
226 of the peak increase with increasing M , which reflect the molar mass dependence
227 of the order-disorder transition (Itou et al, 1987; Yoshiba et al, 2002). In
228 contrast, the DSC curves for the carboxylated SC-1-0.2, SC-1-0.4 and SC-2-0.4
229 have broader peaks and are shifted to lower temperatures. The transition

230 enthalpies, ΔH_r , were calculated from the peak areas of the DSC curves in Figure
 231 2, where the thin lines indicated were chosen as baselines for the samples (Table
 232 2). Compared with previous study of schizophyllan, the present ΔH_r is slightly
 233 underestimated by a tailing at low temperature side. Nonetheless, the values of
 234 ΔH_r for the sclerox solutions were appreciably smaller than those for the parent
 235 schizophyllan, indicating that the carboxylated side chains do not contribute to
 236 the heat of the order-disorder transition.
 237
 238



239
 240 **Figure 2** DSC curves for 10 wt% solutions of schizophyllan and carboxylated
 241 schizophyllan in D₂O containing 0.1 M NaCl at the heating rate of 2.0 K min⁻¹.
 242 Dashed curves: SPG-1 and SPG-2. Dash-dotted curve: SC-1-0.2. Solid curves: SC-
 243 1-0.4 and SC-2-0.4. The data were vertically shifted for clarity.

244
 245
 246 **Table 2** The transition enthalpy and transition temperature obtained from DSC
 247 and OR (T_r only)

Sample	ΔH_r / kJmol ⁻¹	T_r / K (DSC)	T_r / K (OR) ^a
SPG-1	3.9	291.3	291.0
SC-1-0.2	3.0	289.3	288.8
SC-1-0.4	2.7	286.5	287.1
SPG-2	3.4	289.8	288.3
SC-2-0.4	2.3	288.0	286.0

248 a, at $f=0.5$

249

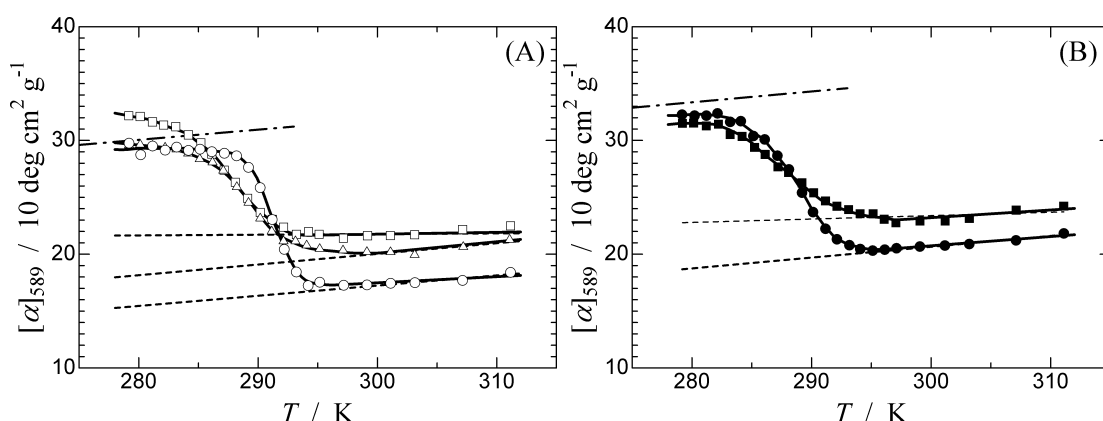
250

251 Figure 3A and 3B show the temperature dependence of the specific rotation
 252 $[\alpha]_{589}$ for the five samples. The wavelength was 589 nm, and the solvent was the
 253 same as in the DSC experiments (D₂O with 0.1 M NaCl). Generally, all samples
 254 displayed a typical transition (decrease in specific optical rotation) in the same
 255 temperature range as in the DSC experiments. The linear part of the curves on
 256 the high temperature side (dashed lines) was taken as the baseline $[\alpha]_{\text{dis}}$ for the
 257 disordered state of each sample. Baselines $[\alpha]_{\text{ord,SPG}}$ for the ordered states of SPG-

258 1 and SPG-2, were drawn parallel to those of the disordered states (dash-dotted
 259 lines).

260

261



262

263 **Figure 3** Specific rotations $[\alpha]_{589}$ at the wavelength of 589 nm for schizophyllan
 264 and carboxylated schizophyllan in 0.1 M NaCl D₂O solutions: (A), unfilled circles,
 265 SPG-1, unfilled triangles, SC-1-0.2, unfilled squares, SC-1-0.4; (B), filled circles,
 266 SPG-2, filled squares, SC-2-0.4. Dashed lines are the baselines for the disordered
 267 state, $[\alpha]_{\text{dis}}$, the dash-dotted lines are the baselines for the ordered state, $[\alpha]_{\text{ord}}$,
 268 for SPG-1 and SPG-2, and solid curves are eye guides.

269

270

271 It is first noted that the optical rotation in the disordered state shifts to higher
 272 values with increasing DS. This suggests that the intact side chain (glucose
 273 residue) and the modified side chain (Figure 1) contribute differently to the total
 274 optical rotation, reflecting differences in the number of asymmetric carbons in
 275 the two cases (Jackson & Hudson, 1937). This finding parallels the observations
 276 for xanthan with partially hydrolysed (removed) side chains (Christensen,
 277 Knudsen, Smidsrød, Kitamura, Takeo, 1993), where the total optical rotation
 278 (and transition enthalpy) changed linearly with changes in the number of side
 279 chains. Secondly, it is noted that the transition temperatures obtained from OR
 280 agree well with those obtained by DSC, i.e. a decrease with increasing DS (Table
 281 2). Thirdly, the transitions detected by OR become broader with increasing DS,
 282 again in qualitative agreement with DSC. Finally, the unmodified SPG-2 appears
 283 to have a broader transition curve than that of SPG-1. This is attributed to the *M*
 284 dependence of the transition, as mentioned above.

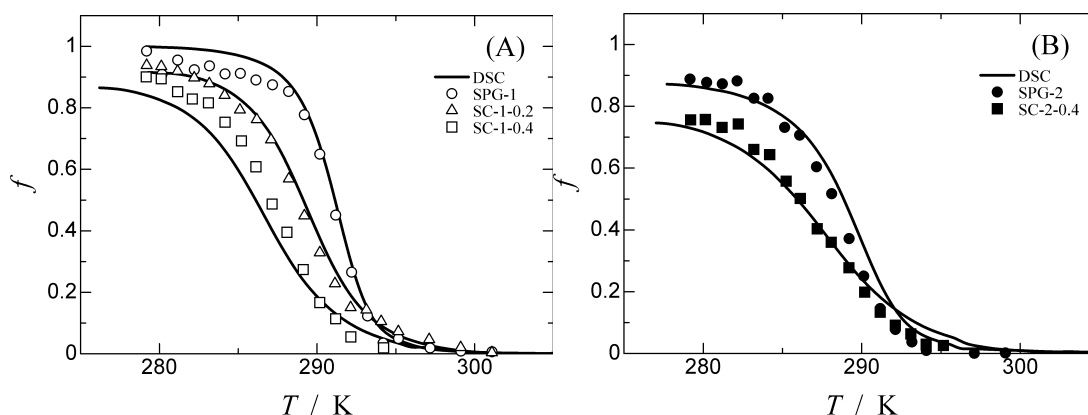
285 The fraction *f* of repeating units in the ordered state was calculated by

$$286 \quad f = \frac{1}{1-DS} \left(\frac{[\alpha]_{589} - [\alpha]_{\text{dis}}}{[\alpha]_{\text{ord,SPG}} - [\alpha]_{\text{dis,SPG}}} \right) \quad (1)$$

287 where $[\alpha]_{\text{dis,SPG}}$ is the baseline for the disordered state of SPG. The midpoint of
 288 the transition was taken as the temperature at $f = 0.5$. The values are included in
 289 Table 2. Figure 4A and 4B show the comparison of *f* between the DSC and the OR
 290 measurements for SPG-1, SC-1-0.2, SC-1-0.4, and SPG-2, SC-2-0.4 to confirm the
 291 consistency between DSC and OR data. The data from the optical rotation were
 292 indicated with same symbols of Figure 3A and 3B. The fraction *f* of repeating
 293 units in the ordered state can be calculated from DSC curves by

294
$$f = \int_{T_i}^T \Delta h dT / (1-DS) \int_{T_i}^{T_f} (\Delta h)_{\text{SPG}} dT \quad (2)$$

295 where Δh is the difference between the DSC curve and the baseline, T_i and T_f are
 296 the temperatures where the DSC curve reaches to the baseline at low and high
 297 temperature sides, respectively, and DS is the degree of substitution. In Figure 4A
 298 and 4B, solid curves indicate the results of f for SPG-1, SC-1-0.2, SC-1-0.4 and
 299 SPG-2, SC-2-0.4 obtained by DSC. Both data agreed satisfactorily in this
 300 experimental temperature range.
 301



302
 303 **Figure 4** Comparison of the fraction of the repeating unit in the ordered state
 304 between DSC and OR measurements; the data points, the values from the specific
 305 rotation $[\alpha]_{589}$; (A), SPG-1, SC-1-0.2 and SC-1-0.4; (B) SPG-2 and SC-2-0.4. Solid
 306 curves are the calculated values from DSC curves.
 307

308 *Analysis of the transition curve*

309 We can summarize present results of SEC-MALS, DSC, and optical rotation (OR)
 310 measurements performed on the order-disorder transition of sclerox, as follows:
 311 Firstly, the sclerox samples have almost the same M before and after the
 312 oxidations. In contrast, a dissociation or aggregation of the trimer would largely
 313 affect M_w and the molar mass distribution (Yoshida et al, 2015). However, these
 314 are almost unchanged for present samples by the chemical modification, which is
 315 attributed to the low DS. This indicates that the sclerox samples maintain the
 316 trimeric state. The second is the dependencies of ΔH_f and OR data on DS. The
 317 modified side chains are most probably randomly distributed along the sclerox
 318 trimer. Since only the unmodified side chains contribute to form the ordered
 319 structure, the reduction of ΔH_f with increasing DS suggests that the modified side
 320 chain cannot take the ordered structure. Thirdly, T_f becomes lower, and the
 321 transition curve broadens with increasing DS. These effects are concerned with
 322 the molecular mechanism of the order-disorder transition. As demonstrated
 323 previously, the order-disorder transition occurring in the schizophyllan triple
 324 helix is described by a statistical theory of one-dimensional cooperative systems
 325 (Itou et al, 1986; Itou et al, 1987; Yoshida et al, 2002). Therefore, we discuss the
 326 above-mentioned effects of the carboxylation on the transition using the same
 327 statistical theory in what follows.
 328

329 As pointed out by Itou et al (1986), this transition depends strongly on the
 330 sample molar mass, indicating that the transition can be regarded as a linear
 331 cooperative phenomenon. It has been shown that such transition can be

332 characterized by the statistical theory for the helix-coil transition established by
 333 Zimm, Bragg (Zimm & Bragg, 1959) and Nagai (Nagai, 1960, 1961). The
 334 schizophyllan triple helix is divided into N transition units. In present analysis,
 335 the unit is taken to be three times of the repeating units of the chemical
 336 structure, i.e. $3M_0$ for schizophyllan ($M_0 = 648.6 \text{ g mol}^{-1}$). At the low temperature
 337 side of the transition, the units except for the terminals are in the ordered state
 338 (the fraction of the ordered units, $f \sim 1$). The sequence of the ordered units is
 339 shortened with a progress of the transition, and finally all of the units become
 340 the disordered state ($f = 0$). The statistical weight of the ordered unit, s , is related
 341 to the Gibbs energy difference, ΔG_r^∞ by

$$342 \quad \ln s = \frac{3\Delta G_r^\infty}{RT} = \frac{3\Delta H_r^\infty}{RT} \left(1 - \frac{T}{T_r^\infty}\right) \quad (3)$$

343 where R is the gas constant and T is the absolute temperature. ΔH_r^∞ and T_r^∞ are
 344 the transition enthalpy and the transition temperature at infinite N , respectively.
 345 Assuming that the transition is concerned only between the adjacent units, the
 346 partition function for the triple helix is given by

$$347 \quad Z_N = (0 \quad 1)\mathbf{M}^{N-2} \begin{pmatrix} 1 \\ 1 \end{pmatrix} \quad (4)$$

348 where \mathbf{M} is the matrix of 2×2 defined by

$$349 \quad \mathbf{M} = \begin{pmatrix} s & 1 \\ \sigma^{1/2}s & 1 \end{pmatrix} \quad (5)$$

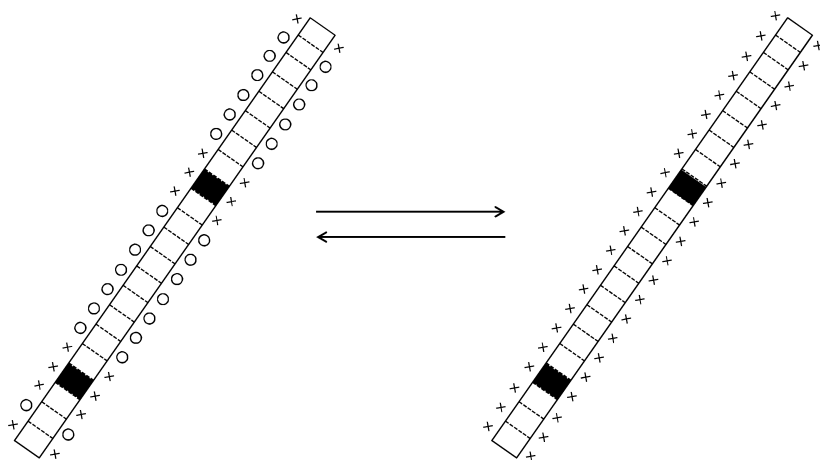
350 with the cooperative parameter $\sigma^{1/2}$, characterizing the sequence number of the
 351 ordered units at the half of the transition at infinite N (Teramoto & Fujita, 1975).
 352 The fraction of the ordered units in the trimer is expressed formally by the
 353 relation,

$$354 \quad f = \frac{1}{N} \frac{d \ln Z_N}{d \ln s} \quad (6)$$

355 Teramoto (2001) summarized the formulas to calculate the numerical value of
 356 the transition curve and the physical meanings of the theoretical parameters
 357 concerned.

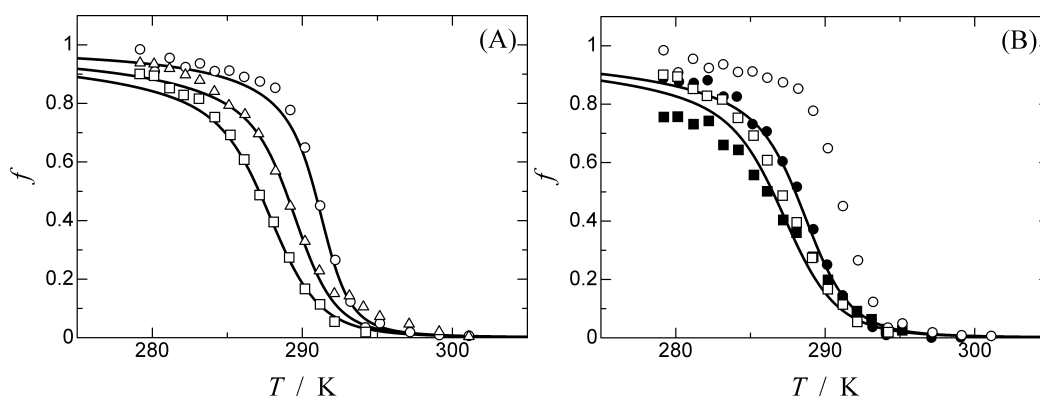
358 For the sclerox trimer, some of the side chains should be replaced by the
 359 modified side chains. The transition unit in the trimer consists of three side
 360 chains, which can take either the ordered or disordered structure. The periodate
 361 oxidation most likely modify the side chains randomly, so that three types of
 362 modified units are produced after the chemical modification, in addition to the
 363 unmodified unit. If all of the side chains in the unit are modified, the unit takes
 364 only the disordered state. Here, this unit is termed the permanent disordered
 365 unit. Since this unit in the trimer interrupts the formation of long sequence of the
 366 ordered units as a terminal, the sequence of the ordered units becomes shorten
 367 even at satisfactorily low temperature. Consequently, the transition curve may
 368 become broader like a shorter chain. This situation is illustrated in Figure 5. In
 369 this figure, the filled units and unfilled units represent the inactive units and the
 370 active units to the order-disorder transition, respectively. The inactive unit has
 371 three modified side chains, that is, the permanent disordered unit, whereas the
 372 others belong to the active units. In this consideration, N can be regarded as the
 373 successive number of the active units for the transition. Using theoretical
 374 parameters (ΔH_r^∞ , T_r^∞ , $\sigma^{1/2}$) for D_2O solution of schizophyllan, the transition

375 curve for sclerox solutions can be fitted by eq. (6) with N as an adjustable
 376 parameter.
 377



378
 379 **Figure 5** Schematic illustrations for the order-disorder transition for
 380 carboxylated schizophyllan. The polysaccharide units in the ordered and
 381 disordered states designate circles and crosses at the side of the units,
 382 respectively. Filled units represent the permanent disordered units.
 383
 384

385 Figure 6A shows the fitting results for SC-1-0.2 and SC-1-0.4 by using eq. (6).
 386 For the calculation of the theoretical curve, known parameters were used; $\Delta H_r^\infty =$
 387 4600 J mol^{-1} , $T_r^\infty = 292.3 \text{ K}$, $\sigma^{1/2} = 0.0135$ (Itou et al, 1987). For SPG-1, the number
 388 average molar mass was used for the theoretical calculation; $N=125$. The
 389 theoretical curve well represents the f data for SPG-1. The results for SC-1-0.2
 390 and SC-1-0.4 can be fitted with $N=69$ and $N=51$, respectively. It is noted that N
 391 for SC-1-0.2 becomes nearly half the parent sample. This means that the
 392 sequence of the ordered units of SC-1-0.2 is divided into two parts by including
 393 the permanent disordered unit in the trimer. N for SC-1-0.4 becomes smaller
 394 than SC-1-0.2. Since the number of the permanent disordered unit in the trimer
 395 is further produced with increasing DS, the sequence of the ordered units
 396 becomes shorter than SC-1-0.2.
 397
 398



399
 400 **Figure 6** Comparison between theoretical curves and experimental data from
 401 the specific rotation for the order-disorder transition of sclerox in D_2O
 402 containing 0.1 M NaCl : (A) unfilled circles, SPG-1 ($N=125$); unfilled triangles, SC-
 403 1-0.2 ($N=69$); unfilled squares, SC-1-0.4 ($N=51$); (B) filled circles, SPG-2 ($N=59$);

404 filled squares, SC-2-0.4 ($N=47$); unfilled circles and unfilled squares, same as
405 those in Panel A. The solid curves were calculated with $\Delta H_r^\infty = 4600 \text{ Jmol}^{-1}$, T_r^∞
406 $= 292.3 \text{ K}$, $\sigma^{1/2} = 0.0135$.

407
408

409 Figure 6B shows the f data for SPG-2 and SC-2-0.4. The f data for SPG-1 and SC-
410 1-0.4 are also shown in Figure 6(B) to compare the DS and M dependencies. It
411 can be seen that the strong M dependence between SPG-1 and SPG2 is well
412 expressed by the theoretical curve with $N=59$ for SPG-2 without fitting
413 parameter. The data for SC-2-0.4 can be fitted by $N=47$, which is comparable to
414 SC-1-0.4. Although the carboxylated side chains of SC-1-0.4 and SC-2-0.4 are
415 distributed with almost equimolar ratio against total number of the side chains,
416 the number of carboxylated side chain per a trimer becomes smaller for SC-2-0.4
417 because of low M . Therefore, the permanent disordered unit in a trimer of SC-2-
418 0.4 is less produced compared with SC-1-0.4. Consequently, the transition curve
419 becomes insensitive to M after the carboxylation.

420
421

422 Conclusion

423 Carboxylated schizophyllan ('sclerox') synthesized by partial periodate oxidation
424 followed by chlorite oxidation dissolves as a trimer, and shows a cooperative
425 order-disorder transition in D_2O solution containing 0.1 M NaCl, analogous to the
426 parent schizophyllan. The transition enthalpy decreases and the transition
427 temperature is lowered with increasing degree of carboxylation. The
428 carboxylated side chains of the trimer introduced by oxidation cannot take the
429 ordered structure, leading the decreasing the enthalpy of the transition.

430 Adopting the linear cooperative theory for the order disorder transition of
431 schizophyllan solution, the length of the ordered unit for sclerox becomes
432 shorter than those of parent schizophyllan, where the disordered unit produced
433 at carboxylation acts as a terminal of the successive ordered sequence.

434 Consequently, the transition temperature and transition enthalpy for sclerox
435 solution depend largely on the degree of carboxylation.

436
437

438 References

- 439 Asakawa, T., Van, K., Teramoto, A. (1984). A thermal transition in a cholesteric
440 liquid crystal of aqueous schizophyllan. *Molecular Crystals and Liquid Crystals*,
441 116, 129-139.
- 442 Carriere, C. J., Amis, E. J., Schrag, J. L., Ferry, J. D. (1985). Dilute-solution dynamic
443 viscoelastic properties of schizophyllan polysaccharide, *Macromolecules*, 18,
444 2019-2023.
- 445 Christensen, B. E., Knudsen, K. D., Smidsrød, O., Kitamura, S., Takeo, K. (1993).
446 Temperature-induced conformational transition in xanthans with partially
447 hydrolyzed side chains. *Biopolymers*, 33, 151-161.
- 448 Coviello, T., Dentini, M., & Crescenzi, V. (1995). Conformation and thermal
449 stability of oxidized scleroglucan chains in aqueous NaOH. *Polymer Bulletin*, 34,
450 337-343.

451 Coviello, T., Dentini, M., Crescenzi, V., & Vincenti, A. (1995). Ionic strength and
452 temperature dependence of oxidized scleroglucan solution properties: optical
453 activity and viscosity data. *Carbohydrate Polymers*, 26,5-10.

454 Crescenzi, V., Gamini, A., Paradossi, G., & Torri, G (1983). Solution properties of a
455 new polyelectrolyte derived from the polysaccharide scleroglucan. *Carbohydrate*
456 *Polymers*, 3, 273-286.

457 Enomoto, H., Einaga, Y., Teramoto, A. (1985). Viscosity of concentrated solutions
458 of rodlike polymers, *Macromolecules*, 18, 2695-2702.

459 Fang, Y., Nishinari, K. (2003) Gelation behaviors of schizophyllan – sorbitol
460 Aqueous solutions, *Biopolymers*, 73, 44-60.

461 Gamini, A., Crescendi, V., Abruzzese, R. (1984). Influence of the charge density on
462 the solution behaviour of polycarboxylates derived from the polysaccharide
463 scleroglucan. *Carbohydrate Polymers*, 4, 461-472.

464 Hayashi, Y., Shinyashiki, N., Yagihara, S., Yoshiba, K., Teramoto, A., Nakamura, N.,
465 Miyazaki, Y., Sorai, M., Wang, Q. (2002). Ordering in aqueous polysaccharide
466 solutions. I. Dielectric relaxation in aqueous solution of a triple-helical
467 polysaccharide schizophyllan. *Biopolymers*, 63, 21-31.

468 Itou, T., Teramoto, A., Matsuo, T., Suga, H. (1986). Ordered structure in aqueous
469 polysaccharide. 5. Cooperative order-disorder transition in aqueous
470 schizophyllan. *Macromolecules*, 19, 1234-1240.

471 Itou, T., Teramoto, A., Matsuo, T., Suga, H. (1987). Isotope effect on the order-
472 disorder transition in aqueous schizophyllan. *Carbohydrate research*, 160, 243-
473 257.

474 Itou, T., Van, K., Teramoto, A. (1985). Molecular weight dependence of the
475 isotropic-cholesteric phase diagram for the system schizophyllan + water,
476 *Journal Applied Polymer Science Applied Polymer Symposia*, 41, 35-48.

477 Jackson, E. L. , Hudson, C. S. (1937). Studies on the cleavage of the carbon chain of
478 glycoside by oxidation. A new method for determining and alfa and beta
479 configurations of glycosides. *Journal of American Chemical Society*, 59, 994-1003.

480 Johnson, J., Kirkwood, S., Misaki, A., Nelson, T. E., Scletti, J. V., Smith, F. (1963).
481 Structure of a new glucan. *Chemistry and Industry*, 820-822.

482 Kashiwagi, Y., Norisuye, T., & Fujita, H. (1981). Triple helix of Schizophyllum
483 commune polysaccharide in dilute solution. 4. Light scattering and viscosity in
484 dilute aqueous sodium hydroxide. *Macromolecules*, 14, 1220-1225.

485 Kikumoto, S., Miyajima, T., Yoshizumi, S., Fujimoto, S., & Kimura, T. (1970).
486 Polysaccharide produced by Schizophyllum commune Part I. Formation and
487 some properties of an extracellular polysaccharide. *Journal of the Agricultural*
488 *Chemical Society of Japan*, 44, 337-342.

489 Kikumoto, S., Miyajima, T., Kimura, K., Okubo, S., & Komatsu, N. (1971).
490 Polysaccharide produced by Schizophyllum commune Part II. Chemical structure
491 of an extracellular polysaccharide. *Journal of the Agricultural Chemical Society of*
492 *Japan*, 45, 162-168.

493 Kitamura, S., & Kuge, T. (1989). A differential scanning calorimetric study of the
494 conformational transitions of schizophyllan in mixtures of water and
495 dimethylsulfoxide *Biopolymers*, 28, 639-654.

496 Kitamura, S., Ozawa, M., Tokioka, H., Hara, C., Ukai, S., Kuge, T. (1990). A
497 differential scanning calorimetric study of the conformational transition of
498 several kinds of (1→6) branched (1→6)-β-D-glucans in a mixture of water and
499 dimethylsulfoxide. *Thermochimica Acta*, 163, 89-96.

500 Mochizuki, S., Sakurai, K. (2009) A noble polysaccharide/polynucleotide complex
501 and its application to bio-functional DNA delivery system. *Polymer Journal*, 41,
502 343-353.

503 Nagai, K. (1960). Configurational change of polypeptide molecules in the helix-
504 coil transition region, I. *Journal of Physical Society of Japan*, 15, 407-416.

505 Nagai, K. (1961). Dimensional change of polypeptide molecules in the helix-coil
506 transition region. II. *The Journal of Chemical Physics*, 34, 887-904.

507 Norisuye, T. (1985). Triple-stranded helical structure of schizophyllan and its
508 antitumor activity in aqueous solution. *Die Makromolekulare Chemie Supplement*,
509 14, 105-118.

510 Norisuye, T., Yanaki, T., & Fujita, H. (1980). Triple helix of a Schizophyllum
511 commune polysaccharide in aqueous solution. *Journal of Polymer Science:*
512 *Polymer Physics Edition*, 18, 547-558.

513 Okamura, K., Suzuki, M., Chihara, T., Fujiwara, A., Fukuda, T., Goto, S., Ichinohe, K.,
514 Jimi, S., Kasamatsu, T., Kawai, N., Mizuguchi, K., Mori, S., Nakano, H., Noda, K.,
515 Sekiba, K., Suzuki, K., Suzuki, T., Takahashi, K., Takeuchi, K., Takeuchi, S., Yajima,
516 A., & Ogawa, N. (1986). Clinical evaluation of schizophyllan combined with
517 irradiation in patients with cervical cancer. *Cancer*, 58, 865-872.

518 Perlin, A. S. (1980). Glycol-Cleavage Oxidation. In Pigman, W., Horton, D. Wander,
519 J. D. Eds. "The Carbohydrates. Chemistry and Biochemistry 2 ed. Vol. IIB", New
520 York, Academic Press.

521 Sakurai, K. & Shinkai, S. (2000) Molecular recognition of adenine, cytosine, and
522 uracil in a single-stranded RNA by a natural polysaccharide: schizophyllan.
523 *Journal of American Chemical Society*, 122, 4520-4521.

524 Sakurai, K., Mizu, M., Shinkai, S. (2001) Polysaccharide-polynucleotide
525 complexes. 2. complementary polynucleotide mimic behavior of the natural
526 polysaccharide schizophyllan in the macromolecular complex with single-
527 stranded RNA and DNA. *Biomacromolecules*, 2, 641-650.

528 Stokke, B. T., Elgsaeter, A., Smidrød, O., & Christensen, B. E. (1995). Carboxylation
529 of scleroglucan for controlled crosslinking by heavy metal ions. *Carbohydrate*
530 *Polymers*, 27, 5-11.

531 Takahashi, Y., Kobatake, T., & Suzuki, H. (1984). Triple helical structure of
532 schizophyllan. *Reports on Progress in Polymer Physics in Japan*, 27, 767-768.

533 Teramoto, A. (2001). Cooperative conformational transitions in linear
534 macromolecules undergoing chiral perturbation. *Progress in Polymer Science*, 26,
535 667-720.

536 Teramoto, A., & Fujita, H. (1975). Conformation-dependent properties of
537 synthetic polypeptide in the helix-coil transition region. *Advances in Polymer*
538 *Science*, 18, 65-149

539 Teramoto, A., Gu, H., Miyazaki, Y., Sorai, M., Mashomo, S. (1995). Dielectric study
540 of the cooperative order-disorder transition in aqueous solutions of
541 schizophyllan, a triple-helical polysaccharide. *Biopolymers*, 36, 803-810.

542 Yanaki, T. & Norisuye, T. (1983). Triple helix and random coil of scleroglucan in
543 dilute solution. *Polymer Journal*, 15, 389-396.

544 Yanaki, T., Norisuye, T., Fujita, H. (1980). Triple helix of Schizophyllum commune
545 polysaccharide in dilute solution. 3. Hydrodynamic properties in water.
546 *Macromolecules*, 13, 1462-1466.

547 Yoshiba, K., Ishino, T., Teramoto, A., Nakamura, N., Miyazaki, Y., Sorai, M., Wang,
548 Q., Hayashi, Y., Shinyashiki, N., Yagihara, S. (2002). Ordering in aqueous

549 polysaccharide solutions. II. Optical rotation and heat capacity of aqueous
550 solutions of a triple-helical polysaccharide schizophyllan, *Biopolymers*, 63, 370-
551 381.

552 Yoshiba, K., Sato, T., Osumi, T., Ulset, A.-S. T., Christensen, B. E. (2015).
553 Conformation of carboxylated schizophyllan in aqueous solution. *Carbohydrate*
554 *Polymers*, 134, 1-5.

555 Yoshiba, K., Teramoto, A., Nakamura, N., Kikuchi, K., Miyazaki, Y., Sorai, M.
556 (2003). Static water structure detected by heat capacity measurements on
557 aqueous solution of a triple-helical polysaccharide schizophyllan.
558 *Biomacromolecules*, 4, 1348-1356.

559 Yoshiba, K., Teramoto, A., Nakamura, Shikata, T., K., Miyazaki, Y., Sorai, M.,
560 Hayashi, Y., Miura, N. (2004). Water structure of differing order and mobility in
561 aqueous solution of schizophyllan, a triple-helical polysaccharide as revealed by
562 dielectric dispersion measurements. *Biomacromolecules*, 5, 2137-2146.

563 Van, K., Norisuye, T.; Teramoto, A. (1981) Liquid crystal formation in aqueous
564 solutions of a polysaccharide schizophyllan, *Molecular Crystals and Liquid*
565 *Crystals*, 78, 123-134.

566 Wang, X., Zhang, Y., Zhang, L., Ding, Y. (2009). Multiple conformation transition of
567 triple helical lentinan in DMSO/water by Microcalorimetry. *Journal of Physical*
568 *Chemistry B*, 113, 9915-9923.

569 Zhang, Y., Xu, X., Zhang, L. (2008). Gel formation and low-temperature
570 intramolecular conformation transition of a triple-helical polysaccharide
571 lentinan in water. *Biopolymers*, 89, 852-861.

572 Zimm, B. H., Bragg, J. K. (1959). Theory of phase transition between helix and
573 random coil in polypeptide chains. *The Journal of Chemical Physics*, 31, 526-535.
574