

1 **Decreased adiponectin/leptin ratio relates to insulin resistance in adults with obesity**

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26 **Running title:** Adiponectin/leptin relates to HOMA-IR

27 **Abstract**

28 Adipose tissue dysfunction is a key mechanism that leads to adiposity-based chronic disease. This study
29 aimed to investigate the feasibility of the adiponectin/leptin ratio (AdipoQ/Lep) as an adipose tissue and
30 metabolic function biomarker in adults with obesity, without diabetes. Data were collected from a
31 clinical trial conducted in 28 adults with obesity (mean body mass index: 35.4 ± 3.7 kg/m²)
32 (NCT02169778). Using a forward stepwise multiple linear regression model to explore the relationship
33 between AdipoQ/Lep and HOMA-IR, it was observed that 48.6% of HOMA-IR variance was explained by
34 triacylglycerols, AdipoQ/Lep and waist-to-hip ratio ($P < 0.001$), being AdipoQ/Lep the strongest
35 independent predictor (Beta = -0.449, $P < 0.001$). A lower AdipoQ/Lep was correlated with a higher body
36 mass index ($R_s = -0.490$, $P < 0.001$), body fat mass ($R_s = -0.486$, $P < 0.001$), waist-to-height ratio ($R_s = -0.290$,
37 $P = 0.037$), and plasma resistin ($R_s = -0.365$, $P = 0.009$). These data highlight the central role of adipocyte
38 dysfunction in the pathogenesis of insulin resistance and emphasize that AdipoQ/Lep may be a
39 promising early marker of insulin resistance development in adults with obesity.

40 **Keywords:** adiponectin/leptin; adipose tissue dysfunction; insulin resistance; metabolic dysfunction;
41 obesity

42

43 **Significance Statement**

44 Adiponectin/leptin ratio, triacylglycerols and waist-to-hip ratio explained almost half of HOMA-IR
45 variance in the context of obesity. This study provides evidence to support adipose tissue dysfunction
46 as a central feature of the pathophysiology of obesity and insulin resistance. Early identification of
47 individuals at higher risk of developing metabolic complications through adipose tissue dysfunction
48 assessment and the staging of obesity and its transient phenotypes can contribute to improve the
49 therapeutic decision-making.

50

51 **Introduction**

52 Adiposity-based chronic disease development and severity are directly related to changes in adipose
53 tissue composition and function (1). Excess adiposity is accompanied by increased production of pro-
54 inflammatory adipokines, such as leptin and resistin, whereas the production of anti-inflammatory
55 cytokines, such as adiponectin, is reduced (2). Dysregulated production or secretion of these adipokines
56 in response to adipose tissue dysfunction can contribute to chronic low-grade inflammation,
57 dyslipidemia, and metabolic impairment (3). Even though there is still no specific parameter for
58 assessing adipose tissue function, the adiponectin/leptin ratio (AdipoQ/Lep) has been proposed as a
59 promising biomarker (4).

60 The hypothesis that adipokines collectively are a major culprit in the development of insulin resistance
61 and metabolic dysfunction in obesity (“adipokine hypothesis”) received widespread attention (5, 6).
62 Despite its association with insulin signaling is well-accepted, the relative contribution of adipose tissue
63 dysfunction in explaining peripheral insulin resistance remains controversial. Thus, this study aimed to
64 analyze the feasibility of the AdipoQ/Lep as an adipose tissue and metabolic function biomarker based
65 on the “adipokine hypothesis”.

66

67 **Materials and methods**

68 Data were collected at two different moments from a clinical trial conducted in 28 adults (79% women;
69 mean age: 39.3±9.8 years) with obesity [mean body mass index (BMI): 35.4±3.7 kg/m²] (NCT02169778).
70 Individuals with other clinically significant comorbidities, including diabetes and cardiovascular diseases,
71 or those who had weight loss surgery were excluded. The detailed study protocol and the clinical
72 characteristics of the participants are published elsewhere (7).

73 The study was approved by the local Regional Ethics Committee (Midt-Norway, Trondheim, Norway)
74 and conducted according to the guidelines laid down in the Declaration of Helsinki. All participants
75 voluntarily agreed to participate in the study and provided their written informed consent.

76 Anthropometric measurements were assessed using standard reference method procedures and are
77 detailed elsewhere (7). Blood samples were collected after an overnight fast and plasma was
78 subsequently separated by centrifugation and stored at -80°C until further analysis. Biochemical
79 parameters were evaluated by a certified laboratory (St. Olavs University Hospital, Trondheim, Norway).
80 Plasma adipokines and inflammatory cytokines were measured through multiplex bead-based flow
81 cytometric immunoassays, as previously described (8). The AdipoQ/Lep was calculated with adiponectin
82 concentration expressed in µg/mL and leptin levels in ng/mL.

83

84 *Statistical analysis*

85 Correlations between the AdipoQ/Lep and clinical parameters were analyzed by the Spearman
86 correlation test (R_s) due to the non-normal distribution of the variables. A forward stepwise multiple
87 linear regression analysis was conducted to identify possible predictors of HOMA-IR and all the variables
88 that significantly correlated with the dependent variable (HOMA-IR) in the univariate analysis were
89 considered as independent variables, namely BMI, fat mass, waist circumference, hip circumference,
90 waist-to-hip ratio, waist-to-height ratio, total cholesterol, triacylglycerols, LDL cholesterol, and
91 AdipoQ/Lep. The overall proportion of variance explained by linear models was calculated using
92 adjusted R^2 measures. Statistical analyses were performed using SPSS Statistics version 27.0 and the
93 results were considered significant when $P < 0.05$.

94

95 **Results**

96 As outlined on Table 1, there were significant correlations between the AdipoQ/Lep and parameters
97 defining body composition and biomarkers of obesity. The AdipoQ/Lep was negatively correlated with
98 weight ($R_s = -0.338$, $P=0.014$), BMI ($R_s = -0.490$, $P<0.001$), and fat mass (in kg) ($R_s = -0.486$, $P<0.001$),
99 while it was positively correlated with the percentage of fat-free mass ($R_s = 0.515$, $P<0.001$).
100 Furthermore, a negative correlation between AdipoQ/Lep and waist-to-height ratio ($R_s = -0.290$,

101 $P=0.037$) was observed. This indicates that a dysfunctional adipose tissue, evidenced by a lower
102 AdipoQ/Lep, is related with a phenotype of individuals with central adiposity.
103 Regarding adipose tissue function, the AdipoQ/Lep was negatively correlated with resistin ($R_s = -0.365$,
104 $P=0.009$), which is an adipose tissue-specific secretory factor. Since resistin appears to be involved in
105 the development of insulin resistance (9, 10), the correlation between the AdipoQ/Lep and surrogate
106 markers of insulin resistance and sensitivity was evaluated. The AdipoQ/Lep was negatively correlated
107 with fasting plasma insulin levels ($R_s = -0.625$, $P<0.001$) and HOMA-IR ($R_s = -0.613$, $P<0.001$). In fact, the
108 AdipoQ/Lep had a stronger correlation with HOMA-IR than HOMA-IR with adiponectin ($R_s = -0.304$,
109 $P=0.033$) or leptin ($R_s = 0.422$, $P=0.003$) separately (Table 1). Moreover, a positive correlation was
110 observed between the AdipoQ/Lep and both HOMA- β ($R_s = 0.591$, $P<0.001$) and HOMA-S ($R_s = 0.596$,
111 $P<0.001$) (Table 2). After adjusting for age, sex, BMI, fat mass (in kg), and waist circumference the
112 correlation between AdipoQ/Lep and HOMA-IR ($R_s = -0.469$, $P=0.002$), HOMA- β ($R_s = 0.406$, $P=0.006$),
113 and HOMA-S ($R_s = 0.535$, $P<0.001$) remained significant (Table 2). No further significant correlations
114 were found between the AdipoQ/Lep and lipid metabolism or inflammatory cytokines (Table 1).
115 Given that AdipoQ/Lep correlated with key features of dysmetabolic obesity, the contribution of adipose
116 tissue function to insulin resistance was evaluated. A multiple linear regression analysis was performed
117 in order to evaluate if AdipoQ/Lep was an independent predictor and could explain the changes in
118 HOMA-IR. Forward stepwise regression analysis revealed that 48.6% of HOMA-IR variance was
119 explained by variations in triacylglycerols, AdipoQ/Lep, and waist-to-hip ratio ($P<0.001$), being
120 AdipoQ/Lep the strongest independent predictor (Beta = -0.449, $P<0.001$) (Table 3).

121

122 Discussion

123 Adipose tissue dysfunction may lead to alterations in adipokine secretion profile, namely adiponectin
124 and leptin, supporting an environment conducive to insulin resistance (2).

125 The main finding of this study was that AdipoQ/Lep may be considered a predictive marker of peripheral
126 insulin resistance in adults with obesity since this marker along with triacylglycerols and waist-to-hip
127 ratio explained nearly half of HOMA-IR variance independently of BMI, fat mass, and waist
128 circumference. Interestingly, this result suggests that different characteristics associated with adiposity
129 (i.e., total amount, distribution, and function of adipose tissue) may be behind the variance of HOMA-
130 IR in the context of obesity, rather than a single component. To some extent, this is in line with the
131 adiposity-based chronic disease's definition (1) and strengthens the evidence that identifies adipose
132 tissue dysfunction as a determinant of obesity-associated metabolic complications.

133 Following the onset of obesity, adipose tissue dysfunction may contribute to local and peripheral insulin
134 resistance through autocrine effects of inflammatory adipose tissue-derived factors on insulin signaling
135 and metabolism in adipocytes, and endocrine effects of adipokines on insulin production and/or
136 sensitivity in other metabolically active organs, particularly in the pancreas, skeletal muscle, and liver
137 (2). For example, leptin up-regulates TNF- α and IL-6, which in turn, can promote insulin resistance by
138 multiple mechanisms, such as reducing the expression of glucose transporter-4 and insulin receptor
139 substrate-1 (2, 10), and inhibiting the production of adiponectin (11). On pancreatic β -cells these pro-
140 inflammatory adipokines and free fatty acids have cytotoxic effects that can lead to a decrease in insulin
141 production, exacerbating β -cells dysfunction, glucose intolerance, and increasing the susceptibility to
142 type 2 diabetes (11). It also bears mention that it is likely that adipokines' dynamic interplay underlies
143 the pathophysiology of insulin resistance rather than the effect of a single adipokine. The lack of
144 correlation between classical pro-inflammatory cytokines (e.g., TNF- α and IL-6) and AdipoQ/Lep or
145 HOMA-IR is noteworthy. Although likely explained by the low detection of these cytokines in plasma
146 samples, it may also suggest the potential involvement of other mechanisms independent of
147 inflammatory pathways, namely lipotoxicity related to the inability of adipose tissue to properly store
148 lipids (9, 12) or the imbalance in the modulation and crosstalk of the aquaporins (integral membrane

149 proteins) and the hepatokines, such as fibroblast growth factors, involved in the regulation of lipid-
150 related metabolism and oxidative stress (13–15, 17).

151 Global risk assessment, including assessing the functionality of adipose tissue, is of clinical interest and
152 may identify high-risk profiles for the development of insulin resistance. However, the efficacy of the
153 current techniques to evaluate obesity’s risk, including BMI, waist circumference, and waist-to-height
154 ratio has been debated because such measurements are unable to accurately evaluate adipose tissue
155 function (16). At individual level, there is no adiposity-adjusted biomarker that separates distinct
156 systemic metabolic phenotypes, for instance between insulin sensitive (healthy) and insulin resistant
157 (unhealthy) obesity phenotypes. Although adiponectin and leptin can be also detected (at lower levels)
158 in other tissues, such as the cardiomyocytes and stomach, respectively, their primarily source is adipose
159 tissue (18–20). Therefore, these adipokines emerge as an attractive candidate to identify these two
160 phenotypes since they are directly or indirectly associated with adipose tissue function (3).

161 This study demonstrated that AdipoQ/Lep was significantly correlated with both abnormal adiposity
162 mass and adipose tissue function characteristics, reinforcing its usefulness as an adipose tissue function
163 biomarker. A higher AdipoQ/Lep was associated with lower weight, BMI, body fat mass, waist-to-height
164 ratio, and plasma resistin. These results are consistent with earlier studies aimed to assess the
165 association between AdipoQ/Lep and metabolic parameters in adults with obesity (21–25).

166 Altogether, these findings appear to support the “adipokine hypothesis”, suggesting that altered
167 adipokine dynamics, as evidenced by a low AdipoQ/Lep, is a feasible marker of adipose tissue
168 dysfunction, and along with other characteristics of adiposity, is a central feature that explains insulin
169 resistance (HOMA-IR) in adults with obesity. Early identification of individuals at higher risk of
170 developing metabolic complications through adipose tissue dysfunction assessment, and the staging of
171 obesity and its transient phenotypes can contribute to improve the therapeutic decision-making. In an
172 era of personalized and precision nutrition, this might be a step forward to a more individualized
173 patient-centered approach that enables improved functional and prognostic assessment for individuals

174 affected by obesity. Although further randomized controlled trials powered for the effect of AdipoQ/Lep
175 on HOMA-IR are needed to confirm this hypothesis, these findings open new possibilities in future
176 research to study therapeutic strategies aimed at improving this dysfunctional adipokine secretion
177 profile.

178

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187

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191 **References**

- 192 1. **Frühbeck G, Busetto L, Dicker D, Yumuk V, Goossens GH, Hebebrand J, Halford JGC, Farpour-**
193 **Lambert NJ, Blaak EE, Woodward E, Toplak H.** The ABCD of obesity: An EASO position statement
194 on a diagnostic term with clinical and scientific implications. *Obes Facts* 12: 131–136, 2019. doi:
195 10.1159/000497124.
- 196 2. **Ouchi N, Parker JL, Lugus JJ, Walsh K.** Adipokines in inflammation and metabolic disease. *Nat Rev*
197 *Immunol* 11: 85–97, 2011. doi: 10.1038/nri2921.
- 198 3. **Assunção M, Guimarães JT, Faria M, Monteiro R.** Adipokines as Emerging Biomarkers for Adipose
199 Tissue Dysfunction. In: *Understanding Obesity: From its Causes to impact on Life*, edited by
200 Monteiro R, Martins MJ. 2020, p. 81–99.
- 201 4. **Frühbeck G, Catalán V, Rodríguez A, Gómez-Ambrosi J.** Adiponectin-leptin ratio: A promising index
202 to estimate adipose tissue dysfunction. Relation with obesity-associated cardiometabolic risk.
203 *Adipocyte* 7: 57–62, 2018. doi: 10.1080/21623945.2017.1402151.
- 204 5. **Chadt A, Scherneck S, Joost H-G, Al-Hasani H.** Molecular links between Obesity and Diabetes. In:
205 *Diabesity*, edited by Feingold KR, Anawalt B, Boyce A, Chrousos G, Herder WW de, Dhatariya K,
206 Dungan K, Hershman JM, Hofland J, Kalra S, Kaltsas G, Koch C, Kopp P, Korbonits M, Kovacs CS,
207 Kuohung W, Laferrère B, Levy M, McGee EA, McLachlan R, Morley JE, New M, Purnell J, Sahay R,
208 Singer F, Sperling MA, Stratakis CA, Trencle DL, Wilson DP. South Dartmouth (MA): Endotext
209 [Internet], 2000.
- 210 6. **Virtue S, Vidal-Puig A.** It's not how fat you are, it's what you do with it that counts. *PLoS Biol* 6:
211 1819–1823, 2008. doi: 10.1371/journal.pbio.0060237.
- 212 7. **Coutinho SR, Halset EH, Gåsbakk S, Rehfeld JF, Kulseng B, Truby H, Martins C.** Compensatory
213 mechanisms activated with intermittent energy restriction: A randomized control trial. *Clin Nutr*
214 37: 815–823, 2018. doi: 10.1016/j.clnu.2017.04.002.
- 215 8. **Castela I, Rodrigues C, Ismael S, Barreiros-Mota I, Morais J, Araújo JR, Marques C, Silvestre MP,**

- 216 **Ângelo-Dias M, Artins C, Borrego LM, Monteiro R, Coutinho SR, Calhau C, Faria A, Pestana D,**
217 **Martins C, Teixeira D.** Intermittent energy restriction ameliorates adipose tissue-associated
218 inflammation in adults with obesity: A randomised controlled trial. *Clin Nutr* 41: 1660–1666,
219 2022. doi: 10.1016/j.clnu.2022.06.021.
- 220 9. **De Ferranti S, Mozaffarian D.** The perfect storm: Obesity, adipocyte dysfunction, and metabolic
221 consequences. *Clin Chem* 54: 945–955, 2008. doi: 10.1373/clinchem.2007.100156.
- 222 10. **Antuna-Puente B, Feve B, Fellahi S, Bastard JP.** Adipokines: The missing link between insulin
223 resistance and obesity. *Diabetes Metab* 34: 2–11, 2008. doi: 10.1016/j.diabet.2007.09.004.
- 224 11. **Burhans MS, Hagman DK, Kuzma JN, Schmidt KA, Kratz M.** Contribution of adipose tissue
225 inflammation to the development of type 2 diabetes mellitus. *Compr Physiol* 9: 1–58, 2019. doi:
226 10.1002/cphy.c170040.
- 227 12. **Stern JH, Rutkowski JM, Scherer PE.** Adiponectin, Leptin, and Fatty Acids in the Maintenance of
228 Metabolic Homeostasis through Adipose Tissue Crosstalk. *Cell Metab* 23: 770–784, 2016. doi:
229 10.1016/j.cmet.2016.04.011.
- 230 13. **Frühbeck G.** Aquaporin enters the picture. *Nature* 24: 436–7, 2005.
- 231 14. **Izaguirre M, Gil MJ, Monreal I, Montecucco F, Frühbeck G, Catalán V.** The Role and Potential
232 Therapeutic Implications of the Fibroblast Growth Factors in Energy Balance and Type 2
233 Diabetes. *Curr Diab Rep* 17, 2017. doi: 10.1007/s11892-017-0866-3.
- 234 15. **Alvarez-sola G, Uriarte I, Latasa MU, Fernandez-barrena MG, Urtasun R, Elizalde M, Barcena-varela**
235 **M, Jiménez M, Chang HC, Barbero R, Catalán V, Rodríguez A, Frühbeck G, Gallego-escuredo JM,**
236 **Gavaldà-navarro A, Villarroya F, Rodríguez-ortigosa CM, Corrales FJ, Prieto J, Berraondo P, Berasain**
237 **C, Avila MA.** Fibroblast growth factor 15/19 (FGF15/19) protects from diet-induced hepatic
238 steatosis: development of an FGF19-based chimeric molecule to promote fatty liver
239 regeneration. *Gut* 66: 1818–1828, 2017. doi: 10.1136/gutjnl-2016-312975.
- 240 16. **Schrover IM, Spiering W, Leiner T, Visseren FLJ.** Adipose Tissue Dysfunction: Clinical Relevance

- 241 and Diagnostic Possibilities. *Horm Metab Res* 48: 213–225, 2016. doi: 10.1055/s-0042-103243.
- 242 17. **Domingo P, Moncada R, Giral M, Villarroya F, Salvador J.** FGF19 and FGF21 serum concentrations
243 in human obesity and type 2 diabetes behave differently after diet- or surgically-induced weight
244 loss. *Clin Nutr* 36: 861–868, 2017. doi: 10.1016/j.clnu.2016.04.027.
- 245 18. **Zhao S, Kusminski CM, Scherer PE.** Adiponectin, Leptin and Cardiovascular Disorders. *Circ Res*
246 128: 136–149, 2021. doi: 10.1161/CIRCRESAHA.120.314458.
- 247 19. **Sobhani I, Bado A, Vissuzaine C, Buyse M, Kermorgant S, Laigneau J, Attoub S, Lehy T, Henin D,**
248 **Mignon M, Lewin MJM, Paris H.** Leptin secretion and leptin receptor in the human stomach. *Gut*
249 47: 178–183, 2000.
- 250 20. **Muruzábal FJ, Frühbeck G, Gómez-Ambrosi J, Archanco M, Burrell MA.** Immunocytochemical
251 detection of leptin in non-mammalian vertebrate stomach. *Gen Comp Endocrinol* 128: 149–152,
252 2002. doi: 10.1016/s0016-6480(02)00072-2.
- 253 21. **Frühbeck G, Catalán V, Rodríguez A, Ramírez B, Becerril S, Salvador J, Colina I, Gómez-Ambrosi J.**
254 Adiponectin-leptin Ratio is a Functional Biomarker of Adipose Tissue Inflammation. *Nutrients* 11:
255 1–13, 2019. doi: 10.3390/nu11020454.
- 256 22. **Finucane FM, Luan J, Wareham NJ, Sharp SJ, O’Rahilly S, Balkau B, Flyvbjerg A, Walker M, Højlund**
257 **K, Nolan JJ, Savage DB.** Correlation of the leptin:adiponectin ratio with measures of insulin
258 resistance in non-diabetic individuals. *Diabetologia* 52: 2345–2349, 2009. doi: 10.1007/s00125-
259 009-1508-3.
- 260 23. **Frühbeck G, Catalán V, Rodríguez A, Ramírez B, Sal J, Portincasa P, Colina I, Gómez- J.** Involvement
261 of the leptin-adiponectin axis in inflammation and oxidative stress in the metabolic syndrome. :
262 1–8, 2017. doi: 10.1038/s41598-017-06997-0.
- 263 24. **Vega GL, Grundy SM.** Metabolic Risk Susceptibility in Men Is Partially Related to
264 Adiponectin/Leptin Ratio. *J Obes* 2013: 1–10, 2013. doi: 10.1155/2013/409679.
- 265 25. **Gupta V, Mishra S, Mishra S, Kumar S, Gupta V.** Association of Leptin:Adiponectin ratio and

266 metabolic risk markers in postmenopausal women. *Immunol Lett* 196: 63–67, 2018. doi:
267 10.1016/j.imlet.2018.01.008.
268
269

270 **Table 1** – Spearman correlations (R_s) between AdipoQ/Lep or HOMA-IR and anthropometric and metabolic
 271 parameters

	AdipoQ/Lep		HOMA-IR	
	R_s	P -value	R_s	P -value
<i>Anthropometric measures and body composition</i>				
Weight, kg	-0.338	0.014*	0.540	< 0.001**
BMI, kg/m ²	-0.490	< 0.001**	0.495	< 0.001**
FM, kg	-0.486	< 0.001**	0.462	< 0.001**
FM, %	-0.515	< 0.001**	0.245	0.090
FFM, kg	0.056	0.691	0.246	0.088
FFM, %	0.515	< 0.001**	-0.245	0.090
Waist circumference, cm	-0.191	0.175	0.513	< 0.001**
Hip circumference, cm	-0.464	< 0.001**	0.333	0.020*
Waist-to-hip ratio	0.113	0.426	0.402	0.004*
Waist-to-height ratio	-0.290	0.037*	0.504	< 0.001**
<i>Glucose homeostasis and insulin sensitivity</i>				
Glucose, mmol/L	-0.061	0.669	NA	NA
Insulin, pg/mL	-0.625	< 0.001**	NA	NA
<i>Blood lipid profile</i>				
Total cholesterol, mmol/L	-0.231	0.099	0.412	0.003*
Triacylglycerols, mmol/L	-0.194	0.168	0.425	0.002*
HDL cholesterol, mmol/L	-0.019	0.895	-0.084	0.564
LDL cholesterol, mmol/L	-0.188	0.183	0.336	0.018*
<i>Adipokines</i>				
Adiponectin, µg/mL	NA	NA	-0.304	0.033*
Adipsin, ng/mL	-0.180	0.226	0.120	0.437
Leptin, ng/mL	NA	NA	0.422	0.003*
Resistin, pg/mL	-0.365	0.009*	0.261	0.077
AdipoQ/Lep	NA	NA	-0.613	< 0.001**
<i>Inflammatory cytokines</i>				
IL-1β, pg/mL	0.073	0.655	-0.185	0.259
IFN-γ, pg/mL	0.025	0.880	-0.070	0.684
MCP-1, pg/mL	-0.014	0.923	0.031	0.835
IL-6, pg/mL	0.014	0.951	0.178	0.440
IL-8, pg/mL	-0.223	0.246	0.187	0.342
IL-10, pg/mL	0.038	0.851	-0.167	0.426
IL-17A, pg/mL	0.038	0.871	0.008	0.970
IL-18, pg/mL	-0.150	0.310	0.272	0.071
IL-23, pg/mL	-0.102	0.527	-0.013	0.939
IL-33, pg/mL	0.067	0.689	-0.010	0.955

272 Data are presented as Spearman's correlation coefficient and associated P -values: * P < 0.05 or ** P < 0.001.

273 Low-density lipoprotein (LDL) cholesterol was estimated by the Friedewald equation (19). Insulin resistance (HOMA-IR) was
 274 estimated by Homeostatic Model Assessment (HOMA) (20).

275 AdipoQ/Lep: adiponectin/leptin ratio; BMI: body mass index; FFM: fat-free mass; FM: fat mass; HDL: high-density lipoprotein
 276 cholesterol; HOMA-IR: homeostatic model assessment of insulin resistance; IFN: interferon; IL: interleukin; LDL: low-density
 277 lipoprotein cholesterol; MCP: monocyte chemoattractant protein; NA: not applicable.

278

279 **Table 2** – Spearman correlations (Rs) between AdipoQ/Lep and HOMA-IR, HOMA-β and HOMA-S, unadjusted and
 280 adjusted for age, sex, BMI, and fat mass (in kg), and waist circumference

	AdipoQ/Lep			
	Crude		Adjusted for age, sex, BMI, and fat mass (in kg), and waist circumference	
	Rs	P-value	Rs	P-value
HOMA-IR	-0.613	< 0.001**	-0.469	0.002*
HOMA-β, %	0.591	< 0.001**	0.406	0.006*
HOMA-S, %	0.596	< 0.001**	0.535	< 0.001**

281 Data are presented as Spearman's correlation coefficient and associated P-values: *P < 0.05 or **P < 0.001.

282 Insulin resistance (HOMA-IR) and sensitivity (HOMA-S) and β-cell function (HOMA-β) were estimated by Homeostatic Model
 283 Assessment (HOMA) (20).

284 AdipoQ/Lep: adiponectin/leptin ratio; BMI: body mass index; HOMA-β: homeostasis model assessment of β-cell function;
 285 HOMA-IR: homeostatic model assessment of insulin resistance; HOMA-S: homeostasis model assessment of insulin
 286 sensitivity.

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288 **Table 3** – Forward stepwise multiple linear regression analysis with HOMA-IR as the dependent variable

Independent variables	HOMA-IR				
	B	Beta	P-value	Adjusted R ²	P-value
Model 1					
Triacylglycerols, mmol/L	1.956	0.519	<0.001**	0.253	<0.001**
Model 2					
Triacylglycerols, mmol/L	1.618	0.429	<0.001**	0.387	<0.001**
AdipoQ/Lep	-0.918	-0.390	0.002*		
Model 3					
Triacylglycerols, mmol/L	0.962	0.255	0.043*	0.486	<0.001**
AdipoQ/Lep	-1.058	-0.449	<0.001**		
Waist-to-hip ratio	11.375	0.368	0.004*		

289 Data are presented as adjusted R² and associated P-value assessed by a forward stepwise multiple linear regression, adjusted
 290 for the variables that were significantly correlated with HOMA-IR.

291 Insulin resistance (HOMA-IR) was estimated by Homeostatic Model Assessment (HOMA) (20).

292 AdipoQ/Lep: adiponectin/leptin ratio; B: unstandardized regression coefficient; Beta: standardized beta coefficient; HOMA-IR:
 293 homeostatic model assessment of insulin resistance.

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