

The biology hidden inside residual within-individual phenotypic variation

David F. Westneat^{1,*}, Jonathan Wright² and Niels J. Dingemanse^{3,4}

¹*Department of Biology and Center for Ecology, Evolution, and Behavior, 101 Morgan Building, University of Kentucky, Lexington, KY 40506-0225, USA*

²*Center for Biodiversity Dynamics, Department of Biology, Norwegian University of Science and Technology (NTNU), N-7491 Trondheim, Norway*

³*Behavioural Ecology, Department of Biology, Ludwig-Maximilians University of Munich, Planegg-Martinsried, Germany*

⁴*Evolutionary Ecology of Variation Research Group, Max Planck Institute for Ornithology, Seewiesen, Germany*

Running Title: The biology of residual phenotypic variance

* Author for correspondence (Tel.: 859-323-9499; Fax: 859-257-1717; E-mail: biodfw@uky.edu).

1 ABSTRACT

2 Phenotypes vary hierarchically among taxa and populations, among genotypes within
3 populations, among individuals within genotypes, and also within individuals for repeatedly
4 expressed labile phenotypic traits. This hierarchy produces some fundamental challenges to
5 clearly defining biological phenomena and constructing a consistent explanatory framework. We

6 use a heuristic statistical model to explore two consequences of this hierarchy. First, although the
7 variation existing among individuals within populations has long been of interest to evolutionary
8 biologists, within-individual variation has been much less emphasized. Within-individual
9 variance occurs when labile phenotypes (behaviour, physiology, and sometimes morphology)
10 exhibit phenotypic plasticity or deviate from a norm-of-reaction within the same individual. A
11 statistical partitioning of phenotypic variance leads us to explore an array of ideas about residual
12 within-individual variation. We use this approach to draw attention to additional processes that
13 may influence within-individual phenotypic variance, including interactions among
14 environmental factors, ecological effects on fitness consequences of plasticity, and various types
15 of adaptive variance. Second, our framework for investigating “variance of variance” reveals that
16 interactions between levels of the hierarchy form the preconditions for the evolution of all types
17 of plasticity, and we extend this idea to the residual level within individuals, where both adaptive
18 plasticity in residuals and canalization-like processes (stability) can evolve. With the statistical
19 tools now available to examine heterogeneous residual variance, an array of novel questions
20 linking phenotype to environment can be usefully addressed.

Key words: plasticity, canalization, variance sensitivity, gene–environment interaction,
phenotypic stability, bet-hedging, reaction norm.

21

22 CONTENTS

23 I. Introduction

24 (1) A statistical framework

- 25 II. Explanations for heterogeneity in residual within-individual variance
- 26 (1) Sampling or measurement error and the influence of bias
- 27 (2) The inaccurate or incomplete model hypothesis
- 28 (a) Non-linear reaction norms
- 29 (b) Slope–intercept covariance
- 30 (c) Multidimensional reaction norms
- 31 (3) Organismal error
- 32 (4) Random residual within-individual variance
- 33 (a) Passive plasticity
- 34 (b) Adaptive residual variance
- 35 III. Interactions across hierarchical levels: genotypic and individual differences in
- 36 residual within-individual variation
- 37 IV. Discussion
- 38 V. Conclusions
- 39 VI. Acknowledgements
- 40 VII. References

41 I. INTRODUCTION

42 Phenotypic variance shows a distinctly hierarchical pattern, with variance existing among taxa,
43 among populations within species, and among individuals within populations (Fig. 1). Among-
44 species and among-individual phenotypic variation have been a central focus of evolutionary
45 thinking since Darwin and Wallace connected the two through the process of natural selection.
46 Many traits, such as behaviour, physiology, and some morphological characteristics, are

47 expressed at different instances multiple times within the lifetime of an individual (Fig. 1). Such
48 traits also exhibit within-individual variation. Increasingly, within-individual variation is being
49 integrated into evolutionary theory (e.g. Nussey, Wilson & Brommer, 2007; Dingemans &
50 Dochtermann, 2013), but major gaps exist in our knowledge of processes affecting this level in
51 the hierarchy of variance. This is surprising given that genetic differences in within-individual
52 phenotypic variance are necessary for the evolution of any mechanism for an individual to
53 respond flexibly to the environment. Such mechanisms range from gene regulation within
54 individual cells to whole nervous systems. Conversely, organisms are also under selection to
55 maintain phenotypic integrity and reduce within-individual variance across environmental
56 conditions that may fluctuate within the lifetime of the individual (e.g. Cannon, 1929). These
57 fundamental attributes of organisms that control phenotypic expression arise out of patterns of
58 within-individual variation.

59 **(1) A statistical framework**

60 A hierarchical structure to phenotypic variance, as shown schematically in Fig. 1, is well
61 suited to descriptions using statistical models. We pursue this idea with four general messages in
62 mind. First, a complete description of the hierarchy will aid biological understanding of
63 phenotypic variance. Second, hierarchical descriptions of phenotypic variance highlight the fact
64 that patterns at one level in the hierarchy are often non-independent from processes acting at
65 other levels. Natural selection leading to evolution is the clearest example of this; variance
66 among individuals is necessary for selection, and this within-population process leads to variance
67 among units at higher levels (e.g. populations, species). A complete partitioning of variance at
68 levels within the individual may reveal other potential examples of cross-level effects. Third,

69 another improvement to understanding arises because being explicit about hierarchical variance
70 and the patterns produced raises challenges for current definitions of a variety of phenomena,
71 including plasticity, developmental stability and canalization. While we do not focus on those
72 issues directly herein, we will point out a few of the important implications that some variance
73 terms have for these concepts. Finally, and perhaps most importantly, fully partitioning variance
74 reveals patterns that demand explanation, and this can lead to new hypotheses about biological
75 processes. Our review begins to identify some potential patterns and some of the intriguing
76 hypotheses that may explain them.

77 Because within-individual phenotypic change constitutes a major subtype of phenotypic
78 plasticity, much is known about particular aspects of the biology of within-individual variance.
79 We suggest, however, that there is an additional level to phenotypic variance that exists inside
80 within-individual variance. This is residual within-individual variance, or unexplained within-
81 individual variance (see Glossary in Table 1). This variance is not well integrated into
82 evolutionary theory, leading to recent calls for more attention to be paid to this variance
83 component (e.g. Cleasby & Nakagawa, 2011; Stamps, Briffa & Biro, 2012; Nicolaus *et al.*,
84 2013). Thus, besides the general goals outlined above, herein we specifically explore three ideas
85 related to residual within-individual variance: (1) patterns of apparently unexplained within-
86 individual variance can provide clues to the existence of several important but possibly hidden
87 biological processes; (2) this component of variance may itself evolve from several interesting
88 types of adaptive processes; and (3) because within-individual variance is a distinct level in the
89 hierarchical structuring of phenotypic variance, interactions with other levels are likely integral
90 to many biological processes linking phenotype to environment. We review what is known about

91 the processes affecting within-individual variance and draw connections between previously
92 poorly linked ideas.

93 A full model of the hierarchy shown in Fig. 1 would be cumbersome, so here we focus
94 first on the among- and within-individual levels within a single population of the same species.
95 We begin with a statistical description of an observed phenotype. One common approach is to
96 partition sources of variation using the quantitative genetics equations where variance in
97 phenotype (V_P) is parceled into variance due to genetics (V_G) versus environment and error (V_E).
98 Many patterns of phenotypic variation have been explored using versions of this equation (e.g.
99 Lynch & Walsh, 1998; Moore, Brodie & Wolf, 1997; Tonsor, Elnaccash & Scheiner, 2013).
100 Here, we use the related ‘phenotypic equation’ (Nussey *et al.*, 2007; Dingemanse *et al.*, 2010)
101 that describes the component parts of each observation of the phenotype, Y . We consider
102 observations taken from a population across a sample that includes replication within each
103 individual, assuming that Y is a continuous character measured for instance i of individual j :

$$104$$
$$105 \quad Y_{ij} = (\beta_0 + ind_{0j}) + (\beta_1 + ind_{1j})E_{ij} + e_{0ij} \quad (1)$$
$$106$$

107 where β_0 is the population-mean phenotype [at the position where the value of the mean-centred
108 environmental gradient (E_{ij}) equals zero; *cf.* Dingemanse & Dochtermann, 2013]; ind_{0j} represents
109 the deviation from that mean for the j^{th} individual, β_1 the population-mean slope with respect to
110 E_{ij} , ind_{1j} the deviation in slope of the j^{th} individual from the population-mean slope, and e_{0ij} the
111 residual deviation of the i^{th} instance from individual j 's estimated reaction norm. The term e_{0ij}
112 represents the focus of this paper: unexplained deviations in phenotype within individuals. We
113 thus explicitly distinguish between among-individual variation (the differences in average value

114 between individuals), within-individual variation (differences between observations of the same
115 individual) and residual variation, which herein we will refer to explicitly as unexplained within-
116 individual variation (see also Table 1).

117 Equation (1) describes a linear mixed-effect model (or “random regression”). This
118 equation is commonly used to investigate phenotypic plasticity (Nussey *et al.*, 2007), defined as
119 the effect of an environmental factor on the phenotype with β_0 and β_1 describing the intercept and
120 slope of the population mean norm-of-reaction (*sensu* Woltereck, 1909). The concept of
121 plasticity cuts across two levels of phenotypic variance: (1) within-genotype among-individual
122 variance, which we will call ‘developmental plasticity’ because this variance is caused by
123 environmental effects during development (Table 1); and (2) within-individual variance. In
124 equation (1), β_1 refers to population average within-individual plasticity (see also the glossary in
125 Table 1 for synonyms). The term ind_{0j} is determined by including ‘random intercepts’ for
126 individual identity into the model, and variation among individuals in intercepts ($V_{ind_{0j}}$) is
127 consequently estimated. This variance component may reflect either genetic variance or
128 environmental factors that have carry-over effects from one instance of expression to another
129 (e.g. developmental plasticity: Lynch & Walsh, 1998; Wilson *et al.*, 2008; Dingemanse & Wolf,
130 2013; Snell-Rood, 2013). The other individual term, ind_{1j} , is similarly determined by including a
131 random effect (on the slope) arising from an interaction between individual and the
132 environmental variable (E_{ij}). This is individual plasticity and the associated estimate of variance
133 among slopes ($V_{ind_{1j}}$) captures differences between individuals in how they change their
134 phenotype in response to changes in the environment they experience. Individual plasticity could
135 also have genetic variance (e.g. $V_{G \times E}$) or also arise from carry-over effects of other
136 environmental factors (e.g. $V_{PE \times E}$ or $V_{G \times PE \times E}$, where PE indicates permanent environmental

137 effects; Schaeffer, 2004; Nussey *et al.*, 2007; Dingemanse *et al.*, 2010). Genetic variance in
138 patterns of within-individual variance is an important element of hypotheses about the evolution
139 of within-individual variance.

140 We focus here on the residual deviation (e_{0ij}) in equation (1): the deviation of observation
141 i from individual j 's reaction norm. Residual variance (variance in e_{0ij} , or V_{e_0}) is thus the amount
142 of within-individual variance not explained by other terms in the model. Residual variance is
143 important statistically because it forms the basis for testing whether sufficient evidence exists to
144 reject a statistical null hypothesis about included terms (Cleasby & Nakagawa, 2011). Most
145 statistical tests assume that residual variance is distributed normally and uniformly (i.e. residual
146 variance should not differ between individuals or along the environmental gradient). However,
147 because residual variance is never actually random and contains overlooked biology, this
148 assumption of homogeneity may often be false (Dutilleul & Potvin, 1995; Cleasby & Nakagawa,
149 2011) and this can have some important effects (Nicolaus *et al.*, 2013).

150 We argue that residual variance is of interest well beyond the question of whether the
151 appropriate statistical model was used to test hypotheses about other terms in the model.
152 Residual within-individual variation often amounts to the largest component of variation for
153 many labile traits, sometimes as much as 60% (e.g. Bell, Hankison & Laskowski, 2009;
154 Westneat *et al.*, 2011; Tonsor, Elnaccash & Scheiner, 2013). An appreciation for the processes
155 that cause residual within-individual phenotypic variance, and particularly heterogeneity in
156 residuals, will generate empirical advances and stimulate new conceptual or theoretical insights.
157 Indeed, because differences between individuals in sources of within-individual variance are
158 required for the evolution of mechanisms for both phenotypic stability and flexibility, most
159 biological phenomena are linked in some way to heterogeneous residual within-individual

160 variances. We explore the causes of this type of variance in more detail, and review the ideas on
161 processes acting at this level of phenotypic variance and the empirical work focused on those
162 ideas.

163 **II. EXPLANATIONS FOR HETEROGENEITY IN RESIDUAL WITHIN-** 164 **INDIVIDUAL VARIANCE**

165 Consider some hypothetical data collected from a single individual across an environmental
166 gradient (Fig. 2). A simplification of equation (1) yields:

$$167 \quad Y_{i1} = \beta_{01} + \beta_{11}E_{i1} + e_{0i1} \quad (2)$$

168 where β_{01} is the focal individual's mean (i.e. $\beta_0 + ind_{01}$ from equation 1), β_{11} is its slope with
169 respect to E_{i1} (i.e., $\beta_1 + ind_{11}$ from equation 1), and e_{0i1} represents the deviation of the i^{th} value
170 from the reaction norm of individual $j=1$. It is clear from Fig. 2 that e_{0i1} is not homogenous,
171 because the values tend to deviate to a greater extent from the individual's reaction norm (i.e. the
172 fitted line) at higher values of E_{i1} . There are a number of possible explanations for such
173 heterogeneous residuals. In order to be complete, we first consider non-biological explanations,
174 but we will focus on interesting but relatively unexplored biological explanations for
175 heterogeneous residual within-individual variance.

176 **(1) Sampling or measurement error and the influence of bias**

177 Sampling and measurement error are inevitable consequences of empirical data collection. We
178 make two brief points about this source of residual variance. First, not all residual variance is due
179 to sampling or measurement error, and it is these other sources that we explore in more detail
180 below. Second, sampling and measurement error may not be homogenous. Measurement error

181 can depend on the magnitude of the measured variable or on the conditions under which it is
182 measured (Viswanathan, 2005). Consider, for example, measures of parental care in which the
183 load of food brought by a parent on each visit to the nest is measured. When nestlings are small,
184 load sizes are small, and measurement error is typically less than the mean load size. When
185 nestlings are older, load sizes can be 4–5 times the size seen at the earlier age, and the
186 measurement error is often several times greater in magnitude than that at the earlier age. Such
187 examples of differences in measurement error across an environmental gradient often have a
188 biological explanation and should be accounted for.

189 **(2) The inaccurate or incomplete model hypothesis**

190 A second common interpretation of residuals in a statistical model is that they include the effects
191 of variables the investigator has not included in the model. This will likely be the case in both
192 field and laboratory studies (see Dingemanse & Dochtermann 2014; Niemelä & Dingemanse
193 2014), and if the research is testing specific, hypothesized influences then lumping everything
194 else into the residual variance is sufficient to proceed. However, an alternative goal may be to
195 seek new explanations, in which case attending to the residual variance may be valuable.
196 Heterogeneity in residual within-individual variance may provide hints concerning the existence
197 of various additional and potentially interesting biological processes that might be affecting
198 phenotypic expression. We discuss here a number of key candidates.

199 *(a) Non-linear reaction norms*

200 Within-individual heterogeneous residuals might occur when reaction norm slopes are modelled
201 as linear but individuals vary in the extent of non-linearity (Fig. 3A). Non-linear reaction norms
202 may exist when there are thresholds for shifting between one phenotype and another (e.g.

203 Moczek *et al.*, 2002), such as in Atlantic salmon (*Salmo salar*) that exhibit genetic variation for
204 maturation thresholds affecting alternative reproductive tactics (Piche, Hutchings & Blanchard,
205 2008). Most studies of thresholds have investigated non-labile traits. Thresholds in labile traits
206 also exist and show individual variation. For example, humans differ in the threshold at which a
207 skin irritant elicits a behavioural response (Smith *et al.*, 2004) and the threshold time to process a
208 perceptual task (e.g. Brock, Xu & Brooks, 2011).

209 Non-linear but continuous (e.g. parabolic) reaction norms may also be common. For
210 example, Brommer, Rattiste & Wilson (2010) found that annual reproductive success in a long-
211 lived gull increased and then declined with age. Provisioning behaviour of parent birds also
212 exhibits non-linearity with respect to offspring age (Westneat *et al.*, 2011). However, both of
213 these examples illustrate the shape of population mean reaction norms; little is known about
214 individual variation in non-linear reaction norms, especially parabolic ones, or the underlying
215 mechanisms that produce them. Individual variation in non-linearity may thus be of considerable
216 biological interest.

217 (b) Slope–intercept covariance

218 Heterogeneous residual within-individual variance may also arise if there is covariance between
219 individual intercepts (ind_{0j}) and slopes (ind_{1j}), and the slope and covariance terms are not
220 included in the phenotypic equation (Fig. 3B) — a frequent practice when a common reaction
221 norm is assumed for all individuals. Only a few studies have documented covariances between
222 intercepts and slopes (Mathot *et al.*, 2012), and in no case is the cause fully understood.
223 Kontiainen *et al.* (2009) found that nest defence intensity of Ural owls (*Strix uralensis*) varied
224 among individuals and yet was plastic with respect to the abundance of voles. Individual
225 aggressiveness varied in how responsive it was to vole abundance, and more aggressive

226 individuals were more plastic (positive covariance between intercept and slope). Slope–intercept
227 covariance of this sort may reflect important biological processes. Mathot *et al.* (2012) suggest
228 that such relationships may arise due to specific adaptations to environmental uncertainty, which
229 cause the magnitude of the intercept (e.g. in sampling effort or fat stores) strategically to
230 predetermine any associated responsiveness in adaptive plasticity to environmental change.
231 Alternatively, such covariances may arise from other types of constraints. For example, the
232 aggressiveness of Ural owls may be state dependent (Konttinen *et al.*, 2009), and state may
233 change with vole abundance, possibly non-linearly. Parent house sparrows (*Passer domesticus*)
234 cannot feed very young nestlings at a high rate perhaps because of nestling digestive constraints,
235 hence either differences in peak provisioning rates (variation in intercept) or in the ability to
236 assess changing offspring need (variation in slope) could drive a positive covariance between
237 them (Westneat *et al.*, 2011). Thus positive (or negative) covariance between intercept and
238 slopes, which can be buried in the residual variance, could potentially be driven by some
239 interesting, yet relatively unknown, biology (e.g. Dingemanse *et al.* 2012).

240 (c) *Multidimensional reaction norms*

241 Interactions among environmental factors affecting plastic phenotypes can also create
242 heterogeneous residuals if not included in the phenotypic equation. Organisms live in
243 environments that vary in many ways, and phenotypes could be a function of more than one
244 environmental factor simultaneously. For example, herbivory and competition for light influence
245 growth and changes in defensive compounds in plants (e.g. *Arabidopsis thaliana*; Cipollini,
246 2004) and temperature interacts with food type to influence growth rate in larval insects (e.g.
247 Kingsolver *et al.*, 2006; Stillwell *et al.*, 2007). These examples involve effects causing between-
248 individual differences *via* developmental plasticity, but multiple environmental factors can

249 obviously also affect within-individual variance in phenotype, and hence the residual variance in
250 equation (1). For example, in house sparrows, breeding attempt order and date in the season
251 interact to affect clutch size (Westneat, Stewart & Hatch, 2009), and nestling age and brood size
252 interact to affect parental feeding rate (Westneat *et al.*, 2011).

253 We label reaction norms that occur in response to more than one environmental factor
254 “multidimensional” norms of reaction (Westneat *et al.*, 2009). Multidimensionality can produce
255 heterogeneous residual within-individual variance in two ways. First, sensitivity of the
256 phenotype to additive effects of two or more environmental variables can produce this type of
257 heterogeneity if individuals experience only subsets of both environments. All individuals in a
258 population might share the same reaction norm that is responsive additively to two
259 environmental factors (E_1 and E_2). If E_1 is more variable at some values of E_2 , such as if
260 territories with good food supplies also had more stable temperatures, then individuals on good
261 territories might be less variable than those on poor territories (e.g. Charmantier & Garant,
262 2008). While some of these effects could be fixed by better sampling by the researcher, the case
263 of territory effects illustrates the more interesting possibility that expression of one phenotype
264 could alter the environments experienced (a phenotype–environment correlation) and thereby
265 affect expression of another phenotype; in this way multidimensionality combined with a
266 phenotype–environment correlation may be the underlying cause of heterogeneous residuals.

267 Second, the phenotype may be sensitive to a non-additive (i.e. interactive) effect of two
268 or more environmental factors. This interaction can create unequal variances across one of the
269 environmental gradients (e.g. Fig. 4). For datasets of repeatedly expressed traits,
270 multidimensionality can easily be incorporated in the phenotypic equation by constructing

271 models with more than one environmental gradient (e.g. E_1 and E_2 instead of just E) plus their
272 interactions (e.g. parental provisioning rates: Westneat *et al.*, 2011).

273 Non-additive effects have implications for understanding both the ecology of plasticity
274 and the organismal mechanisms producing it. An interaction effect may arise because of some
275 constraint to a process involved in the trait of interest. For example, an influence of host plant
276 diet on the thermal reaction norm of insects may arise in part because phenolic compounds
277 present in some diets are harder to process at cooler temperatures (e.g. Diamond & Kingsolver,
278 2012). Alternatively, interaction terms may arise because environmental variables affect fitness
279 trade-offs in ways that produce multiple fitness peaks. For example, within-individual variance
280 in clutch size in sparrows is influenced by an interaction between date in the season and nesting
281 attempt order (Westneat *et al.*, 2009). This appears consistent with life-history theory that
282 incorporates a seasonal decline in offspring quality (Rowe, Ludwig & Schluter, 1994). In this
283 model, multiple breeding episodes create separate adaptive ridges with respect to date for each
284 nesting attempt, producing multidimensional reaction norms affected by interactions between
285 date and nesting attempt order. Such circumstances could select for the integration of multiple
286 environmental cues.

287 Multidimensionality in reaction norms affects interpretations about tests of theory. For
288 example, evolutionary theory on pleiotropic effects leading to senescence suggests that genetic
289 variation in fitness should increase at older ages. Brommer *et al.* (2010) analysed declines with
290 age in reproductive performance (annual fitness) in common gulls (*Larus canus*), showing
291 among-individual variance, but little additive genetic variance, in slope with respect to age. Yet,
292 residual variance increased with age. This heterogeneity in residuals suggests the possibility of
293 multidimensionality — that is, as individuals age they may be increasingly susceptible to the

294 impact of other environmental factors, possibly in non-additive ways. Multidimensionality would
295 explain the change in residual variance with age, and the expected impact of pleiotropy on
296 genetic variance in fitness might be resurrected if there was genetic variation in the interaction
297 between age and environment on performance. Evidence supports the idea that genetic variance
298 for such interactions exists (e.g. Kingsolver *et al.*, 2006; Stillwell *et al.*, 2007), although we
299 know of no studies demonstrating such effects on within-individual variation. In general, the
300 biology of multidimensional reaction norms is likely to be quite important both for evolutionary
301 hypotheses regarding plasticity and for understanding underlying mechanisms of phenotypic
302 development, with non-additivity raising challenging questions about the mechanisms by which
303 variance in environment produces phenotypic variance.

304 **(3) Organismal error**

305 Many reaction norms arise from some mechanism of assessing an environmental factor (called
306 “active plasticity”; Scheiner, 2006). Errors in assessment (e.g. Reeve, 1989; Wiley, 1994;
307 Sherman, Reeve & Pfennig, 1997; DeWitt, Sih & Wilson, 1998; Auld, Agrawal & Relyea, 2010)
308 can produce phenotypes that deviate from the correct one. In other words, the deviations from
309 the line in Fig. 2 occur horizontally, and they arise from the organism misidentifying the cue to
310 the environment on the x -axis and producing a phenotype that would be better suited to a
311 different environment. Such errors in plasticity occur at both the among-individual and within-
312 individual levels; we refer to this phenomenon as “organismal error” (see glossary in Table 1 for
313 synonyms) simply to separate these errors from researcher measurement error. These types of
314 error contribute to limited plasticity (Moran, 1992; Getty, 1996; DeWitt *et al.*, 1998; Auld *et al.*,
315 2010), and occur whenever assessment mechanisms (broadly defined) are involved in phenotype

316 production. For example, misidentification of self as non-self by the immune system leads to
317 inappropriate activation of the immune system producing auto-immune disorders (Golub &
318 Green, 1991). Errors in growth processes during development may underlie fluctuations in the
319 symmetry of paired attributes (e.g. Van Valen 1962; Hansen, Carter & Pélarbon, 2006). Finally,
320 inappropriate behaviour may arise because of inadequacies of assessment at the sensory level
321 (e.g. Wollerman & Wiley, 2002) or the ways in which information is integrated as is exhibited
322 by increased error when attention is divided (e.g. Dukas, 1998).

323 Organismal errors could be heterogeneous for several reasons. First, organismal error is
324 likely proportional to the cue's scale (known as Weber's Law; Ross & Murray, 1996). This
325 effect is known in all sensory modalities and impacts many types of cues, including assessment
326 of time (e.g. Gibbon, 1977). Heterogeneity might also arise because multidimensionality leads to
327 problems with integration. For example, the inappropriate activation (response to internal
328 environment) of the immune system that leads to autoimmune disease can be exacerbated by
329 exposure to some bacterial pathogens, a second environment due to an external invader (Playfair,
330 1995). Phenotypic imprecision due to error may itself be influenced by developmental processes.
331 Deviations from a target phenotype can be compensated for in some cases (e.g. Kellner &
332 Alford, 2003), and therefore the magnitude of such noise may vary through ontogeny. Finally,
333 heterogeneous phenotypic expression could also reflect heterogeneous selection if either the
334 fitness consequences of the inaccurate phenotype or the costs to improving precision differ along
335 the range of the environment gradient. For example, in birds, brood parasites produce
336 circumstances in which errors in egg recognition by hosts reduce host fitness. As the rate of
337 parasitism increases, the costs of acceptance increase, and indeed, acceptance rates decline with
338 increases in parasitism (e.g. Lindholm & Thomas, 2000; Stokke *et al.*, 2008). Preventing

339 parasitism appears costly because acceptance increases when parasitism levels decline (Brooke,
340 Davies & Noble, 1998). Acceptance errors may be influenced by learning (e.g. Rothstein, 1978),
341 and so they can exhibit within-individual plasticity (Lotem, Nakamura & Zahavi, 1995), but it is
342 not known if variance in either the costs of accepting a parasite egg or the cost of discriminating
343 among eggs influences acceptance errors within an individual.

344 (4) **Random residual within-individual variance**

345 A final possibility is that phenotypes vary due to truly random processes. We describe two major
346 ways this could occur, which differ in the mechanism that connects the environment with the
347 phenotypic effect.

348 (a) *Passive plasticity*

349 The phenotype could exhibit “passive” plasticity (Scheiner, 2006) in which purely physical
350 processes create phenotypic variation. Fluctuations in body temperature in ectotherms due to the
351 physics of heat transfer and changes in ambient air temperature could be seen as one example of
352 passive plasticity. More convincingly, foraging success, measured as the time to find the next
353 food item, might exhibit passive plasticity as a result of changes in the density or distribution of
354 prey items. Thus food intake rate will have a component of variation that arises from the physical
355 constraint that food cannot be ingested before it is found, and the time taken to find the next prey
356 item will show some unpredictable variance because the location of a particular prey item is
357 usually not known by the forager when they start foraging. In both cases, some portion of the
358 phenotypic variance arises due to passive plasticity and the environmental factor causing this
359 may be unpredictable. The phenotype thus contains some stochastic variation that, if not
360 otherwise accounted for, would be present in the residual phenotypic variance. Because the

361 environmental factor causing passive plasticity may be associated with other factors, then, as
362 may occur with multidimensional reaction norms, this could create heterogeneity of residual
363 variance (e.g. Stearns & Kawecki, 1994).

364 Residual variance caused by unpredictable passive plasticity could have fitness
365 consequences creating stabilizing selection on reaction norms. This could produce a process akin
366 to canalization, occurring at the within-individual level, whereby the phenotype is stabilized
367 around the optimal reaction norm (Stearns & Kawecki, 1994). However, there are also
368 circumstances when increased residual variance may be favoured. Variance-prone foraging is
369 one example (Stephens, 1981). Encounters with prey may be unpredictable, but if the variance in
370 encounter times can be assessed by foragers, then individuals can make decisions to experience
371 either more or less unpredictable passive plasticity in the instantaneous food-capture rate (Shafir,
372 2000). Certain state variables, such as energy reserves, are predicted to create selection favouring
373 either variance-averse or variance-prone behaviour (Caraco, Martindale & Whittam, 1980;
374 Stephens, 1981). Studies of this idea in captivity have produced some equivocal results that may
375 be resolved by accounting for the scale of environmental variance (reviewed in Shafir, 2000), but
376 there remains a lack of studies conducted in the wild that properly test for such effects of
377 ecologically relevant state variables (but see Ratikainen, Wright & Kazem, 2010).

378 *(b) Adaptive residual within-individual variation*

379 Adaptive residual phenotypic variation could be induced by mechanisms incorporating
380 stochasticity into phenotype expression. When this occurs at the level of among-individual but
381 within-genotype variation, it can be a mechanism for adaptive phenotypic polymorphisms
382 (phenotype switching, e.g. Kussel & Leibler, 2005), polyphenisms (Mayr, 1963; Van Dooren,
383 2001) and diversification bet-hedging (Gillespie, 1973; Frank & Slatkin, 1990; Simons, 2011).

384 Residual within-individual variation in a wide variety of traits could be adaptive (Table
385 2). Some phenotypes emerge from processes that initially generate (possibly random) variation
386 and then involve mechanisms that reduce this variation within individuals (Frank, 1997). In the
387 vertebrate immune system, clonal selection within a highly diverse population of B-cells is a
388 central process for adaptive immunity (Golub & Green, 1991). Similar processes may occur at
389 the cellular level for epidermal or neural tissues (e.g. Changeux & Danchin, 1976; Kagan,
390 Novoplansky & Sachs, 1992). Likewise, some learning processes such as trial-and-error learning
391 may involve generating variation followed by a mechanism of sorting among options within the
392 individual (Frank, 1997). For example, in jumping spiders, individuals produce a large array of
393 signals oriented toward potential prey (other spiders); appropriate feedback from the prey then
394 leads to repetition of the effective signal (Jackson & Wilcox, 1993). Such mechanisms would
395 produce heterogeneous residuals in phenotypic variation across either time or specific
396 environmental gradients, and could evolve through differential selection among genotypes
397 producing different patterns of within-individual variation.

398 Reduced residual within-individual variance may be adaptive. Theory suggests that the
399 presence of conspecific observers during contests can favour predictable levels of aggression
400 (e.g. Johnstone, 2001; Nesse, 2001), which would reduce within-individual variance in
401 aggression. In complex social groups, particular social niches may exist, and individuals taking
402 on those roles may behave less variably (e.g. Bergmüller & Taborsky, 2010). Similarly, Schuett,
403 Tregenza & Dall (2010) hypothesize that sexual selection on male behaviour might produce
404 sexual dimorphism in within-individual variance in behaviour.

405 Alternatively, unpredictability *per se* may be favoured. Being unpredictable could in
406 some conditions lead to higher rates of winning in contests (e.g. Whiten & Byrne, 1997).

407 Variable display intensity by individual combatants is favoured in war-of-attrition contests
408 (Maynard Smith, 1974). Similarly, variable waiting times for both predator and prey, possibly at
409 both the among- and within-individual levels, are favoured when the prey is in refuge and the
410 predator waits for them to emerge (Hugie, 2003). Briffa (2013), for example, found that residual
411 variation in a startle response, after controlling for individual identity and mean plasticity,
412 increased in the presence of additional cues to a predator. This type of unpredictable behaviour
413 might also increase as individuals become familiar with individual predators (Stamps *et al.*,
414 2012).

415 An intriguing example of potentially adaptive stochastic variance may occur at the sub-
416 cellular level. There is growing evidence that a number of molecular events, including gene
417 regulation, are subject to stochastic variation from cell to cell within the individual (e.g. Eldar &
418 Elowitz, 2010). This may arise because of relatively small copy numbers of some key molecules
419 (e.g. DNA, some large regulatory proteins) within the cell that produce differences in rates of
420 chemical contacts from cell to cell. Some of this variation might be considered passive plasticity,
421 but in some cases it may be adaptive. For example, in yeast a suite of genes is regulated by
422 calcium, and the main transcription factor Crz1 exhibits apparently stochastic bursts of up-
423 regulation. Cai, Dalal & Elowitz (2008) show that stochastic bursting, which varies in response
424 to calcium, produces more uniform co-regulation across an array of downstream genes because
425 of proportional control — that is, the proportion of time Crz1 is bursting produces stronger
426 correlations between downstream products than would more modulated amplitudes or durations
427 of individual bursts. Hence within-individual residual variation in Crz1 activation oddly results
428 in more coordinated control of other genes than would less stochastic Crz1 regulation. Stochastic
429 bursts of up-regulation may also be the underlying molecular explanation for the generation of

430 variable populations of cells involved in internal selection during development (Losick &
431 Desplan, 2008).

432 In summary, adaptive residual within-individual phenotypic variation may exist across
433 several levels in organismal organization, from sub-cellular to organismal. We have described
434 some hypotheses that might explain such variation, but the array of studies that have directly
435 focused on these is remarkably small.

436 **III. INTERACTIONS ACROSS HIERARCHICAL LEVELS: GENOTYPIC AND** 437 **INDIVIDUAL DIFFERENCES IN RESIDUAL WITHIN-INDIVIDUAL VARIATION**

438 We have emphasized that residual within-individual variance occurs in the context of a hierarchy
439 of variances, ranging from the within-individual level on up to higher levels of taxonomic
440 organization (Fig. 1). Thus, instances of phenotypic expression are nested within individuals,
441 individuals within genotypes, genotypes within populations, populations within species, and so
442 forth. A fascinating feature of this structure is that interactions between levels occur, and they
443 have far-reaching consequences.

444 One well-known example involves the genotype by environment interactions ($G \times E$)
445 depicted in quantitative genetics theory concerning the evolution of phenotypic plasticity (Via &
446 Lande, 1985; Gomulkiewicz & Kirkpatrick 1992). This is typically viewed as an interaction
447 between the among-genotype-within-a-population level (multiple genotypes exist within a
448 population) and the among-individuals-within-genotype level (multiple environments can be
449 experienced by different individuals with the same genotype, producing “permanent
450 environment”, PE, variation). But, since individuals can experience different environments in
451 their lifetimes, $G \times E$ also captures an interaction between the among-genotype level and the

452 within-individual level, leading some explicitly to distinguish between these two types of gene
453 by environment interaction ($G \times PE$ versus $G \times E$; Nussey *et al.*, 2007; Dingemanse *et al.*, 2010). A
454 potentially interesting possibility is multidimensionality across levels, with an interaction
455 between an environmental effect at the within-genotype level (E_{1j}) and another factor at the
456 within-individual level (E_{2ij}) (e.g. Weinig & Delph, 2001). A suitable modification of equation
457 (1) to include the genotypic level would account for developmental plasticity in behavioural
458 flexibility (Piersma & Drent, 2003; Stamps & Groothuis, 2010; Dingemanse *et al.*, 2010), and
459 this can be extended to include effects of environmental variables that interact but do so across
460 different timescales. For example, in birds, variation in maternal androgens present in the yolk of
461 eggs may affect the mean level of aggressive behaviour by these individuals as adults (Gil, 2008;
462 Müller *et al.*, 2012). Levels of aggression in any particular interaction are also influenced by the
463 value of a food resource (e.g. Chancellor & Isbell, 2008). An interesting but untested possibility
464 is that yolk androgens influence the way in which food value influences aggression (i.e. the slope
465 of a reversibly plastic response), producing a between-individual by within-individual
466 multidimensional reaction norm ($PE \times E$), with the possibility of there being genetic variance for
467 this (e.g. $G \times PE \times E$). We note that the distinction between environments that have developmental
468 and those that have only activational effects is often more subtle than typically portrayed; some
469 activational environmental effects (cue to a predator) can also have carryover effects through
470 processes such as learning (Dingemanse & Wolf, 2013), potentially producing complexities not
471 captured by current variance equations.

472 The hierarchical phenotypic variance structure may produce interactions or covariances
473 between elements of residual within-individual variance and the among-individual or the among-

474 genotype levels. To illustrate, we take the phenotype equation (1) and expand the residual
475 within-individual deviations (e_{0ij}) into its own equation:

$$476 \quad \sigma_{eij} = (\beta_{\sigma 0} + ind_{\sigma 0j}) + (\beta_{\sigma 1} + ind_{\sigma 1j})E_{ij} \quad (3)$$

477 where σ_{eij} describes the residual variance (Ve_{0i}) as having a population mean variance ($\beta_{\sigma 0}$), an
478 individual-specific deviation in variance from the mean ($ind_{\sigma 0j}$), and an effect of both population
479 and individual effects of environment on the variance ($\beta_{\sigma 1} + ind_{\sigma 1j}$). These latter terms capture
480 the heterogeneous nature of residual variance due to, in many cases, factors that influence the
481 phenotypic sensitivity to environmental factors.

482 This double equation, with equation (1) describing effects on means and the simultaneous
483 equation (3) capturing patterns in variances, has several important consequences. One is that
484 there may be interactions between elements of the residual variance (equation 3) and terms
485 present in the mean portion (equation 1). Equation (3) already includes one such interaction —
486 residual within-individual variance could vary among individuals. Equation (3) could be
487 expanded to include between-genotype, between-population, and between-species differences in
488 residual within-individual variation. Such effects would make the residual variance in a
489 particular trait behave as if it is a trait itself (Biro & Adriaenssens, 2013).

490 A second implication of the double equation is that there are new potential covariances
491 between terms within and between the two linked equations that are, as we detail below, of
492 biological interest. Some of these are evident in Fig. 5; we describe two in more detail here.

493 **Cov (ind_{0j} , $ind_{\sigma 0j}$):** the magnitude of an individual's reaction norm intercept could covary with
494 the magnitude of an individual's residual variance. Either positive or negative covariances are
495 possible; Fig. 5 depicts a negative covariance. This covariance seems likely to have a biological
496 basis since the magnitude of a phenotype and tight control over its variance in expression may be

497 linked. For example, aggressive individuals might exhibit less residual variance because they
498 may be less sensitive to extraneous stimuli (e.g. Natarajan *et al.*, 2009). In general terms,
499 processes involved in changing residual variance (e.g. canalization or behavioural stability) may
500 be integrated with processes producing mean phenotypes. A review of genetic variation in
501 environmental variance reports a handful of studies that have measured a genetic correlation
502 between mean phenotype and variance in phenotype, the majority of which are negative (Hill &
503 Mulder, 2010).

504 **Cov (ind_{1j} , $ind_{\sigma_{0j}}$):** the magnitude of an individual's reaction norm slope covaries with its within-
505 individual residual variance; also shown as negative in Fig. 5. Several potential examples of this
506 covariance exist; a positive covariance could perhaps be due to increases in plasticity making the
507 phenotype more sensitive to organismal error or the impact of other environmental factors. This
508 covariance is similar to one suggested for a relationship between developmental plasticity and
509 developmental instability (e.g. Hansen *et al.*, 2006; Tonsor *et al.*, 2013), which is a covariance
510 between a genotype's intercept and within-genotype among-individual deviations from the
511 genotype's reaction norm. Alternatively, an individual with a strong reaction to a particular
512 environmental gradient might be less sensitive to stochastic influences of other cues (e.g.
513 attentional focus; Dukas, 1998).

514 Other covariances with elements of stochastic residual within-individual variance are
515 possible, especially if other hierarchical levels of phenotypic variance are included. We also
516 expect interactions with other levels. For example, if residual within-individual variation itself is
517 to evolve, as we suspect it might, then there must be genetic variation for residual deviations.
518 Indeed, studies have uncovered evidence of genetic variation for environmental variance (e.g.
519 Hill & Mulder, 2010). Often these have lumped together many of the processes acting within the

520 individual (plasticity, all of the sources of heterogeneous residual variance discussed above). It is
521 not clear in any case that the genetic variance of any specific cause of heterogeneity in residuals
522 has been estimated. Hill & Mulder (2010) review a variety of methodological approaches and
523 some of the problems with each. Here, we note that an important implication of our treatment is
524 that attending to different potential sources of unexplained residual within-individual variance
525 and being able to assess the genetic variance in specific causes would fine-tune tests of
526 hypotheses about the evolution of phenotypic variance.

527 **IV. DISCUSSION**

528 Labile phenotypes, especially behavioural and physiological characters, exhibit substantial
529 within-individual variation. We emphasize that the presence of this variation is a large, mostly
530 untapped, opportunity to understand better the ecology of selection and evolution. The basic
531 logic here is powerful: if variance in phenotype within an individual has fitness consequences
532 and differences in within-individual variance exist between genotypes, then patterns of within-
533 individual variance can evolve. Presumably, it is exactly this process that has driven the variety
534 of mechanisms for assessing environments and producing adaptive reversible, or irreversible,
535 phenotypic plasticity that we now see in most organisms. Although within-individual plasticity
536 may be one of the most widespread of biological phenomena, it has usually been studied
537 indirectly and is not well integrated conceptually. More importantly for the purposes of this
538 review, variation in residual within-individual variance (that not explained by active plasticity)
539 also likely underlies how individuals maintain consistent phenotypes in the face of considerable
540 environmental variance. Such stability is an example of within-individual canalization, and has
541 also been understudied from the perspective of the evolution of reaction norms. Finally, we

542 expect residual within-individual residual variation to differ between individuals and genotypes,
543 and if so, heterogeneous residual within-individual variance may be as common as genetic
544 variance itself.

545 Residual within-individual variance in the phenotype is neither “noise” nor “random”
546 variance, despite the labels given it from the statistical assumptions needed for hypothesis
547 testing. It is, in fact, a rich source of clues about the biology of phenotypes. It is likely to be
548 heterogeneous for many reasons, and so it should be the explicit focus of investigation more so
549 than it currently is. Residual within-individual variance is often the largest component of
550 phenotypic variance for some phenotypes, such as behavioural traits (e.g. Bell *et al.*, 2009).
551 Clues as to its underlying biology can be gained by statistically exploring the structure of
552 residual variance, particularly for patterns of heterogeneity. To that end, several statistical
553 approaches have been developed to account appropriately for heterogeneous within-individual
554 residual variance in tests of hypotheses about other terms in a model, but they can be adapted to
555 explore patterning in residual variance directly (e.g. Breusch & Pagan, 1979; White, 1980; Lee &
556 Nelder, 1996; Smyth & Verbyla, 1999; Cleasby & Nakagawa, 2011; Westneat, Schofield &
557 Wright, 2013). Of critical importance here is that residual within-individual variance is modelled
558 analogously to means. Because residual within-individual variance can vary simultaneously with
559 respect to several variables (including individual identity), a mixed-model structure that accounts
560 for influences on both mean effects and residual variances within a single model has the most
561 potential to uncover new patterns. Recent techniques appear to accomplish this (Lee & Nelder,
562 1996; Smyth & Verbyla, 1999; Westneat *et al.*, 2013) and can be applied to datasets containing
563 repeated measures of phenotypes within individuals.

564 Models of mixed effects for both means and variances require large datasets. For
565 example, good estimates of variance terms in the mean portion of the model need 1000 data
566 points or more (e.g. Martin *et al.*, 2011; van de Pol, 2012). While applying our approach to some
567 rarely expressed traits may be a challenge, there are many morphological, physiological, and
568 behavioural traits that are expressed quite often. Consider, for example, feathers on a bird that
569 moults twice a year, leaves on a plant, eggs per spawn in a fish, or tendency to attack an
570 opponent in crickets. These traits are expressed dozens to hundreds of times in each individual,
571 and so massive datasets can be relatively easily collected. Empirical studies have detected
572 heterogeneous residuals in several different traits (reviewed by Nicolaus *et al.*, 2013) and in
573 some cases from modest-sized datasets collected for other purposes (e.g. Westneat *et al.*, 2013).
574 We think the phenotypic equation combined with other conceptual and empirical tools has the
575 potential to lead to a variety of novel hypotheses and experiments for many types of traits.

576 Our review emphasizes that the nature of residual within-individual variance is not
577 merely an empirical issue; several potentially important conceptual ideas have emerged from
578 considering the underlying reasons for residual phenotypic variance and the impact that such
579 variance might have on the evolutionary process. For example, our examination of residual
580 within-individual variance intersects with concepts of phenotypic plasticity, canalization, and
581 developmental stability. The specific relationships between these terms are often confusing and
582 there appears to be no general agreement on definitions (Dworkin, 2005). Some authors, for
583 example, view plasticity and canalization as opposites (e.g. Gibson & Wagner, 2000; Debat &
584 David, 2002; Nijhout & Davidowitz, 2003; Ghalambor, Angeloni & Carroll, 2010), whereas
585 others treat them as potentially independent phenomena (e.g. Stearns & Kawecki, 1994) although
586 they may be correlated (e.g. Tonsor *et al.*, 2013). Our focus on the phenotypic equation and our

587 treatment of within-individual residual variance as a component of variance within a hierarchy of
588 variances favours distinct but overlapping definitions. We do not have the space here to explore
589 all the nuances, but a brief example illustrates our point that the concepts of plasticity and
590 canalization can cut across several levels of phenotypic variance. Selection could act on a
591 particular trait to reduce environmentally induced variation in phenotype among individuals
592 within a genotype. We might call this within-genotype canalization of intercepts.
593 Simultaneously, selection might favour a more flexible within-individual phenotypic response to
594 the environment. A possible by-product of this might be higher within-individual residual
595 variance due to organismal error, meaning that at the within-individual level the organism is
596 simultaneously more plastic (steeper slope) and less canalized (higher residual variance), even
597 though the genotype is more canalized developmentally around the intercept. Improved clarity
598 about concepts and processes may be achieved by taking a more statistical approach to such
599 definitions and attending to the full hierarchical structure of variance, including residual within-
600 individual variance.

601 We also claim that residual within-individual variance deserves more attention because it
602 would bring renewed focus on the ecology of phenotypes. Molecular and quantitative genetics
603 have contributed major new insights into the genetics of phenotypes. Yet, our focus on reaction
604 norms and residual within-individual variance rests on how environments affect phenotypes.
605 Genotypes can interact with the environment at two levels — among individuals within genotype
606 and within individuals. The environment also has effects on within-individual phenotypic
607 variation in three distinct ways: *(i)* via within-individual plasticity, *(ii)* through several possible
608 impacts on within-individual residual variance, and *(iii)* due to effects on developmental
609 plasticity that change either within-individual plasticity or the nature of residual variance.

610 Finally, the environment influences the fitness consequences of phenotypic variation at each of
611 these levels. These influences of ecology have important ramifications, and while we have made
612 great strides in understanding the interface between ecology and phenotypic diversity, our
613 analysis here suggests that we could gain even more by attending to the ecology of individual
614 phenotypes in greater detail. This may be especially important in this time of rapid ecological
615 change.

616 Another emergent conclusion is that statistical models are more than a means to evaluate
617 particular biological hypotheses. As we have done here, the phenotypic equation can clearly also
618 be used to generate biological hypotheses. It is effective precisely because it is
619 phenomenological — it is a description of pattern in phenotype. Too often in biology we conflate
620 pattern and process in our terminology. Statistical descriptions allow for clearer definitions of
621 pattern, which then demand explanation. Phenotypic variation is an unusual blend of processes
622 that mimic statistical properties and those that actually incorporate variance, all combined in a
623 hierarchical structure (from individual to phylogeny) that is especially well suited for statistical
624 modelling.

625 Thus, the phenotypic equation may be viewed as a biological hypothesis in itself. It
626 models a hierarchical structure, and so thereby constitutes a hypothesis about the hierarchical
627 nature of phenotypic variance. This draws attention to each term in the equation and leads to
628 hypotheses regarding its potential biological importance. In this context, the residual term
629 becomes as important as the population mean. Moreover, we suggest that extensions of the
630 phenotypic equation can integrate patterns of phenotypic variance from within the individual up
631 to among taxa. Employing the phenotypic equation fully might catalyse a new integration of
632 micro and macro evolutionary processes, overcoming some of the problems with such

633 integration (e.g. Martin, Ton & Niklison, 2013). It could also provide the structure for assessing
634 the role of ecology on multiple scales (e.g. within individuals, among individuals, among
635 populations) simultaneously. Such considerations go beyond understanding the biology
636 underlying residual within-individual variance, but our systematic exploration of this one
637 element of the phenotypic equation is illustrative of the potential value of more fully integrating
638 statistical thinking into biology (e.g. Bolker *et al.*, 2009).

639

640 V. CONCLUSIONS

- 641 1. The hierarchical structure of phenotypic variance is especially amenable to hierarchical
642 statistical models, and applying such models highlights the potential importance of within-
643 individual residual variance. This variance term is more than “error”, and could contain
644 interesting patterns, such as heterogeneous residual variance. We review hypotheses that may
645 explain heterogeneity in within-individual residual variance in phenotype.
- 646 2. Our review reveals many relatively poorly studied phenomena that have potential theoretical
647 importance, including non-linear reaction norms, intercept-slope covariance,
648 multidimensional phenotypic plasticity, various forms of passive plasticity, and several types
649 of adaptive variance.
- 650 3. We find that the biology of within-individual residual variance cuts across multiple levels of
651 biological organization, from gene regulation within cells, to whole organism traits such as
652 physiology and behavior. Our investigation of heterogeneous residual variation also links
653 concepts from multiple fields. For example, canalization in developmental biology and
654 variance sensitivity in behavioral ecology have elements in common. Moreover, explicitly
655 considering the causes of phenotypic variance in a hierarchical framework reveals multiple

656 scales at which particular processes may occur, with some seemingly opposite processes
657 (e.g., canalization and plasticity) occurring simultaneously but at different levels in the
658 hierarchy.

- 659 4. By embedding within-individual residual variance at its appropriate level in the hierarchy of
660 phenotypic variance, we establish that residual variance can evolve. It is nested several levels
661 down from genotypic variance, and so may evolve in ways that are linked to individual
662 plasticity (within-individual level), developmental plasticity (among-individual within-
663 genotype level), and mean phenotype (among-genotype level). Such interactions may have
664 important implications for the ecology of selection and the process of evolution.
- 665 5. Methods are available to assess within-individual residual variance in a variety of repeatedly
666 expressed traits and statistically explore pattern in these residuals. With these tools, new
667 understanding of the ecology of phenotypes can be obtained.

668

669 **VI. ACKNOWLEDGEMENTS**

670 We thank our respective institutions and especially the Max Planck Society for supporting our
671 initial conversations about these ideas. We thank members of the Evolutionary Ecology group at
672 Kentucky for feedback following a presentation of the ideas in this paper, and Kim Mathot,
673 Marion Nicolaus, Christophe Pélabon, and two anonymous reviewers for their useful
674 suggestions. NJD was supported by the Max Planck Society.

675 **VII. REFERENCES**

676 AULD, J. R., AGRAWAL, A. A. & RELYEA, R. A. (2010). Re-evaluating the costs and limits of
677 adaptive phenotypic plasticity. *Proceedings of the Royal Society of London B*, **277**, 503–11.

- 678 BELL, A. M., HANKISON, S. L. & LASKOWSKI, K. L. (2009) The repeatability of behaviour: a
679 meta-analysis. *Animal Behaviour*, **77**, 771–783.
- 680 BERGMÜLLER, R. & TABORSKY, M. (2010) Animal personality due to social niche specialisation.
681 *Trends in Ecology and Evolution*, **9**, 504–511.
- 682 BIRO, P. A. & ADRIAENSSENS, B. (2013). Predictability as a personality trait: consistent
683 differences in intra-individual behavioral variation. *The American Naturalist*, **182**, 621–629
- 684 BOLKER, B. M., BROOKS, M. E., CLARK, C. J., GEANGE, S. W., POULSEN, J. R., STEVENS, M. H. H.
685 & WHITE, J. S. S. (2009). Generalized linear mixed models: a practical guide for ecology and
686 evolution. *Trends in Ecology and Evolution*, **24**, 127–135.
- 687 BREUSCH, T.S. & PAGAN, A.R. (1979). Simple test for heteroscedasticity and random coefficient
688 variation. *Econometrica*, **47**, 1287–1294.
- 689 BRIFFA, M. (2013). Plastic proteans: reduced predictability in the face of predation risk in hermit
690 crabs. *Biology Letters*, **9**, 20130592.
- 691 BROCK, J., XU, J. Y. & BROOKS, K. R. (2011). Individual differences in visual search: relationship
692 to autistic traits, discrimination thresholds, and speed of processing. *Perception*, **40**, 739–742.
- 693 BROMMER, J. E., RATTISTE, K. & WILSON, A. (2010). The rate of ageing in a long-lived bird is
694 not heritable. *Heredity*, **104**, 363–70.
- 695 BROOKE, M. D., DAVIES, N.B. & NOBLE, D. G. (1998). Rapid decline of host defences in
696 response to reduced cuckoo parasitism: behavioural flexibility of reed warblers in a changing
697 world. *Proceedings of the Royal Society of London B*, **265**, 1277–1282.
- 698 CAI, L., DALAL, C. K. & ELOWITZ, M. B. (2008). Frequency-modulated nuclear localization
699 bursts coordinate gene regulation. *Nature*, **455**, 485–490.

- 700 CANNON, W. B. (1929) Organization for physiological homeostasis. *Physiological Reviews*, **9**,
701 399-431.
- 702 CARACO, T. MARTINDALE, S. & WHITTAM, T. S. (1980). An empirical demonstration of risk
703 sensitive foraging preferences. *Animal Behaviour*, **28**, 820-830.
- 704 CHANCELLOR, R. L. & ISBELL, L. A. (2008). Punishment and competition over food in captive
705 rhesus macaques, *Macaca mulatta*. *Animal Behaviour*, **75**, 1939-1947.
- 706 CHANGEUX, J. P. & DANCHIN, A. (1976). Selective stabilization of developing synapses as a
707 mechanism for specification of neuronal networks. *Nature*, **264**, 705-712.
- 708 CHARMANTIER, A. & GARANT, D. (2008). Environmental quality and evolutionary potential:
709 lessons from wild populations. *Proceedings of the Royal Society of London B*, **272**, 1415–
710 1425.
- 711 CIPOLLINI, D. (2004). Stretching the limits of plasticity: can a plant defend against both
712 competitors and herbivores? *Ecology*, **85**, 28–37.
- 713 CLEASBY, I. R. & NAKAGAWA, S. (2011). Neglected biological patterns in the residuals.
714 *Behavioral Ecology and Sociobiology*, **65**, 2361–2372.
- 715 DEBAT, V. & DAVID, P. (2002). Mapping phenotypes: canalization, plasticity and developmental
716 stability. *Trends in Ecology and Evolution*, **16**, 555-561.
- 717 DEWITT, T. J., SIH, A. & WILSON, D. S. (1998). Costs and limits of phenotypic plasticity. *Trends*
718 *in Ecology and Evolution*, 13,77–81.
- 719 DIAMOND, S. E. & KINGSOLVER, J. G. (2012). Host plant adaptation and the evolution of thermal
720 reaction norms. *Oecologia*, **169**, 353–60.

- 721 DINGEMANSE, N. J., BARBER, I., WRIGHT, J. & BROMMER, J. E. (2012). Quantitative genetics of
722 behavioural reaction norms: genetic correlations between personality and behavioural
723 plasticity vary across stickleback populations. *Journal of Evolutionary Biology*, **25**, 485-496.
- 724 DINGEMANSE, N. J. & DOCHTERMANN, N. A. (2013). Quantifying individual variation in
725 behaviour: mixed-effect modelling approaches. *Journal of Animal Ecology*, **82**, 39-54.
- 726 DINGEMANSE, N. J. & DOCHTERMANN, N. A. (2014). Individual behaviour: Behavioural ecology
727 meets quantitative genetics. In: *Quantitative Genetics in the Wild* (eds. A. CHARMANTIER, D.
728 GARANT & L. E. B. KRUK). In press, Oxford University Press, Oxford, UK.
- 729 DINGEMANSE, N. J., KAZEM, A. J. N., RÉALE, D. & WRIGHT, J. (2010). Behavioural reaction
730 norms: animal personality meets individual plasticity. *Trends in Ecology and Evolution*, **25**,
731 81–89.
- 732 DINGEMANSE, N. J. & WOLF M. (2013). Between-individual differences in behavioural plasticity
733 within populations: causes and consequences. *Animal Behaviour*, **85**, 1031-1039
- 734 DUKAS, R. (1998). Constraints on information processing and their effects on behavior. In:
735 *Cognitive Ecology: The Evolutionary Ecology of Information Processing and Decision-*
736 *Making* (ed. R. DUKAS), pp. 89–128. University of Chicago Press, Chicago, IL.
- 737 DUTILLEUL, P. & POTVIN, C. (1995). Among-environment heteroscedasticity and genetic
738 autocorrelation: implications for the study of phenotypic plasticity. *Genetics*, **139**, 1815–
739 1829.
- 740 DWORKIN, I. (2005). Canalization, cryptic variation, and developmental buffering. In: *Variation:*
741 *A Central Concept in Biology* (eds. B. HALGRIMSSON & B. K. HALL), pp. 131-158. Academic
742 Press, New York, NY.

- 743 ELДАР, A. & ELOWITZ M. B. (2010). Functional roles for noise in genetic circuits. *Nature*, **467**,
744 167-173.
- 745 FRANK, S. A. (1997). The design of adaptive systems: optimal parameters for variation and
746 selection in learning and development. *Journal of Theoretical Biology*, **184**, 31-39.
- 747 FRANK, S. A. & SLATKIN, M. (1990). Evolution in a variable environment. *The American*
748 *Naturalist*, **136**, 244–260.
- 749 GETTY, T. (1996). The maintenance of phenotypic plasticity as a signal detection problem. *The*
750 *American Naturalist*, **148**, 378–385.
- 751 GHALAMBOR, C. K., ANGELONI, L. M. & CARROLL S. P. (2010). Behavior as phenotypic
752 plasticity. In: *Evolutionary Behavioral Ecology* (eds. D. F. WESTNEAT & C. W. FOX), pp 90-
753 107. Oxford University Press, New York, NY.
- 754 GIBBON, J. (1977). Scalar expectancy theory and Weber's law in animal timing. *Psychology*
755 *Reviews*, **84**, 279–325.
- 756 GIBSON, G. & WAGNER, G. (2000). Canalization in evolutionary genetics: A stabilizing theory?
757 *Bioessays*, **22**, 372-380.
- 758 GIL, D. (2008). Hormones in avian eggs: physiology, ecology and behavior. *Advances in the*
759 *Study of Behavior*, **38**, 337–398
- 760 GILLESPIE, J. H. (1973). Polymorphism in random environments. *Theoretical Population*
761 *Biology*, **4**, 193–195.
- 762 GOLUB, E. S. & GREEN, D. R. (1991). *Immunology: A Synthesis*, 2nd ed. Sinauer, Sunderland,
763 MA.
- 764 GOMULKEIWICZ, R. & KIRKPATRICK, M. (1992). Quantitative genetics and the evolution of
765 reaction norms. *Evolution*, **46**, 390-411.

- 766 HANSEN, T. F., CARTER, A. J. R. & PÉLABON, C. (2006). On adaptive accuracy and precision in
767 natural populations. *The American Naturalist*, **168**, 168–81.
- 768 HILL, W. G. & MULDER, H. A. (2010). Genetic analysis of environmental variation. *Genetics*
769 *Research*, **92**, 381-395.
- 770 HUGIE, D. M. (2003). The waiting game: a "battle of waits" between predator and prey.
771 *Behavioral Ecology*, **14**, 807-817.
- 772 JACKSON, R. R. & WILCOX, R. S. (1993). Spider flexibly chooses aggressive mimicry signals for
773 different prey by trial and error. *Behaviour*, **127**, 21-36.
- 774 JOHNSTONE, R. A. (2001). Eavesdropping and animal conflict. *Proceedings of the National*
775 *Academy of Sciences USA*, **98**, 9177–9180.
- 776 KAGAN, M. L., NOVOPLANSKY, N. & SACHS, T. (1992). Variable cell lineages from the functional
777 pea epidermis. *Annals of Botany*, **69**, 303-312.
- 778 KELLNER, J. R. & ALFORD, R. A. (2003). The ontogeny of fluctuating asymmetry. *The American*
779 *Naturalist*, **161**, 931-947.
- 780 KINGSOLVER, J. G., SHLICHTA, J. G., RAGLAND, G. J. & MASSIE, K. R. (2006). Thermal reaction
781 norms for caterpillar growth depend on diet. *Evolutionary Ecology Research*, **8**, 703–715.
- 782 KONTIAINEN, P., PIETIÄINEN, H., HUTTUNEN, K., KARELL, P., KOLUNEN, H. & BROMMER, J. E.
783 (2009). Aggressive Ural owl mothers recruit more offspring. *Behavioral Ecology*, **20**, 789-
784 796.
- 785 KUSSEL, E. & LEIBLER, S. (2005). Phenotypic diversity, population growth, and information in
786 fluctuating environments. *Science*, **309**, 2075–2078.
- 787 LEE, Y. & NELDER, J. A. (1996). Hierarchical generalized linear models (with Discussion).
788 *Journal of the Royal Statistical Society B*, **58**, 619-678.

- 789 LINDHOLM, A. & THOMAS, R. (2000). Differences between populations of reed warblers in
790 defences against brood parasitism. *Behaviour*, **137**, 25–42.
- 791 LOSICK, R. & DESPLAN, C. (2008). Stochasticity and cell fate. *Science*, **320**, 65–68.
- 792 LOTEM, A., NAKAMURA, H. & ZAHAVI, A. (1995). Constraints on egg discrimination and cuckoo-
793 host co-evolution. *Animal Behaviour*, **49**, 1185-1209.
- 794 LYNCH, M. & WALSH, B. (1998). *Genetics and Analysis of Quantitative Traits*. Sinauer Press,
795 Sunderland, MA.
- 796 MARKOW, T. A. (1995). Evolutionary ecology and developmental instability. *Annual Reviews of*
797 *Entomology*, **40**, 105-120.
- 798 MARTIN, T. M., TON, R. & NIKLISON, A. (2013). Intrinsic versus extrinsic influences on life
799 history expression: metabolism and parentally induced temperature influences on embryo
800 development rate. *Ecology Letters*, **16**, 738-745.
- 801 MARTIN, J. G. A., NUSSEY, D., WILSON, A. & RÉALE, D. (2011). Measuring individual differences
802 in reaction norms in field and experimental studies: a power analysis of random regression
803 models. *Methods in Ecology and Evolution*, **2**, 362-374.
- 804 MATHOT, K. J., WRIGHT, J., KEMPENAERS, B. & DINGEMANSE, N. J. (2012). Adaptive strategies
805 for managing uncertainty may explain personality-related differences in behavioural
806 plasticity. *Oikos*, **121**, 1009–1020.
- 807 MAYNARD SMITH, J. (1974). Theory of games and the evolution of animal contests. *Journal of*
808 *Theoretical Biology*, **47**, 209-221.
- 809 MAYR, E. (1963). *Animal species and evolution*. Belknap Press/Harvard University Press,
810 Cambridge, MA.

- 811 MOCZEK, A. P., HUNT, J., EMLEN, D. J. & SIMMONS, L. W. (2002). Threshold evolution in exotic
812 populations of a polyphonic beetle. *Evolutionary Ecology Research*, **4**, 587–601.
- 813 MOORE, A. J., BRODIE III, E. D. & WOLF, J. B. (1997). Interacting phenotypes and the
814 evolutionary process: I. Direct and indirect genetic effects of social interactions. *Evolution*,
815 **51**, 1352-1362.
- 816 MORAN, N. A. (1992). The evolutionary maintenance of alternative phenotypes. *The American*
817 *Naturalist*, **139**, 971–989.
- 818 MÜLLER, M. S., ROELOFS, Y., ERIKSTAD, K. E. & GROOTHUIS, T. G. G. (2012). Maternal
819 androgens increase sibling aggression, dominance, and competitive ability in the siblicidal
820 black-legged kittiwake (*Rissa tridactyla*). *PLoS ONE*, 7(10), e47763.
821 doi:10.1371/journal.pone.0047763
- 822 NATARAJAN, D., DEVRIES, H., SAALTINK, D. J., DE BOER, S. F. & KOOLHAAS, J. (2009).
823 Delineation of violence from functional aggression in mice: an ethological approach.
824 *Behavioral Genetics*, **39**, 73-90.
- 825 NESSE, R. M. (2001). *Evolution and the Capacity for Commitment*. Russell Sage Foundation,
826 New York, NY.
- 827 NICOLAUS, M., BROMMER, J. E., UBELS, R., TINBERGEN, J. M. & DINGEMANSE, N. J. (2013).
828 Exploring patterns of variation in clutch size-density reaction norms in a wild passerine bird.
829 *Journal of Evolutionary Biology*, **26**, 2031-2043.
- 830 NIEMELÄ, P. T. & DINGEMANSE, N. J. (2014). Artificial environments and the study of "adaptive"
831 personalities. *Trends in Ecology and Evolution*, **29**, 245-247.

- 832 NIJHOUT, H. F. & DAVIDOWITZ, G. (2003). Developmental perspectives on phenotypic plasticity,
833 canalization, and fluctuating asymmetry. In: *Developmental Instability: Causes and*
834 *Consequences* (ed. M. POLAK) , pp. 3-13. MIT Press, Boston, MA.
- 835 NUSSEY, D. H., WILSON, A. J. & BROMMER, J. E. (2007). The evolutionary ecology of individual
836 phenotypic plasticity in wild populations. *Journal of Evolutionary Biology*, **20**, 831–44.
- 837 PICHE, J., HUTCHINGS, J. A. & BLANCHARD, W. (2008). Genetic variation in threshold reaction
838 norms for alternative reproductive tactics in male Atlantic salmon, *Salmo salar*. *Proceedings*
839 *of the Royal Society of London B*, **275**, 1571-1575.
- 840 PIERSMA, T. & DRENT, J. (2003). Phenotypic flexibility and the evolution of organismal design.
841 *Trends in Ecology and Evolution*, **18**, 228–233.
- 842 PLAYFAIR, J. H. L. (1995). *Infection and Immunity*, 2nd ed. Oxford University Press, New York,
843 NY.
- 844 RATIKAINEN, I. I., WRIGHT, J. & KAZEM, A. J. N. (2010). Social class influences degree of
845 variance sensitivity in wild Siberian jays. *Behavioral Ecology*, **21**, 1067-1072.
- 846 REEVE, H. K. (1989). The evolution of conspecific acceptance thresholds. *The American*
847 *Naturalist*, **133**, 407–435.
- 848 ROSS, H. E. & MURRAY, D. J. (1996). *E. H. Weber on the tactile senses*, 2nd ed. Erlbaum, Taylor
849 & Francis, Hove, UK.
- 850 ROTHSTEIN, S. I. (1978). Mechanisms of avian egg-recognition: Additional evidence for learned
851 components. *Animal Behaviour*, **26**, 671–677.
- 852 ROWE, L., LUDWIG, D. & SCHLUTER, D. (1994). Time, condition, and the seasonal decline of
853 avian clutch size. *The American Naturalist*, **143**, 698–722.

- 854 SCHAEFFER, L. R. (2004). Application of random regression models in animal breeding.
855 *Livestock Production Science*, **86**, 35–45.
- 856 SCHEINER, S. M. (2006). Genotype-environment interactions and evolution. In: *Evolutionary*
857 *genetics: concepts and case studies* (eds. C. W. FOX & J. B. WOLF) , pp. 326–338. Oxford
858 University Press, New York, NY.
- 859 SCHUETT, W., TREGENZA, T. & DALL, S. R. X. (2010). Sexual selection and animal personality.
860 *Biological Reviews*, **85**, 217-246.
- 861 SHAFIR, S. (2000). Risk-sensitive foraging: the effect of relative variability. *Oikos*, **88**, 663–669.
- 862 SHERMAN, P. W., REEVE, H. K. & PFENNIG, D. W. (1997). Recognition systems. In: *Behavioural*
863 *Ecology: An Evolutionary Approach*. Fourth edition. (eds J. R. KREBS & N. B. DAVIES), pp.
864 69–96. Blackwell Scientific, Oxford, UK.
- 865 SIMONS, A. M. (2011). Modes of response to environmental change and the elusive empirical
866 evidence for bet-hedging. *Proceedings of the Royal Society of London B*, **278**, 1601-1609.
- 867 SMITH, H. R., ROWSON, M., BASKETTER, D. A. & MCFADDEN, J. P. (2004). Intra-individual
868 variation of irritant threshold and relationship to trans-epidermal water loss measurement of
869 skin irritation. *Contact Dermatitis*, **51**, 26-29.
- 870 SMYTH, G. K. & VERBYLA, A. P. (1999). Adjusted likelihood methods for modeling dispersion in
871 generalized linear models. *Environmetrics*, **10**, 695-710.
- 872 SNELL-ROOD, E. C. (2013). An overview of the evolutionary causes and consequences of
873 behavioural plasticity. *Animal Behaviour*, **85**, 1004-1011.
- 874 STAMPS, J. A., BRIFFA, M. & BIRO, P. A. (2012). Unpredictable animals: individual differences in
875 intraindividual variability (IIV). *Animal Behaviour*, **83**, 1325–1334.

- 876 STAMPS, J. A. & GROOTHUIS, T. G. G. (2010). Developmental perspectives on personality:
877 implications for ecological and evolutionary studies of individual differences. *Philosophical*
878 *Transactions of the Royal Society B*, **365**, 4029-4041.
- 879 STEARNS, S. C. & KAWECKI, T. J. (1994). Fitness sensitivity and the canalization of life-history
880 traits. *Evolution*, **48**, 1438-1450.
- 881 STEPHENS, D. W. (1981). The logic of risk-sensitive foraging preferences. *Animal Behaviour*, **29**,
882 628–629.
- 883 STILLWELL, R. C., WALLIN, W. G., HITCHCOCK, L. J. & FOX, C. W. (2007). Phenotypic plasticity
884 in a complex world: interactive effects of food and temperature on fitness components of a
885 seed beetle. *Oecologia*, **153**, 309–321.
- 886 STOKKE, B. G., HAFSTAD, I., RUDOLFSEN, G., MOKSNES, A., MØLLER, A. P., ROSKRAFT, E. &
887 SOLER, M. (2008). Predictors of resistance to brood parasitism within and among reed
888 warbler populations. *Behavioral Ecology*, **19**, 612–620.
- 889 TONSOR, S. J., ELNACCASH, T. W. & SCHEINER, S. M. (2013). Developmental instability is
890 genetically correlated with phenotypic plasticity, constraining heritability, and fitness.
891 *Evolution*, **67**, 2923-2935.
- 892 VAN DE POL, M. (2012). Quantifying individual variation in reaction norms: how study design
893 affects the accuracy, precision and power of random regression models. *Methods in Ecology*
894 *and Evolution*, **3**, 268-280.
- 895 VAN DOOREN, T. J. M. (2001). Reaction norms with bifurcations shaped by evolution.
896 *Proceedings of the Royal Society of London B*, **268**, 279–287.
- 897 VAN VALEN, L. (1962). A study of fluctuating asymmetry. *Evolution*, **16**, 125–142.

- 898 VIA, S. & LANDE, R. (1985). Genotype-environment interaction and the evolution of phenotypic
899 plasticity. *Evolution*, **39**, 505-522.
- 900 VISWANATHAN, M. (2005). *Measurement error and research design*. Sage Publications,
901 Thousand Oaks, CA.
- 902 WADDINGTON, C. H. (1942). Canalization of development and the inheritance of acquired
903 characters. *Nature*, **150**, 563-565
- 904 WEINIG C. & DELPH, L. F. (2001). Phenotypic plasticity early in life constrains developmental
905 responses later. *Evolution*, **55**, 930-936.
- 906 WEST-EBERHARD, M. J. (2003). *Developmental plasticity and evolution*. Oxford University
907 Press, Oxford, UK.
- 908 WESTNEAT, D. F., HATCH, M. I., WETZEL, D. P. & ENSMINGER, A. L. (2011). Individual variation
909 in parental care reaction norms: integration of personality and plasticity. *The American*
910 *Naturalist*, **178**, 652–667.
- 911 WESTNEAT, D. F., SCHOFIELD, M. & WRIGHT, J. (2013). Parental behavior exhibits between-
912 individual variance, plasticity and heterogeneous residual variance. *Behavioral Ecology*, **24**,
913 598-604.
- 914 WESTNEAT, D. F., STEWART, I. R. K. & HATCH, M. I. (2009). Complex interactions among
915 temporal variables affect the plasticity of clutch size in a multi-brooded bird. *Ecology*, **90**,
916 1162–1174.
- 917 WHITE, H. (1980). A heteroskedasticity-consistent covariance matrix estimator and a direct test
918 for heteroskedasticity. *Econometrica*, **48**, 817–838.
- 919 WHITEN, A. & BYRNE, R.W. (1997). *Machiavellian Intelligence II*. Cambridge University Press,
920 Cambridge, UK.

- 921 WILEY, R. H. (1994). Errors, exaggeration and deception in animal communication. In:
922 *Behavioural Mechanisms in Evolutionary Ecology* (ed. L. A. REAL), pp. 157–189. University
923 of Chicago Press, Chicago, IL.
- 924 WILSON, A. J., RÉALE, D., CLEMENTS, M. N., MORRISSEY, M. M., POSTMA, E., WALLING, C. A.,
925 KRUUK, L. E. B. & NUSSEY, D. H. (2008). An ecologist's guide to the animal model. *Journal*
926 *of Animal Ecology*, **79**, 13–26.
- 927 WOLLERMAN, L. & WILEY, R. H. (2002). Background noise from a natural chorus alters female
928 discrimination of male calls in a Neotropical frog. *Animal Behaviour*, **63**, 15–22.
- 929 WOLTERECK, R. (1909). Weitere experimentelle Untersuchungen uber Artveränderung, speziell
930 uber das Wesen quantitativer Artunterschiede bei Daphniden. *Versuche Deutsche Zoologische*
931 *Gesellschaft*, **19**, 110–172.
- 932 YDENBERG, R. C. (1994). The behavioral ecology of provisioning in birds. *Ecoscience*, **1**, 1-14.

933 Table 1. Glossary of terms used in the text, a short definition, and related terms with the same or
 934 similar meaning.

935

Term	Definition	Similar terms
Active plasticity	Phenotypic plasticity in which the phenotype responds to environmental cues through a biological mechanism (<i>sensu</i> Scheiner, 2006)	Adaptive plasticity
Among-individual phenotypic variance	Variance among individuals in average phenotype in a specified environment	-
Canalization	The reduction of residual phenotypic variance at either the within-genotype-among-individual or within-individual levels	Developmental stability; behavioural stability; individual stability (Dingemans <i>et al.</i> , 2010; Stamps & Groothuis, 2010)
Developmental plasticity	Phenotypic plasticity occurring earlier in the lifetime that has long-lasting effects on the phenotype	Permanent environmental effect; irreversible plasticity (West-Eberhard, 2003)
Heterogeneous residual within-individual variance	Differences in residual within-individual variance across any terms in a model of phenotypic variance	Non-normal residual variance
Measurement error	Variance in phenotypic measures due to the way the trait is measured	Observer error
Multidimensional reaction norm	A function relating a phenotype to two or more environmental factors	
Organismal error	Variance in phenotype due to mismeasures of the environment by the subject	Phenotype–environment mismatching (DeWitt <i>et al.</i> , 1998); developmental instability (Waddington, 1942; Markow, 1995; Tonsor <i>et al.</i> , 2013); recognition error (Sherman <i>et al.</i> , 1997) or imprecision (Hansen <i>et al.</i> , 2006)
Passive plasticity	Phenotypic plasticity in which the effect of the environment can	Non-adaptive plasticity

	be explained by non-biological processes (<i>sensu</i> Scheiner, 2006)	
Phenotypic plasticity	A change in the phenotype expressed by a genotype or individual with respect to a difference in environment, either passive or active plasticity	Plasticity; flexibility
Residual within-individual variance	Amount of within-individual variance not explained in a specific statistical model (i.e. the average squared deviations of observations from an individual's reaction norm), averaged over a sample of individuals	Unexplained within-individual variance
Within-genotype among-individual variance	Variance in mean phenotype among individuals of a given genotype, measured in a specified environment	Among-individual variation
Within-individual plasticity	Variation in an individual's phenotype with respect to variation in the environment. Quantified at the individual level or averaged across individuals ("population average")	Reversible plasticity; behavioural flexibility (Piersma & Drent, 2003); activational plasticity (Snell-Rood, 2013); labile phenotype
Within-individual variance	Amount of phenotypic variance among instances of phenotypic expression of an individual. Quantified at the individual level or averaged across individuals ("population average")	Intra-individual variation (Stamps <i>et al.</i> , 2012)

936

937

938 Table 2. Examples of traits exhibiting patterns of residual variance that differ from that expected
 939 under passive plasticity. Such deviations have been suggested to be adaptive *via* the listed
 940 selective agent.

Trait exhibiting adaptive residual variance	Selective agent	Reference
Gene expression	Stochasticity leads to more efficient coregulation	Cai <i>et al.</i> (2008)
B-cells (antibody types)	Diversity followed by internal selection leads to more effective adaptive immunity	Golub & Green (1991)
Components of neural networks	Diversity followed by self-selection leads to more finely tuned neural processing	Changeux & Danchin (1976); Kagan <i>et al.</i> (1992)
Homeostatic temperature control	Multiple mechanisms across endotherms and ectotherms reduce variation leading to more effective physiological functions	
Task roles	Reduced variance leads to more effective output of social group	Bergmüller & Taborsky (2010)
Male courtship	Stereotyped and predictable courtship may be favoured through female preference	Schuett <i>et al.</i> (2010)
Prey responses	Variable and unpredictable emergence from refuge reduces predation	Hugie (2003); Briffa (2013)
Aggression levels in consecutive contests	Reduced variability increases ability to assess outcome and reduce costs to both contestants	Johnstone (2001)
Food intake rate (individual or provisioning parent)	Reduced variability beneficial to forager in high condition; increased variability beneficial to forager in poor condition	Stephens (1981); Ydenberg (1994)
Trial-and-error learning	Increased diversity of solutions, followed by self-selection, may lead to novel solutions to common problems	Frank (1997)

941

942

943 **Fig. 1.** Schematic representation of the hierarchical organization of phenotypic variance, with
944 directional arrows indicating that replicates of the next level (e.g. populations within species,
945 individuals within genotypes) are nested within the upper level. Variance in trait expression
946 among instances (i.e., within-individual variance) is relatively poorly studied, and so we focus on
947 phenotypes that have multiple instances of expression within an individual. We explore
948 processes that produce patterns of variance among instances. We also emphasize that variation in
949 patterns of variation can occur due to the hierarchical structure. That is, patterns of variation in
950 expression among instances can vary among individuals, genotypes, populations, etc.

951

952 **Fig. 2.** Plot of phenotypic measures (Y_{i1}) taken from a single individual ($j=1$) across an
953 environmental gradient (E_{i1}). The mean phenotype (β_{01}) is the elevation and is appropriately
954 taken at the mean-centred environment, and the slope (β_{11}) describes the individual's plasticity,
955 with elevation and slope together producing a norm of reaction. In this case there is
956 heterogeneous residual variance, with confidence limits indicated by the dashed lines that 'fan
957 out' over the gradient.

958 **Fig. 3.** Two examples of incomplete models producing heterogeneous within-individual residual
959 variance. (A) Modelling a phenotype with a linear reaction norm (solid line) produces
960 heterogeneous residuals when the reaction norm is actually non-linear (dashed line). (B)
961 Individuals (one in red, the other in blue) vary in how they respond to changes in the
962 environmental gradient (e.g. $I \times E$) and slope covaries with intercept. Omission of these terms
963 from the model will produce heterogeneous residual within-individual variance (if each is

964 assumed to have the average reaction norm, black dotted line). The vertical line indicates the
965 mean environment for E_1 .

966 **Fig. 4.** Multidimensional reaction norm depicted in two dimensions: gradient E_{1i1} (x -axis)
967 interacts with gradient E_{2i1} (indicated by colour) to affect the phenotype of an individual. This
968 non-additive effect of two different environmental parameters creates heterogeneous residual
969 within-individual variance if it is not modelled.

970 **Fig. 5.** Graphical depiction of the extended phenotypic equation applied to hypothetical data
971 from two individuals. The solid black line represents the population-average reaction norm. The
972 two individuals deviate from the population intercept (blue = ind_{01} and red = ind_{02}) and they
973 differ in slopes (blue line, $ind_{11} < red\ line, ind_{12}$). Individual 1, with the larger intercept, also has
974 a shallower slope, hinting at a negative covariance between intercept and slope. The two
975 individuals also differ in residual variance ($ind_{\sigma 01} < ind_{\sigma 02}$), indicated by the spread of points at
976 the intercept. Finally, the residual variance changes with E_{ij} differently for the two individuals
977 ($ind_{\sigma 11} < ind_{\sigma 12}$) and the change is positively correlated with individual residual variance
978 [$Cov(ind_{0j}, ind_{\sigma 1j}) > 0$]. Moreover, the individual with the smaller intercept has the larger residual
979 variance, indicating a negative covariance across levels [$Cov(ind_{0j}, ind_{\sigma 0j}) < 0$].

Figure 1.

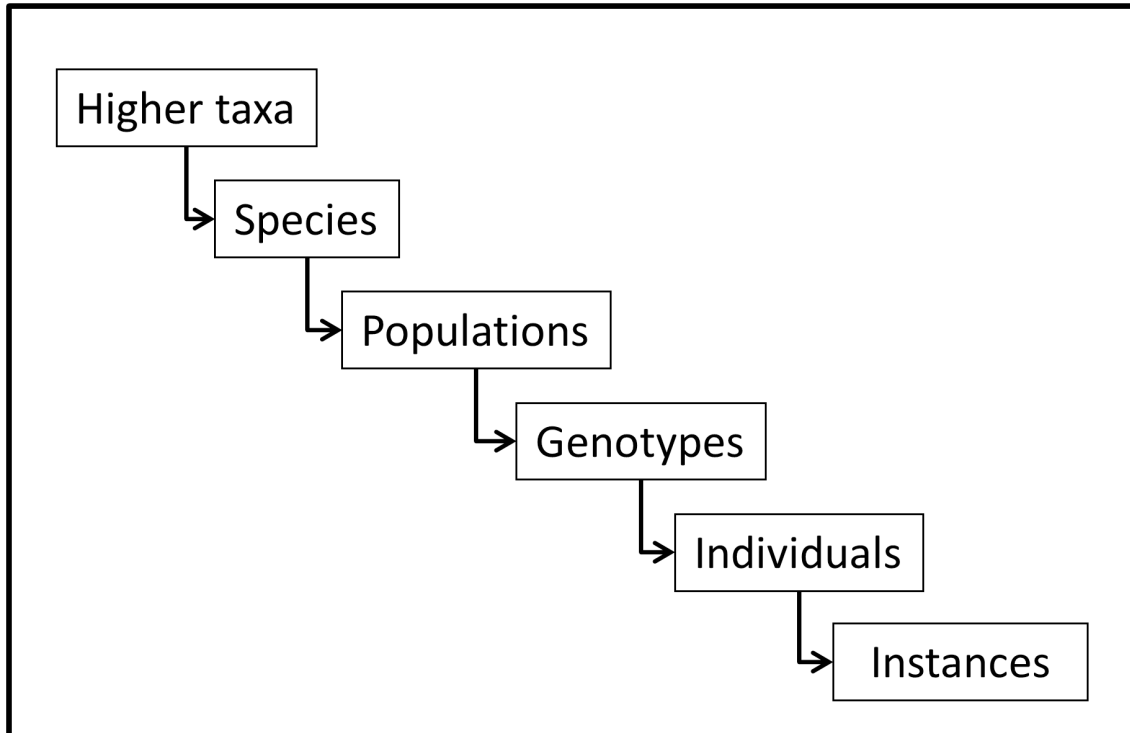


Figure 2.

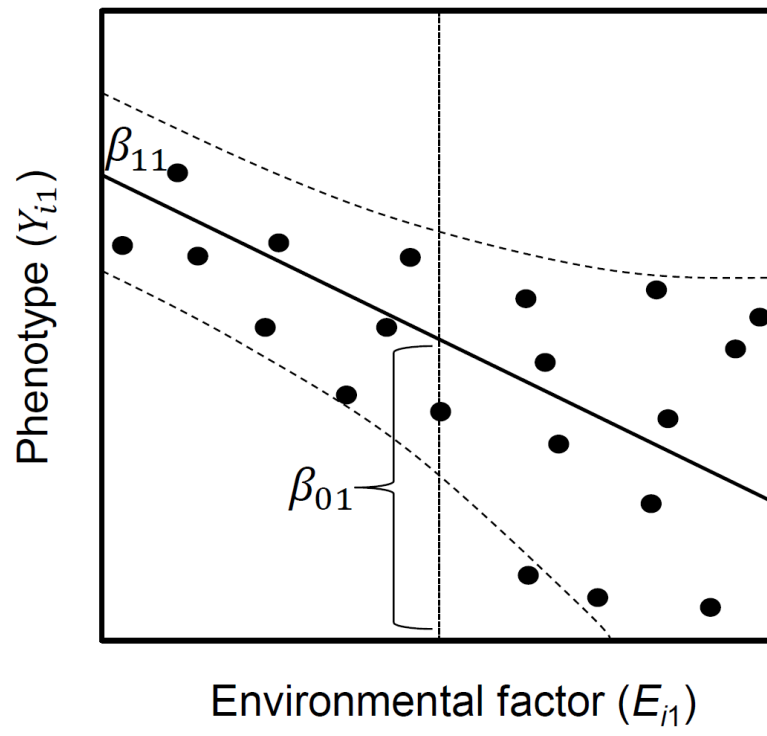


Figure 3.

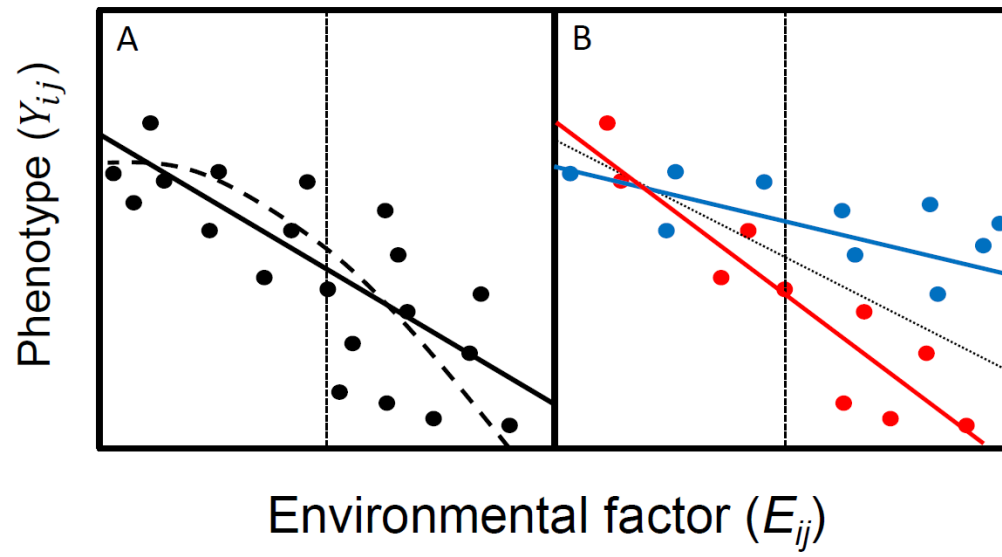


Figure 4.

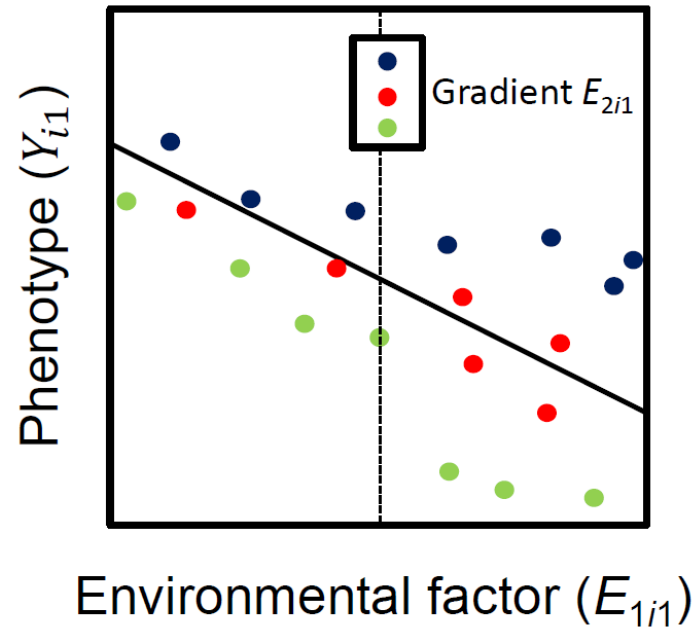


Figure 5.

