# The effects of nutritional state, sex and body size on the marine migration behaviour of sea trout 

S. H. Eldøy ${ }^{*}$, X. Bordeleau ${ }^{2,3}$, M. J. Lawrence ${ }^{4,5}$, E. B. Thorstad ${ }^{6}$, A. G. Finstad ${ }^{1}$, F. G. Whoriskey $^{7}$, G. T. Crossin ${ }^{2}$, S. J. Cooke ${ }^{4}$, K. Aarestrup ${ }^{8}$, L. Rønning ${ }^{1}$, A. D. Sjursen ${ }^{1}$ and J. G. Davidsen ${ }^{1}$.

${ }^{1}$ S. H. Eldøy (sindre.eldoy@ntnu.no), J. G. Davidsen (jan.davidsen@ntnu.no), A. G. Finstad (anders.finstad@ntnu.no), L. Rønning (lars.ronning@ntnu.no), A. D. Sjursen (aslak.sjursen@ntnu.no). Department of Natural History, NTNU University Museum, Norwegian University of Science and Technology, NO-7491 Trondheim, Norway.
${ }^{2}$ X. Bordeleau (Xavier.Bordeleau@dfo-mpo.gc.ca) and G. Crossin (glenn.crossin@dal.ca). Department of Biology, Dalhousie University, 1355 Oxford Street, Halifax, NS B3H 4R2, Canada.
${ }^{3}$ X. Bordeleau (Xavier.Bordeleau@,dfo-mpo.gc.ca). Maurice Lamontagne Institute, Department of Fisheries and Oceans Canada, 850 Route de la mer, Mont-Joli, QC G5H 3Z4, Canada.
${ }^{4}$ M. J. Lawrence (m_lawrence27@live.ca) and S. J. Cooke (StevenCooke@cunet.carleton.ca). Fish Ecology and Conservation Physiology Laboratory, Department of Biology, Carleton University, Ottawa, ON, K1S 5B6, Canada.

# ${ }^{5}$ M. J. Lawrence (m lawrence27@live.ca). Department of Biological Sciences, University of Manitoba, Sifton Rd, Winnipeg, MB R3T 2N2 

${ }^{6}$ E. B. Thorstad (eva.thorstad@nina.no). Norwegian Institute for Nature Research, NO-7485 Trondheim, Norway.

${ }^{7}$ F. G. Whoriskey (fwhoriskey@dal.ca). Ocean Tracking Network, Dalhousie University, 1355 Oxford St., Halifax, NS B3H 4J1, Canada.

${ }^{8} \mathrm{~K}$. Aarestrup (kaa@aqua.dtu.dk). National Institute of Aquatic Resources, Technical University of Denmark, DK-8600 Silkeborg, Denmark.

Running headline: Physiological drivers for marine behaviour in sea trout Salmo trutta.
*Corresponding author: Sindre Håvarstein Eldøy, Department of Natural History, NTNU University Museum, Norwegian University of Science and Technology, NO-7491 Trondheim; Norway. Tel.: +47 922 66586; email: sindre.eldoy@ntnu.no.


#### Abstract

The sea trout (anadromous brown trout, Salmo trutta) displays extensive among-individual variation in marine migration behaviour. We studied the migration behaviour of 286 sea trout (27-89 cm ) tagged with acoustic transmitters in the spring, in seven populations located in two distinct marine fjord systems in Norway. We examined whether individual nutritional state, sex, and body size influenced marine migration behaviour in terms of $i$. the decision to migrate to the sea or remain resident in freshwater and/or estuarine habitats, ii. seasonal


timing of sea entry, iii. duration of the marine residency, and $i v$. migration distance at sea from the home river. Most sea trout were in a poor nutritional state in the spring prior to migration. Sea trout with low body condition factors and low plasma triglyceride levels were more likely to migrate to sea, and low triglyceride levels were also associated with earlier sea-entry. Poor body condition also increased the probability of individuals remaining at sea longer and migrating further offshore compared to fish in better condition. Females were more likely to migrate to the sea than males. Larger fish were also more likely to migrate to sea instead of remaining in freshwater and estuaries and dispersed over greater distances from the river than smaller fish. In conclusion, this study documented general trends across multiple populations and showed that nutritional state, sex and body size influence important aspects of the marine migration behaviour of sea trout.

## 1. INTRODUCTION

Migration behaviour is observed in a wide range of taxa (Dingle 2014). There are various proximate explanations for why animals migrate, but the ultimate reason is for the optimization of individual growth and survival in order to increase lifetime fitness (Dingle \& Drake 2007). Throughout their lifetimes, individuals must continually allocate energy to various life-history activities, while balancing the metabolic demands for somatic growth, maturation, and reproduction (Zera \& Harshman 2001). Diadromy, which refers to migrations between marine and freshwater habitats, is thought to have evolved because of differences in food availability between these habitats (Gross et al. 1988). Among the fish family Salmonidae, all species spawn in freshwater, but many are anadromous, which means individuals migrate to sea at some point in their lives to exploit the richer marine food resources (Jonsson \& Jonsson 1993). Salmonid populations often consist of both freshwater
resident and sea migrating individuals (i.e partial migration, Chapman et al. 2012). Body size is positively correlated with fecundity within salmonids (Elliott 1995). Marine migration is advantageous if the fitness benefits of larger body size outweighs the cost of migration, including increased risk of mortality, disease and failure to reach spawning grounds (Klemetsen et al. 2003, Thorstad et al. 2016).

The brown trout (Salmo trutta) is a highly adaptable salmonid species, which through natural dispersal or human transport is found in all continents except Antarctica (MacCrimmon et al. 1970). Wide variation in environmental conditions and food availability influences the physiology of individuals, and further determine whether they migrate to sea (anadromous brown trout, hereafter referred to as sea trout) or remain freshwater resident (Forseth et al. 1999, Wysujack et al. 2009, Archer et al. 2020). After leaving freshwater, sea trout display plasticity in migratory tactics, with some individuals using nearshore and estuarine habitats, while others use marine areas more than 500 km away from their natal watercourse (Thorstad et al. 2016, Birnie-Gauvin et al. 2019b). However, variation in migration patterns and life history strategies is not fully understood, thus limiting our understanding of ecological and evolutionary dynamics of sea trout populations (BirnieGauvin et al. 2019b, Ferguson et al. 2019). Understanding the drivers of marine migration behaviour is crucial for evaluating susceptibility of sea trout to large scale climate change, and to human induced stressors that can vary both temporally and spatially in coastal zone ecosystems (Thorstad et al. 2015, Nevoux et al. 2019). In general, migration is regarded as a biological phenomenon that is particularly sensitive to environmental change and anthropogenic disturbance (Wilcove \& Wikelski 2008) such that it is important to understand how different taxa respond to such challenges (Lennox et al. 2016).

Energy status is known to impact migratory strategies of individual trout (Cucherousset et al. 2005, Boel et al. 2014, Bordeleau et al. 2018). For mature sea trout,
reproduction is energetically expensive (Lien 1978, Jonsson \& Jonsson 2005), and so they must recondition between spawning events. In watercourses suitable for overwintering, sea trout can remain in the spawning river system throughout the winter after autumn spawning (Berg \& Berg 1989, Östergren \& Rivinoja 2008), but in these oligotrophic systems feeding and growth are usually limited. Post-spawned individuals are therefore in a generally poor nutritional state prior to the seaward migration in the spring (Jonsson \& Jonsson 1998, Jonsson \& Jonsson 2011a). How individual variation in energy status relates to variation in marine migration behaviour of sea trout is not well understood. Body condition factor, which is based on the relationship between body length and mass (i.e., the relative stoutness of fish), is commonly used as an index of somatic energy status in salmonids and other fishes, but might not always be a precise predictor of energy status (Weatherley \& Gill 1983, Simpson 1992, Næsje et al. 2006). In addition to body condition factor, nutritional correlates derived from blood plasma samples can be used to assess the nutritional state of fishes. For salmonids, low levels of plasma triglycerides, total protein, and calcium levels can indicate poor nutritional state (Congleton \& Wagner 2006), which has previously been observed to promote marine migratory decisions in brown trout and Atlantic salmon (Boel et al. 2014, Bordeleau et al. 2018, Bordeleau et al. 2019). Elevated levels of cortisol possibly due to low food availability, have previously been found to promote earlier seaward migration (BirnieGauvin et al. 2019a). These previous studies suggest that sea trout in poor nutritional state will display more risk-taking behaviour than individuals in better nutritional state in order to compensate for their depleted energy stores.

In this study, we tested the hypothesis that poor nutritional state promotes adopting a more high-risk ocean migratory behaviour in sea trout in terms of timing, duration and migration distance. We used both body condition factor and blood plasma metabolites as measures of individuals' nutritional state. Migration behaviour in sea trout has also been
observed to be influenced by sex (Pemberton 1976, Knutsen et al. 2004, Jensen et al. 2019) and body size (Jensen et al. 2014, Jonsson \& Jonsson 2014). Hence, sex determined by genetic analyses and body size were also included in the analyses. We included 286 individual sea trout from seven populations in two fjord systems in Northern Norway in this study to test the general hypotheses that poor nutritional state, females, and large size would promote initiation and greater extent of the marine migration. Specifically, we examined whether nutritional state, sex, and body size (length) influenced $i$. the tendency to migrate to the sea or remain resident in freshwater and/or estuarine habitats, $i i$. the timing of sea entry, iii. the duration of the marine residency, and $i v$. the distance moved out to sea from the river where the fish were tagged.

## 2. MATERIAL \& METHODS

2. 1 Study site

This study was conducted in two Norwegian fjord systems in Nordland County; Skjerstadfjorden $\left(67^{\circ} \mathrm{N}\right)$ and Tosenfjorden $\left(65^{\circ} \mathrm{N}\right)$ (Figure 1) as part of two larger tracking studies, enabling sampling from seven river systems. In Tosenfjorden, fish were captured and tagged in the period 27 March to 10 June in the Rivers Åbjøra and Urvold (Figure 1). In Skjerstadfjorden, fish were captured and tagged during 28 April to 15 June in the rivers Saltdalselva, Botnvassdraget, Lakselva, Laksåga, and Kosmovatnet (Figure 1). Estuaries were defined as the transition zone between the freshwater and marine environment, where the water masses were expected to be brackish throughout the year. For all rivers, this included receivers that were deployed less than 600 meters from the river mouth, except for River Saltdalselva where receivers deployed up to 1 km from the river mouth were categorized as estuarine habitats.

In the Tosenfjorden study area, River Åbjøra has 24 km of river stretch available for
anadromous fish, and includes a large estuarine area influenced by the tide (about $1.6 \mathrm{~km}^{2}$ of tidal affected surface area including shoreline areas inundated at high tide and the lower sections of the river including an estuarine pool), and Lake Åbjørvatnet (surface area of 4.8 $\mathrm{km}^{2}, 81$ meters above the sea). River $\AA$ Abjøra is regulated for hydropower production and has a minimum discharge of $7 \mathrm{~m}^{3} / \mathrm{s}$. River Urvold has an average water discharge of $5 \mathrm{~m}^{3} / \mathrm{s}$, is not developed for hydropower production, and consists of a 200 m steep river stretch from the sea to Lake Urvoldvatnet (surface area of $0.6 \mathrm{~km}^{2}, 8.6$ meters above sea level). In the inner end of Lake Urvoldvatnet, River Urvold has about 1 km of river stretch available for anadromous fish. The estuary of River Urvold is small (about $0.002 \mathrm{~km}^{2}$ of tidal influenced area inside the littoral zone) because the steep river drains straight into the open fjord.

In the Skjerstadfjorden study area, River Saltdalselva is a large river with average discharge of $55 \mathrm{~m}^{3} / \mathrm{s}$ and 66 km of river stretch available for anadromous fish. Due to its large size, and relatively slow-running areas in its lower part, River Saltdalselva has a relatively large estuary (about $0.47 \mathrm{~km}^{2}$ tidal influenced areas inside the littoral zone). There is only one lake available for anadromous fish in River Saltdalselva, Lake Vassbotnvatn, which is located in a tributary. River Botnvassdraget has a 500 m steep river stretch to Lake Botnvatnet ( 12 m above the sea), and continues upstream of the lake, making about 8 km of river stretch available for anadromous fish. River Botnvassdraget's confined estuary covers about 0.002 $\mathrm{km}^{2}$ of tidal influenced area. River Lakselva has about 7 km of river stretch with no lakes available to anadromous fish, and has a tidally influenced surface area of about $0.08 \mathrm{~km}^{2}$ in its estuarine area. River Laksåga hasabout 6.5 km of river stretch available for anadromous fish and drains into two large brackish-water lakes influenced by the sea (about $15 \mathrm{~km}^{2}$ surface area). River Laksåga is regulated for hydropower purposes. River Kosmovatnet has about 6 km of river stretch available to anadromous fish, and drains into a brackish-water
lake of about $8 \mathrm{~km}^{2}$, separated from the sea by a 1 km narrow channel where the tide governs the direction of the current.

### 2.2. Capture and tagging of fish

A total of 286 sea trout, divided into 10 groups based on location and year (Table 1), were captured and tagged with individually coded acoustic transmitters (Thelma Biotel AS; 9 mm and 13 mm , expected battery life 10-24 months, tag size depended on body size) during 2016-2018. Fish from River Åbjøra, River Kosmovatnet and River Laksåga were caught, in the estuarine parts of the river systems. Fish from River Urvold were caught in Lake Urvold (freshwater), except for 13 individuals during spring 2017 (Table 1) that were caught in the river mouth. All fish caught in River Saltdalselva were caught in the river. The fish were captured by angling or by gillnets in the rivers, lakes and/or estuarine areas. Gill nets were continuously monitored and quickly tended when a fish was detected to minimize the time fish were entangled in the nets, and the fish were released using scissors to cut the netting to prevent damage to the skin and gills. A non-lethal blood sample was drawn shortly after capture (max 5 ml blood per kg body mass, Lawrence et al., 2020). The fish were held in keep nets for up to four hours before they had a transmitters implanted.

Prior to tagging, each sea trout was anesthetized for 4 minutes using $0.5 \mathrm{ml}^{-1} 2$ -phenoxy-ethanol (EC No. 204-589-7, Sigma-Aldrich, USA). For most fish, tags were inserted through a $1.5-3 \mathrm{~cm}$ incision in the body cavity (Cooke et al. 2011, Eldøy et al. 2015). For the fish tagged in River Åbjøra in 2017 (Table 1), the transmitters were externally attached using a wire through the dorsum about 1 cm below the dorsal fin, with a silicone plate between the tag and the fish and a plastic plate on the opposite side of the dorsum to prevent erosion on skin and flesh. The sea trout were subsequently placed in a holding tank until recovery from anaesthesia and released in a slow flowing area as close to the capture location as possible.

The experimental procedures were approved by the Norwegian National Animal Research Authority (permission number 2015/8518, 1614092 \& 18/67706).

### 2.3 Fish tracking

The tagged sea trout were tracked using 74 acoustic receivers in Tosenfjorden and 82 acoustic receivers in Skjerstadfjorden (Vemco Inc. models: VR2, VR2-W and VR2-AR). The fish in Tosenfjorden were tracked from May 2016 to December 2017, while the fish tagged in Skjerstadfjorden were tracked from May 2017 to December 2018, although not all receivers were operative throughout these periods (Figure 1).

Tracking data were filtered for false registrations generated by code collisions with simultaneously transmitting tags, or by noise in the environment (Pincock 2012). After empirically assessing the frequency of false detections (i.e., a receiver reported detection of an unused transmitter ID) recorded by each receiver, automated filtering was applied to 16 receivers in Tosenfjorden, and 4 receivers in Skjerstadfjorden (see Figure 1 for filtered receiver locations). The filter required that a tagged sea trout had to be registered at least two times by a receiver within a 10-minute time period to be accepted as a true registration, and resulted in removal of 68682 of 2191047 detections (3.1\%) in Tosenfjorden and 2402 of 594345 detections ( $0.4 \%$ ) in Skjerstadfjorden. The data were subsequently visually inspected by plotting a timeline of all recordings for each fish, and registrations that did not fit with the overall migration track of each fish were also removed (i.e. detections suggesting unrealistic migration speed and/or passing multiple receiver gates without detection).

### 2.4 Processing and analysing blood samples

Blood samples were stored in tubes and immediately placed in an ice water bath for up to 3 hours before being centrifuged at 1163 g for 10 min . Blood plasma was flash-frozen in a liquid nitrogen dry shipper and subsequently stored at $-80^{\circ} \mathrm{C}$ until biochemical analyses could be carried out. Plasma triglyceride levels were assessed in duplicates using the
manufacturer's suggested protocols with a commercially available colorimetric kit (Cayman Chemical Company, USA). Total plasma protein levels were determined using a Bradford's assay (Bradford 1976, Kruger 2009) using commercial reagents (Bio-Rad Laboratories (Canada) Ltd., Mississauga, ON, Canada). Plasma $\mathrm{Ca}^{2+}$ concentrations were determined using flame spectrophotometry (Varian Spectra AA 220FS, Varian Inc., Palo Alto, CA, USA) for a single replicate. Due to the limited volume of blood that could be taken from each fish (see Lawrence et al. 2020), there was not enough blood for all individuals to run all biochemical assays. Therefore, 214 fish were tested for blood plasma triglycerides, 204 fish were tested for blood plasma protein and 185 fish were tested for blood plasma calcium.

### 2.5 Determining the sex of individuals

DNA samples from adipose fin clips taken at the time of tagging and preserved in ethanol were used to genetically determine the sex of the sea trout. DNA was extracted using the QuickExtract kit (Epigen) using the manufacturer's protocol but with extraction volume reduced to $150 \mu$. Using $10 \mu 1$ reactions of the Qiagen Multiplex PCR kit and Salmo-sdY-F and Samo sdYR primers, PCR amplification was applied to a 200 base pair fragment from the first intron of the male-specific SDY gene (Quéméré et al. 2014). PCR steps for denaturation, annealing and extension were: incubation at $95^{\circ} \mathrm{C}$ for $15 \mathrm{~min}, 11$ cycles of touchdown PCR, held at $94{ }^{\circ} \mathrm{C}$ for $30 \mathrm{~s}, 63-52{ }^{\circ} \mathrm{C}$ for 30 s , then $72{ }^{\circ} \mathrm{C}$ for 1 min followed by 25 cycles at $94{ }^{\circ} \mathrm{C}$ for $30 \mathrm{~s}, 52^{\circ} \mathrm{C}$ for $30 \mathrm{~s}, 72^{\circ} \mathrm{C}$ for 1 min and a final extension at $72^{\circ} \mathrm{C}$ for 10 min . Sex was determined by running PCR products in $1 \%$ agarose gels.
2.6 Migrating to the sea $v s$. remaining in freshwater and estuaries

The trout could remain resident in the habitat where they were captured and tagged (river, lake and estuarine habitats), or migrate to the fjord. Individual sea trout were considered sea migrants if they were recorded by any marine receiver except those categorized as estuarine
of the river where they were tagged. Fish that we lost track of shortly after tagging, or which showed a "permanent residency" at a particular receiver indicative of mortality or tag loss within the receiver's detection range were excluded from statistical analyses $(\mathrm{n}=7)$.

### 2.7 Timing of sea entry and duration of the marine migration

The timing of sea entry was calculated for all individuals that were recorded leaving the freshwater and estuarine areas of the watercourse where they were tagged. The timing of sea entry was recorded as the first detection of a tagged fish in the estuary for Rivers Urvold, Saltdalselva and Botnvassdraget. In River Åbjøra, the timing of sea entry was defined as the time of the last recorded detection in the river's estuary, provided the fish was subsequently detected by a receiver in the fjord. For the fish tagged in Laksåga, the time of sea entry was defined as the time of first detection by a receiver in the fjord. The different definitions for timing of sea entry were due to logistical and hydrologic constraints that required different approaches to receiver deployment in the estuarine areas of the different watercourses. Fish tagged in rivers Lakselva and Kosmovatnet were excluded from these analyses due to small sample sizes ( $n \leq 2$ ).

Residence time at sea was calculated as the period between the time a fish entered the sea to its last detection in the sea prior to entering the river during the first year of tracking. In River Åbjøra, the sea journey was considered to have ended at the last detection on a receiver deployed in the river mouth, provided that detection was followed by subsequent detections within the watercourse. In some cases, sea trout transitioned between freshwater and marine habitats multiple times within a year. Time spent in the freshwater habitat between migrations to the sea was not included in the total marine migration time. Fish tagged in Lakselva and Kosmovatnet were excluded from the analyses of residence time at sea due to a low sample size $(n=1)$. At these sites most tagged fish stayed in the river and estuary.
2.8 Maximum migration distance at sea

For marine migrants, the maximum migration distance at sea from the river mouth was calculated for the fish from Rivers Åbjøra, Urvold, Saltdalselva, Botnvassdraget and Laksåga. Because receivers were only deployed in the inner part of Skjerstadfjorden in 2016, the fish tagged in Skjerstadfjorden in 2016 were excluded from the analyses of migration distance. The maximum migration distance for each fish was calculated as the distance from the receiver deployed closest to the mouth of the river in which the fish were tagged to the furthermost receiver at sea where the fish was recorded. This was done by estimating the shortest migration route avoiding land, using the 'costDistance' function in the gdistance R package (van Etten 2018). Fish that were not detected after 20 July of the year of tagging (which was the date corresponding to the upper $95 \%$ percentile for reaching maximum distance for fish observed returning to watercourses), that were not last observed returning to freshwater, or were last observed at the outer arrays of receivers in the fjord were not included in the maximum migration distance analysis $(\mathrm{n}=21)$.

### 2.9 Statistical analyses

To test for effects of nutritional state, sex and body size (natural length) on migratory behaviour, we used a set of generalized mixed effect models. The behavioural traits that were used as response variables were either binomial (migrated or did not migrate) or continuous and normally distributed (timing, duration and distance of sea migration). Independent variables (fixed explanatory effects) were sex, body size and nutritional state (body condition and blood plasma triglycerides). Tagging years were nested within populations (watercourse) and used as random effects. We did not aim to investigate which of the nutritional state variables employed were the best proxies. As such, we fitted one full model for each of the nutritional state variables (i.e. the nutritional state variables were used simultaneously in a full model). As all nutritional indicators were found to be correlated, and to simplify the
presentation and interpretation of results in this study, modelling using blood plasma protein and blood plasma triglycerides was excluded from the manuscript, but can be found in Appendices $1-8$. This approach allowed us to avoid issues with co-linearity among nutritional indicators.

All statistical analyses were conducted in R Studio version 1.2.1335 (RStudio Team 2019) and $R$ version 3.5.3 ( $R$ Core Team 2019) with the 'glmer' function in the 'lme4' $R$ package (Bates et al. 2015) for the model with a binomial dependent variable. The 'lme' function in the nlme R package (Pinheiro et al. 2018) was used for models with normally distributed dependent variables. Collinearity within models was checked using the 'check_collinearity' function in the performance R package (Lüdecke et al. 2020), and collinearity was found to be low (VIF $\leq 1.09$ ). All variables were standardised prior to modelling using the 'scale' function in the R 'base' package. Blood plasma triglycerides values were log-transformed in order to stabilise variance. Body condition factor was calculated from the formula $K=100 \times$ mass $(\mathrm{g}) \times$ total length $(\mathrm{cm})^{-3.028}$, as the regression coefficient of the mass-length relationship was 3.028 for the tagged individuals. Model selection was conducted using Akaike information criterion (AIC) (Anderson et al., 2001), with the 'dredge' function in the MuMIn R package (Barton, 2019). In cases when model selection left us with support for multiple alternative models ( $\Delta$ AIC $<2$ ), conditional model averaging was applied, using all alternative models ( $\Delta \mathrm{AIC}<4$ ) to estimate the coefficients of the explanatory variables. Kruskal-Wallis-tests were applied for comparisons among groups of tagged fish (based on tagging year and population) in terms of body size, body condition factor, blood plasma triglycerides, blood plasma protein and blood plasma calcium. Spearman correlation tests were applied to test for correlations between pairs of nutritional indicators (body condition factor, blood plasma triglycerides, blood plasma protein and blood plasma calcium). For visualization purposes, linear regression lines were fitted to the
relationships among nutritional correlates in Figure 3 using the 'geom_smooth' function in the ggplot2 R package (Wickham 2016). Spearman correlation tests were applied to test for correlations between behavioural traits (timing of sea entry, marine residency and marine migration distance).

The raw tracking dataset on individual fish generated and analysed during the current study is uploaded to the Ocean Tracking Network data system (www.oceantrackingnetwork.org).

## 3. RESULTS

### 3.1 Characteristics of tagged fish

The results were based on 286 trout ( 165 females, 121 males, i.e. $58 \%$ females, $42 \%$ males) with a body size ranging from 270 to 890 mm (mean $=471 \mathrm{~mm}, \mathrm{SD}=129 \mathrm{~mm}$ ) (Figure 2). The fish were divided into ten groups based on the river and year they were tagged. The proportion of females within these groups varied between $38 \%$ and $78 \%$ (Figure 2). There were significant differences in body condition factor among the groups (Kruskal-Wallis test $n$ $=286, P<0.001$, Figure 2). Average concentrations of the nutritional metabolites (pooled samples for all fish from all rivers) derived from blood plasma sampling were $0.71 \mathrm{mmol}^{-1}$ triglycerides $(\mathrm{SD}=0.85$, range $0.004-4.36), 25.03 \mathrm{mg} \mathrm{ml}^{-1}$ of protein $(\mathrm{SD}=5.70$, range $8.87-$ $45.85)$ and $3.21 \mathrm{mmol} \mathrm{l}^{-1}$ of calcium $(\mathrm{SD}=0.43$, range $=1.94-5.56)$ among the groups (Figure 2). However, there were significant differences among the groups in concentrations of blood plasma triglycerides (Kruskal-Wallis test; $n=214, P<0.001$ ), plasma protein ( $n=$ 204, $P<0.001$ ) and plasma Calcium ( $n=185, P<0.001$, Figure 2).
3.2 Correlations between the nutritional indicators

There were significant positive correlations between all the measured variables reflecting nutritional state (Appendix 1). There was a positive correlation between body condition factor and 1) $\log$ transformed blood plasma triglycerides (Spearman's correlation; $n=214, r_{\mathrm{s}}$ $=0.36, P<0.001)$, 2) blood plasma protein $\left(n=204, r_{\mathrm{s}}=0.42, P<0.001\right)$, and 3) plasma calcium ( $n=185, \mathrm{r}_{\mathrm{s}}=0.23, \mathrm{P}<0.001$ ). There were positive correlations between the $\log$ transformed blood plasma triglycerides and both blood plasma protein $\left(n=194, r_{\mathrm{s}}=0.45, P\right.$ $<0.001$ ) and blood plasma calcium ( $n=178, r_{\mathrm{s}}=0.31, P<0.001$ ). There was also a positive correlation between blood plasma protein and blood plasma calcium ( $n=173, r_{\mathrm{s}}=0.60, P<$ $0.001)$.
3.3 To migrate to the sea or stay in freshwater and estuaries Of the 286 tagged trout, 173 individuals migrated to the sea, while 106 individuals remained in freshwater and estuaries during the rest of the year. For 7 individuals, migratory decision could not be determined due to absence of detections, or detection records suggesting mortality, tag loss or tag malfunction. Overall, the models (see below) suggested that the fish migrating to the sea had greater body sizes than those remaining in freshwater, that females had a greater tendency to migrate to the sea than males, and that those migrating had lower body condition factors and lower blood plasma triglyceride levels than individuals remaining resident (Figure 3 and 4, Appendix 2 and 3).

Using body condition factor as the nutritional indicator, a model for migratory decision which included condition factor, sex and body size, and an alternative model excluding sex, were of equally good ( $\Delta \mathrm{AIC}=0.62$, Appendix 2 ). Conditional model averaging indicated that body size had the strongest effect on migratory decision, followed by condition factor and sex, respectively (Figure 3, Appendix 3).

Using plasma triglycerides as nutritional indicator, the model on migratory decision which included sex and body size was the best model ( $\triangle$ AIC 2.22 to second best model;

Appendix 2). The model estimates showed that sex had the strongest effect on migratory decision, followed by body size and blood plasma triglycerides, respectively (Figure 4, Appendix 3).

### 3.4 Timing of sea entry

Timing of sea entry could be determined for 161 individuals. There were four models with $\Delta$ AIC $<2$ (Appendix 4) for the timing of sea entry, including the null model, when using body condition factor as a nutritional indicator. Here, the model averaging estimates generally suggested limited effects of all explanatory variables (Figure 3, Appendix 5).

Three equally well supported models for timing of sea entry were identified when using plasma triglycerides as the nutritional indicator ( $\Delta \mathrm{AIC}=1.89$, Appendix 4 ). The model averaging estimates indicated that sea trout with higher plasma triglyceride levels entered the sea later in the season, while the effect of sex and body size on timing of sea entry was limited (Figure 4, Appendix 5).
3.5 Marine residence time

The marine residence times could be determined for 74 individuals. Three equally well supported models for the marine residence time were identified when using body condition factor as the nutritional indicator ( $\Delta \mathrm{AIC}=1.65$, Appendix 6). The model estimates from conditional model averaging showed that condition factor had the strongest effect on marine residence time, showing that sea trout with lower condition factors spent longer times at sea (Figure 3, Appendix 7).

There were four models with $\Delta \mathrm{AIC}<2$ (Appendix 6) for the marine residence time, including the null model, when using body plasma triglycerides as the nutritional indicator. Conditional model averaging for these models showed that the standard errors exceeded the estimated effects of the explanatory variables (Figure 4, Appendix 7).

There was a significant negative correlation between timing of sea entry and marine residence time (Spearman's correlation; $n=80, P<0.001$ ), where fish entering the sea earlier spent more time at sea.
3.6 Maximum migration distance in the sea

Maximum migration distance could be determined for 111 individuals. The full model on migration distance, which included condition factor, sex and body size, and an alternative model excluding sex, were equally well supported ( $\Delta \mathrm{AIC}=1.97$, Appendix 8 ). Model averaging showed that larger fish and fish with lower body condition factors migrated further out in the marine habitat, and that sex had limited effect on the migration distance (Figure 3, Appendix 9).

The migration distance model selection process where blood plasma triglyceride was used as nutritional indicator found that a model only including body size and an alternative model including body size and sex were equally well supported ( $\Delta \mathrm{AIC}=1.73$, Appendix 8 ). Here, model conditional averaging showed that larger fish tended to migrate further out to sea than smaller fish, while sex and blood triglycerides had limited effect on the migration distance (Figure 4, Appendix 9).

The maximum migration distance at sea was not correlated with the timing of sea entry (Spearman's correlation; $n=167, P=0.89$ ) or the duration of the marine residency ( $n=$ $69, P=0.49$ ).

## 4 DISCUSSION

Overall, nutritional state, sex and body size (length) influenced the marine migration behaviour of sea trout from the seven study populations in two distinct fjord systems in northern Norway. Sea trout with poor body conditions and low triglyceride levels tended to leave the river and estuaries and migrate to the sea, and individuals with low triglyceride levels migrated to the sea earlier. Fish with poor body condition prior to the migration remained at sea for a longer time-period and migrated further out in the fjords than fish in better condition. Although all the nutritional indicators were found to be highly correlated, this study suggests that measuring both body condition and blood plasma metabolites gave a better evaluation of the nutritional state of the individuals and the impacts of nutritional state on behaviour. While body condition results from the balance between energy intake vs. expenditure over time frames of weeks or months, blood plasma triglycerides have previously been observed to change in response to food intake over much shorter time scales (Sheridan 1988, Congleton \& Wagner 2006).

Migrating to the marine environment is believed to provide better feeding opportunities and potentially increased growth and reproductive capacity (Klemetsen et al. 2003, Thorstad et al. 2016) because of the higher productivity in marine habitats than freshwater habitats in high latitude areas (Gross et al. 1988). On the other hand, energetic costs related to migration and osmoregulation, the risk of predation, disease, or other factors that could prevent a migratory individual from returning to freshwater spawning grounds are all risk factors presumed to be higher when an individual migrates to the sea (Thorstad et al. 2015, Jensen et al. 2019). In this study, the observed effects of nutritional state on sea trout migratory behaviour suggest that individuals in a poor nutritional state tend to engage in riskier migration behaviour than fish in a better nutritional state. The results from this study also suggest that individuals in a poor nutritional state were energetically limited in
freshwater, tipping the cost $v s$. benefit trade-off in favour of migration. In a previous study, Boel et al. (2014) found that sea trout with poor body condition were most likely to migrate towards the sea. Davidsen et al. (2014) observed that starved hatchery-reared sea trout released to the wild utilized sea habitat to a greater extent than well fed hatchery fish, which tended to remain in the lower parts of the river and estuarine areas to which they were released. The observed relationship between nutritional state and timing of sea entry is also consistent with previous studies on reconditioning post-spawn Atlantic salmon S. salar which exhibited earlier sea entry for individuals in poor body condition (Halttunen et al. 2018, Bordeleau et al. 2019). Birnie-Gauvin et al. (2019a) showed that elevated baseline cortisol levels, possibly in response to nutritional need, were associated with earlier migration towards the sea for post-spawned sea trout. Interestingly, we observed a stronger effect of blood plasma triglycerides levels than of body condition factor on the timing of sea entry. This may suggest behavioural response to nutritional need, as acute triglyceride deprivation was a strong predictor for migratory initiation. Alternatively, it may suggest that opportunistic feeding in the freshwater or estuarine habitats during early spring may recondition nutritional state and delay the initiation of marine migration.

Once migration has occurred, sea trout with poor body condition factors tended to spend more time at sea and migrate further out in the marine habitat, possibly reflecting a greater need to recondition compared to individuals with better body condition factors. There was a significant relationship between timing of sea entry and duration of marine residency, where fish that migrated early resided longer at sea. The prolonged residency at sea may enable sea trout to recondition for the next spawning and overwintering season but may also include higher risk of mortality as marine habitats often have greater abundance of potential predators. A previous study by Haraldstad et al. (2018) showed that post-spawned sea trout in poor body condition were more likely to skip spawning the following season compared to
individuals in better body condition. Bordeleau et al. (2018) reported that the pre-migratory level of blood plasma triglycerides was negatively correlated with the duration of marine residency in veteran sea trout migrants. According to previous studies, sea trout with a low body condition tended to migrate further out to sea compared with individuals in a better body condition (Davidsen et al. 2014, Eldøy et al. 2015, Bordeleau et al. 2018). In the present study, differences in characteristics of the near marine habitats among the multiple sites we studied probably impacted how far the sea trout from the various rivers needed to migrate to meet their metabolic demands. No correlation was found between migration distance and timing of marine entry or marine residence time.

As expected, there were significant positive correlations among all measured nutritional indicators. A previous lab experiment by Congleton et al. (2006) documented low levels of blood plasma triglycerides, blood plasma protein and blood plasma calcium in starved juvenile salmonids. Overall nutritional state is determined by net differences over periods of weeks or months between energy intake and energy expenditure (Congleton \& Wagner 2006). Poor nutritional state could likely be explained by limited feeding while overwintering, and for fish that reproduced, also by the energy expenditure during spawning the previous autumn (Bordeleau et al. 2018). The energy investment in spawning can be substantial for brown trout (Jonsson \& Jonsson 2011b). In Lake Vangsvatnet in Norway, Jonsson and Gravem (1985) documented that immature migrants fed little while in freshwater and that mature migrants stopped feeding after the spawning season.

Although the nutritional status of most fish in the study was poor, there was large variation in nutritional state both among individuals and groups of fish. There may be several reasons for this, including differences in nutritional state after the previous growth season, energy investment in previous spawning, overwintering conditions, metabolic rate and feeding activity (Midwood et al. 2015, Auer et al. 2016). Some individuals had elevated
nutritional indicators indicative of a better nutritional status. These individuals mainly belonged to the groups of fish tagged in the estuarine habitats of River Åbjøra, River Lakselva and the Kosmovatnet watercourse. For these fish, elevated blood plasma triglyceride level was the most obvious signal. This might suggests that the fish had started feeding prior to tagging, because triglycerides become elevated a few hours after feeding (Sheridan 1988), and a previous laboratory experiment documented that blood plasma triglycerides recovered quickly when refeeding began after a starvation period in rainbow trout Oncorhynchus mykiss (Congleton \& Wagner 2006). Common for these groups of tagged fish was that they were captured and tagged in lower parts of watercourses which have relatively large estuarine areas likely suitable for opportunistic feeding during early spring.

Females were more likely to migrate to the sea than males, instead of remaining in the freshwater and estuarine areas of the river where they were tagged. Previous studies with this species have also noted that females are more likely to migrate than males (Pemberton 1976, Knutsen et al. 2004, Jensen et al. 2019). In a study in Tosenfjorden, Bordeleau et al. (2018) found that females in the Åbjøra watercourse were more likely to leave the estuarine areas than males. This is likely caused by a greater benefit of increased feeding opportunities for females than males due to the strong correlation between female body size and the number of eggs the female can produce (Elliott 1995). Sexual bias in migratory behaviours is a wellknown phenomenon that has previously been observed in a range of salmonid species (reviewed by Dodson et al. 2013).

The models in the present study provided limited support for the influence of sex on migration timing, duration and distance migrated at sea. Some previous studies have reported that male sea trout tended to migrate earlier (Jensen 1968, Östergren \& Rivinoja 2008), while others at different sites have suggested an earlier migration timing for females (Berg \& Berg 1989). $\operatorname{Berg} \& \operatorname{Berg}$ (1989) also observed that females had a longer duration of marine
residency than males. Bordeleau et al. (2018) found that females migrated further from the river than males and were more likely to migrate to the outer fjord areas of Tosenfjorden. Although the reasons female and males differed in their migration patterns among these different sites remain obscure, it suggests that a combination of local environmental conditions and population characteristics may plays an important role for the trade-off mechanisms shaping the migratory decisions of individuals within sea trout populations. This plasticity is one of the reasons the species has been so successful.

Larger fish of both sexes were more likely to migrate to the sea and migrated greater distances at sea than smaller fish. These tendencies are likely driven by the need of larger individuals to find more prey of larger size than the smaller fish, that larger fish are less susceptible to predation than smaller fish, and possibly because larger fish may be more powerful swimmers (Dill 1983, Klemetsen et al. 2003). Individual sea trout tend to repeat their migratory patterns among successive years (Eldøy et al. 2019) although some studies suggest that iteroparous salmonids may reduce their migration distances as they become larger and older (Svärdson \& Fagerström 1982, Bond et al. 2015). The earlier seaward migration of large fish observed in the present study is similar to the timing observed in previous studies (Pemberton 1976, Bohlin et al. 1996, Jonsson \& Jonsson 2009). The positive correlation we noted between body size and the duration of the marine migration is consistent with previous work (Eldøy et al. 2015). However, the tendency we found for larger fish to migrate further out to sea compared to smaller individuals has only been noted in a few of the previous studies on this species (e.g. Berg \& Berg 1989, Jensen et al. 2014, Jonsson \& Jonsson 2014).

In conclusion, despite the large individual and among-group variation observed in both nutritional state and migratory behaviour, this study showed that sex, body size and premigratory nutritional state strongly influenced the migratory patterns of sea trout. Anadromy
is considered a quantitative threshold trait, where environmental thresholds for triggering behavioural responses are genetically determined (Ferguson 2006, Ferguson et al. 2019). Previous studies have suggested that the migratory behaviour of brown trout is a continuum of behavioural responses to the environmental cues experienced by the individuals in coastal trout populations (Cucherousset et al. 2005, Boel et al. 2014, Villar-Guerra et al. 2014). However, the importance of different factors affecting the pre-migratory nutritional state, and the influence of carry-over effects are poorly understood (O'Connor et al. 2014). For example, it is unknown how the success of a previous feeding migration interacts with spawning investment and over-wintering conditions to determine the nutritional state and subsequent marine migrations of post-spawned, veteran sea trout migrants (Bordeleau 2019). As shown by Jensen et al. (2020), life history patterns or decisions adopted early in life may persist throughout an individual's lifetime, and significantly affect the animal's lifetime fitness. Jensen et al. (2020) showed that early migrants continued to migrate early throughout their life time, had better growth, and a larger lifetime fecundity. This suggests that individuals developing under favourable conditions will gain fitness benefits throughout their lifetime. Jensen et al. (2020) therefore concluded that individuals that experience environmental conditions as juveniles in freshwater and/or with genes that contribute to a large smolt size and early smolt migration may benefit preferentially from growth opportunities in the sea, and the benefits of the early adoption of anadromy enables them to continue with early and longer migrations during following years. However, the fact that sea trout populations do not evolve to contain exclusively early migrants highlights again that there are costs that counterbalance the strategy.

While at sea, sea trout commonly reside in habitats heavily affected by human activity (Nevoux et al. 2019). Salmon lice infestation related to open cage farming of Atlantic salmon was recently evaluated as the biggest threat for Norwegian sea trout stocks (Norwegian

Scientific Advisory Committee for Atlantic Salmon 2019) . Serra-Llinares et al. (2020) documented that sea trout infested with salmon lice altered their migration behaviour and experienced increased mortality. The results of the present study, where fish in poor nutritional state seemed to migrate to the sea earlier and spent more time at sea, suggest that fish in poor nutritional state may display behavioural patterns that make them especially vulnerable to such negative anthropogenic factors at sea. This is both because their longer stay at sea increases the risk of being infested by salmon lice, and because they migrate to areas with high salinity favourable for sea lice instead of remaining in brackish water areas where the lice do not survive well. The links between migration behaviour, human induced stressors and reproductive success throughout the lifetime of sea trout remain obscure. Future studies examining the link between marine migration behaviour and reproductive investment over consecutive years are therefore advocated.

## 5. ACKNOWLEMENTS

This study was part of the CHASES (Consequences of land-use change and human activity on anadromous salmonids and the ecosystem services that they provide) project funded by the Research Council of Norway (ref: 255110/E50) with additional funding from the Norwegian Environmental Agency, the County Governor of Nordland, Nordland County Authority, Saltdal, Fauske and Bodø municipalities, Sinkaberg-Hansen AS, Salten Aqua AS, NCA-Aquaculture, The River Åbjøra landowners association, Plahtes Eiendommer, River Saltdalelva landowner association, SKS Produksjon, SISO Energi, the IRIS-foundation, the Natural Sciences and Engineering Research Council of Canada, Ocean Tracking Network and the NTNU University Museum.

The crew of RV Gunnerus, Lars Rønning, Jan Ivar Koksvik, Aslak Darre Sjursen,

Vegard Pedersen Sollien, Paul Skarsvåg, Stein Hugo Hemmingsen, Kristian Lian, Embla Østebrøt, Hilde Dørum, Kristina Johansen, Ashley Vold, Ole Johan Hornenes, Torjus

Haukvik, Charlotte Hallerud, Frode Tjønn, Adam Piper, Petter Kristensen, Johan Åsbakk, Peder Straume, Jo Vegar Arnekleiv, Karstein Hårsaker, Kjell Christian Rambech, Geir Johny Monsen, Sveinung Kristiansen, Geir Jensen and Flavier Morin-Doré are thanked for their extensive help during fieldwork. Marc André Francis Daverdin is thanked for producing the map of the study area. Colin Adams and two anonymous reviewers are thanked for commenting on earlier drafts of the manuscript.

## 6. REFRENCES

Archer LC, Hutton SA, Harman L, McCormick SD and others (2020) Food and temperature stressors have opposing effects in determining flexible migration decisions in brown trout (Salmo trutta). Global Change Biology 26:2878-2896
Auer SK, Salin K, Anderson GJ, Metcalfe NB (2016) Flexibility in metabolic rate and activity level determines individual variation in overwinter performance. Oecologia 182:703-712
Bates D, Maechler M, Bolker B, Walker S (2015) Fitting linear mixed-effects models using Ime4. Journal of Statistical Software 67:1-48
Berg OK, Berg M (1989) The duration of sea and freshwater residence of the sea trout, Salmo trutta, from the Vardnes River in northern Norway. Environmental Biology of Fishes 24:23-32
Birnie-Gauvin K, Flávio H, Kristensen ML, Walton-Rabideau S and others (2019a) Cortisol predicts migration timing and success in both Atlantic salmon and sea trout kelts. Scientific Reports 9:1-9
Birnie-Gauvin K, Thorstad EB, Aarestrup K (2019b) Overlooked aspects of the Salmo salar and Salmo trutta lifecycles. Reviews in Fish Biology and Fisheries 29:749-766
Birnie-Gauvin K, Lennox RL, Guglielmo CG, Teffer AK and others (In Press) The value of experimental approaches in migration biology. Physiological and Biochemical Zoology 00:000-000
Boel M, Aarestrup K, Baktoft H, Larsen T and others (2014) The physiological basis of the migration continuum in brown trout (Salmo trutta). Physiological and biochemical zoology 87:334-345
Bohlin T, Dellefors C, Faremo U(1996) Date of smolt migration depends on body-size but not age in wild sea-run brown trout. Journal of Fish Biology 49:157-164
Bond MH, Miller JA, Quinn TP (2015) Beyond dichotomous life histories in partially migrating populations: cessation of anadromy in a long-lived fish. Ecology 96:1899-1910
Bordeleau X, Davidsen JG, Eldøy SH, Sjursen AD, Whoriskey FG, Crossin GT (2018) Nutritional correlates of spatiotemporal variations in the marine habitat use of brown trout (Salmo trutta) veteran migrants. Canadian Journal of Fisheries and Aquatic Sciences 75:1-11
Bordeleau X (2019) The post-spawning ecology of iteroparous salmonids: Basis of variability in migratory behaviour and survival, ecological importance and conservation implications. Ph.D. Thesis. 168 p. Halifax: Department of Biology, Dalhousie University.

Bordeleau X, Hatcher BG, Denny S, Whoriskey FG, Patterson DA, Crossin GT (2019) Nutritional correlates of the overwintering and seaward migratory decisions, and long-term survival of post-spawning Atlantic salmon. Conservation Physiology 7:1-13
Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Analytical Biochemistry 72:248-254
Chapman BB, Hulthén K, Brodersen J, Nilsson PA, Skov C, Hansson L-A, Brönmark C (2012) Partial migration in fishes: causes and consequences. Journal of Fish Biology 81:456-478
Congleton JL, Wagner T (2006) Blood-chemistry indicators of nutritional status in juvenile salmonids. Journal of Fish Biology 69:473-490
Cooke SJ, Crossin GT, Patterson DA, English KK and others (2005) Coupling non-invasive physiological assessments with telemetry to understand inter-individual variation in behaviour and survivorship of sockeye salmon: development and validation of a technique. Journal of Fish Biology 67:1342-1358
Cooke SJ, Hinch SG, Farrell AP, Patterson DA and others (2008) Developing a mechanistic understanding of fish migrations by linking telemetry with physiology, behavior, genomics and experimental biology: An interdisciplinary case study on adult Fraser River sockeye salmon. Fisheries 33:321-339
Cooke SJ, Murchie S, Mc Connachie S, Goldberg T (2011) Standardized surgical procedures for the implantation of electronic tags in key Great Lakes fishes. Technical Report. Great Lakes Fishery Commission, Ann Arbor, MI. 44 pp.
Crossin GT, Cooke SJ, Goldbogen JA, Phillips RA (2014) Tracking fitness in marine vertebrates: current knowledge and opportunities for future research. Marine Ecology Progress Series 496:1-17
Crossin GT, Heupel MR, Holbrook CM, Hussey NE and others (2017) Acoustic telemetry and fisheries management. Ecological Applications 27:1031-1049
Cucherousset J, Ombredane D, Charles K, Marchand F, Baglinière J-L (2005) A continuum of life history tactics in a brown trout (Salmo trutta) population. Canadian Journal of Fisheries and Aquatic Sciences 62:1600-1610
Davidsen JG, Daverdin M, Sjursen AD, Rønning L, Arnekleiv JV, Koksvik JI (2014) Does reduced feeding prior to release improve the marine migration of hatchery brown trout Salmo trutta smolts? Journal of Fish Biology 85:1992-2002
Dill LM (1983) Adaptive flexibility in the foraging behavior of fishes. Canadian Journal of Fisheries and Aquatic Sciences 40:398-408
Dingle H, Drake VA (2007) What Is Migration? BioScience 57:113-121
Dingle H (2014) Migration: the biology of life on the move, Vol. Oxford University Press, New York
Dodson JJ, Aubin-Horth N, Thériault V, Páez DJ (2013) The evolutionary ecology of alternative migratory tactics in salmonid fishes. Biological Reviews 88:602-625
Eldøy SH, Davidsen JG, Thorstad EB, Whoriskey F and others (2015) Marine migration and habitat use of anadromous brown trout Salmo trutta. Canadian Journal of Fisheries and Aquatic Sciences 72:1366-1378
Eldøy SH, Bordeleau X, Crossin GT, Davidsen JG (2019) Individual repeatability in marine migratory behavior: A multi-population assessment of anadromous brown trout tracked through consecutive feeding migrations. Frontiers in Ecology and Evolution 7:1-12
Elliott JM (1995) Fecundity and egg density in the redd for sea trout. Journal of Fish Biology 47:893901
Ferguson A (2006) Genetics of sea trout, with particular reference to Britain and Ireland. In: Harris G, Milner $N$ (eds) Sea trout: biology, conservation and management. Blackwell publishing Ltd, Oxford, p 155-182
Ferguson A, Reed TE, Cross TF, McGinnity P, Prodöhl PA (2019) Anadromy, potamodromy and residency in brown trout Salmo trutta: the role of genes and the environment. Journal of Fish Biology 95:692-718

Forseth T, Nesje TF, Jonsson B, Hårsaker K (1999) Juvenile migration in brown trout: a consequence of energetic state. Journal of Animal Ecology 68:783-793
Gross MR, Coleman RM, McDowall RM (1988) Aquatic productivity and the evolution of diadromous fish migration. Science 239:1291-1293
Halttunen E, Gjelland K- $\varnothing$, Hamel S, Serra-Llinares R-M and others (2018) Sea trout adapt their migratory behaviour in response to high salmon lice concentrations. Journal of Fish Diseases 41:953-967
Haraldstad T, Höglund E, Kroglund F, Lamberg A, Olsen EM, Haugen TO (2018) Condition-dependent skipped spawning in anadromous brown trout (Salmo trutta). Canadian Journal of Fisheries and Aquatic Sciences 75:2313-2319
Jachowski DS, Singh NJ (2015) Toward a mechanistic understanding of animal migration: incorporating physiological measurements in the study of animal movement. Conservation Physiology 3:1-12
Jensen AJ (1968) Sea trout (Salmo trutta L.) of the River Istra, western Norway. Report: Institute of Fresh-water Research, Drottningholm 48, 187-213.
Jensen AJ, Finstad B, Fiske P (2019) The cost of anadromy: marine and freshwater mortality rates in anadromous arctic char and brown trout in the arctic region of Norway. Canadian Journal of Fisheries and Aquatic Sciences 76:2408-2417
Jensen AJ, Finstad B, Fiske P, Diserud OH, Thorstad EB (2020) Repeatable individual variation in migration timing in two anadromous salmonids and ecological consequences. Ecology and Evolution 10:11727-11738
Jensen JLA, Rikardsen AH, Thorstad EB, Suhr AH, Davidsen JG, Primicerio R (2014) Water temperatures influence the marine area use of Salvelinus alpinus and Salmo trutta. Journal of Fish Biology 84:1640-1653
Jonsson B, Gravem FR (1985) Use of space and food by resident and migrant brown trout, Salmo trutta. Environmental Biology of Fishes 14:281-293
Jonsson B, Jonsson N (1993) Partial migration: niche shift versus sexual maturation in fishes. Reviews in Fish Biology and Fisheries 3:348-365
Jonsson B, Jonsson N (2005) Lipid energy reserves influence life-history decision of Atlantic salmon (Salmo salar) and brown trout (S. trutta) in fresh water. Ecology of Freshwater Fish 14:296301
Jonsson B, Jonsson N (2009) Migratory timing, marine survival and growth of anadromous brown trout Salmo trutta in the River Imsa, Norway. Journal of Fish Biology 74:621-638
Jonsson B, Jonsson N (2011a) Habitats as template for life histories. In: Jonsson B, Jonsson N (eds) Ecology of Atlantic Salmon and Brown Trout: Habitat as a template for life histories. Springer Netherlands, Dordrecht, p 1-21
Jonsson B, Jonsson N (2011b) Maturation and Spawning. In: Ecology of Atlantic Salmon and Brown Trout: Habitat as a template for life histories. Springer Netherlands, Dordrecht, p 327-414
Jonsson B, Jonsson N (2014) Naturally and hatchery produced European trout Salmo trutta: Do their marine survival and dispersal differ? Journal of Coastal Conservation 18:79-87
Jonsson N, Jonsson B (1998) Body composition and energy allocation in life-history stages of brown trout. Journal of Fish Biology 53:1306-1316
Klemetsen A, Amundsen P-A, Dempson JB, Jonsson B, Jonsson N, O’Connell MF, Mortensen E (2003) Atlantic salmon Salmo salar L., brown trout Salmo trutta L., and Arctic charr Salvelinus alpinus (L.): a review of aspects of their life histories. Ecology of Freshwater Fish 12:1-59
Knutsen JA, Knutsen H, Olsen EM, Jonsson B (2004) Marine feeding of anadromous Salmo trutta during winter. Journal of Fish Biology 64:89-99
Kruger NJ (2009) The Bradford method for protein quantitation. In: Walker JM (ed) The Protein Protocols Handbook. Humana Press, Totawa, p 17-24
Lawrence MJ, Raby GD, Teffer AK, Jeffries KM and others (2020) Best practices for non-lethal blood sampling of fish via the caudal vasculature. Journal of Fish Biology 97:4-15

Lennox RJ, Chapman JM, Souliere CM, Tudorache C, Wikelski M, Metcalfe JD, Cooke SJ (2016) Conservation physiology of animal migration. Conservation Physiology 4:1-15
Lien $L$ (1978) The energy budget of the brown trout population of $\varnothing$ vre Heimdalsvatn. Ecography 1:279-300
Lüdecke D, Makowski D, Waggoner P, Patil I (2020) Performance: Assessment of regression models performance. R package version 0.4.5. https://CRAN.R-project.org/package=performance.
MacCrimmon HR, Marshall TL, Gots BL (1970) World distribution of brown trout, Salmo trutta: Further observations. Journal of the Fisheries Research Board of Canada 27:811-818
Midwood JD, Larsen MH, Boel M, Aarestrup K, Cooke SJ (2015) An experimental field evaluation of winter carryover effects in semi-anadromous brown trout (Salmo trutta). Journal of Experimental Zoology Part A: Ecological Genetics and Physiology 323:645-654
Nevoux M, Finstad B, Davidsen JG, Finlay R and others (2019) Environmental influences on life history strategies in partially anadromous brown trout (Salmo trutta, Salmonidae). Fish and Fisheries 20:1051-1082
Norwegian Scientific Advisory Committee for Atlantic Salmon (2019) Classification of the state of 430 Norwegian sea trout populations. Temarapport no. 7, Norwegian Scientific Advisory Committee for Atlantic Salmon, Trondheim
Næsje TF, Thorstad EB, Forseth T, Aursand M, Saksgård R, Finstad AG (2006) Lipid class content as an indicator of critical periods for survival in juvenile Atlantic salmon (Salmo salar). Ecology of Freshwater Fish 15:572-577
O'Connor CM, Norris DR, Crossin GT, Cooke SJ (2014) Biological carryover effects: linking common concepts and mechanisms in ecology and evolution. Ecosphere 5:1-11
Pemberton R (1976) Sea trout in North Argyll Sea lochs, population, distribution and movements. Journal of Fish Biology 9:157-179
Pincock DG (2012) False detections: what they are and how to remove them from detection data. 11 p. Halifax: AMIRIX Systems Inc. DOC-004691-03.

Pinheiro J, Bates D, DebRoy S, Sarkar D, Team RC (2018) nlme: Linear and nonlinear mixed effects models. R package version 3.1-137, URL: https://CRAN.R-project.org/package=nlme.
Quéméré E, Perrier C, Besnard A-L, Evanno G, Baglinière J-L, Guiguen Y, Launey S (2014) An improved PCR-based method for faster sex determination in brown trout (Salmo trutta) and Atlantic salmon (Salmo salar). Conservation Genetics Resources 6:825-827
R Core Team (2019) R: A language and environment for statistical computing. Vienna: R Foundation for Statistical Computing. URL: http://www.R-project.org/.
RStudio Team (2019) RStudio: Integrated development for R. Boston: RStudio Inc. URL: http://www.rstudio.com/.
Serra-Llinares RM, Bøhn T, Nilsen R, Karlsen $\varnothing$ and others (2020) Increased mortality and altered behaviour of sea trout (Salmo trutta) post-smolts infested with salmon lice (Lepophtheirus salmonis). Marine Ecology Progress Series 635:151-168
Sheridan MA (1988) Lipid dynamics in fish: aspects of absorption, transportation, deposition and mobilization. Comparative Biochemistry and Physiology Part B: Comparative Biochemistry 90:679-690
Simpson AL (1992) Differences in body size and lipid reserves between maturing and nonmaturing Atlantic salmon parr, Salmo salar L. Canadian Journal of Zoology 70:1737-1742
Svärdson G, Fagerström $\AA$ (1982) Adaptive differences in the long-distance migration of some trout (Salmo trutta L.) stocks. Report Instute of Freshwater Research Drottningholm 60:51-80
Thorstad EB, Todd CD, Uglem I, Bjørn PA and others (2015) Effects of salmon lice Lepeophtheirus salmonis on wild sea trout Salmo trutta - a literature review. Aquaculture Environment Interactions 7:91-113
Thorstad EB, Todd CD, Uglem I, Bjørn PA and others (2016) Marine life of the sea trout. Marine Biology 163:1-47
van Etten J (2018) gdistance: Distances and routes on geographical grids. R package version 1.2-2. URL: https://CRAN.R-project.org/package=gdistance.
Villar-Guerra D, Aarestrup K, Skov C, Koed A (2014) Marine migrations in anadromous brown trout (Salmo trutta). Fjord residency as a possible alternative in the continuum of migration to the open sea. Ecology of Freshwater Fish 23:594-603
Weatherley AH, Gill HS (1983) Protein, lipid, water and caloric contents of immature rainbow trout, Salmo gairdneri Richardson, growing at different rates. Journal of Fish Biology 23:653-673
Wickham H (2016) ggplot2: Elegant graphics for data analysis. New York: Springer Publishing.
Wilcove DS, Wikelski M (2008) Going, going, gone: Is animal migration disappearing. PLOS Biology 6:e188
Wysujack K, Greenberg LA, Bergman E, Olsson IC (2009) The role of the environment in partial migration: food availability affects the adoption of a migratory tactic in brown trout Salmo trutta. Ecology of Freshwater Fish 18:52-59
Zera AJ, Harshman LG (2001) The physiology of life history trade-offs in animals. Annual Review of Ecology and Systematics 32:95-126
Östergren J, Rivinoja P (2008) Overwintering and downstream migration of sea trout (Salmo trutta L.) kelts under regulated flows-northern Sweden. River Research and Applications 24:551563

1 Table 1: Description of fjord, watercourse, tracking year, number of individuals, date 2 oftagging and mean ( $\pm \mathrm{SD}$ ) sea trout body size ( mm ) and body mass ( g )

3

| Tracking |  |  |  |  | Body size | Body |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Site | Watercourse | year | Tagging date | $n$ | (mm) | Mass (g) |
| 21/05/2016- |  |  |  |  | $405 \pm 62$ | $705 \pm 269$ |
| Tosenfjorden | Åbjøra | 2016 | 26/05/2016 | 23 | (290-520) | (240-1280) |
| 27/03/2017- |  |  |  |  | $451 \pm 58$ | $847 \pm 318$ |
| Tosenfjorden | Åbjøra | 2017 | 29/03/2017 | 37 | (350-560) | (400-1540) |
| 04/05/2016- |  |  |  |  | $415 \pm 81$ | $639 \pm 401$ |
| Tosenfjorden | Urvoll | 2016 | 10/06/2016 | 24 | (270-620) | (160-1940) |
|  |  |  | 19/04/2017- |  | $463 \pm 98$ | $949 \pm 816$ |
| Tosenfjorden | Urvoll | 2017 | 14/05/2017 | 46 | (330-730) | (290-4550) |
|  |  |  | 01/05/2016- |  | $574 \pm 164$ | $2068 \pm 1812$ |
| Skjerstadfjorden | Botnvassdraget | 2016 | 20/05/2016 | 21 | (300-850) | (200-5800) |
|  |  |  | 11/05/2017- |  | $379 \pm 71$ | $531 \pm 284$ |
| Skjerstadfjorden | Kosmovatnet | 2017 | 15/06/2017 | 19 | (275-470) | (180-990) |
|  |  |  | 30/05/2017- |  | $384 \pm 78$ | $538 \pm 447$ |
| Skjerstadfjorden | Laksåga | 2017 | 31/05/2017 | 34 | (290-630) | (180-2590) |
|  |  |  | 02/05/2017- |  | $442 \pm 53$ | 902 $\pm 377$ |
| Skjerstadfjorden | Lakselva | 2017 | 09/05/2017 | 13 | (360-520) | (410-1580) |
|  |  |  | 28/04/2016- |  | $520 \pm 157$ | $1574 \pm 1611$ |
| Skjerstadfjorden | Saltdalselva | 2016 | 30/04/2016 | 40 | (360-860) | (350-5600) |
|  |  |  | 01/05/2018- |  | $637 \pm 113$ | $2459 \pm 1626$ |
| Skjerstadfjorden | Saltdalselva | 2018 | 10/05/2018 | 29 | (450-890) | (830-8400) |



Figure 1: Map over the study areas in the two fjords Skjerstadfjorden (upper) and Tosenfjorden (lower), with tagging sites and receiver positions indicated. "Automated filtering" indicates receivers where automatic data filtering was applied to remove false detections. Light blue water surface indicates watercourses. Purple to deep blue surface colour indicate the depth of estuarine and marine habitats.


Figure 2: Sex (a), body size (mm) (b), body condition factor (c), blood plasma triglycerides (d), blood plasma proteins (e) and blood plasma calcium (f) for the study's groups of tagged fish (location and year of tagging for the groups indicated on the x -axes). The stacked bar plots (a) shows the proportion of males and females in each group. The box plots show the interquartile range (boxes), median (horizontal line in boxes), the 5th and 95th percentiles (whiskers) and outliers (dots), with number of individuals in each group denoted at the top of the panels.


Figure 3: Estimated effect of body condition factor, sex (male) and body size on the decision to migrate to sea (a), timing of sea entry (b), marine residence time (c) and migration distance at sea (d). The bar plots show the estimated parameter coefficients and their standard error (whiskers) for the best fitted model ( $\Delta \mathrm{AIC}<2$ ) or from conditional model averaging
(including models with $\Delta \mathrm{AIC}<4$ ) where model selection identified multiple models of similar support.


Figure 4: Estimated effect of plasma triglycerides, sex (male) and body size on the decision to migrate to sea (a), timing of sea entry (b), marine residence time (c) and migration distance at sea (d). The bar plots show the estimated parameter coefficients and their standard error (whiskers) for the best fitted model $(\Delta \mathrm{AIC}<2)$ or from conditional model averaging (including models with $\Delta \mathrm{AIC}<4$ ) where model selection identified multiple models of similar support.

