1	The effects of nutritional state, sex and body size on the
2	marine migration behaviour of sea trout
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19	ABSTRACT				
20	The sea trout (anadromous brown trout, Salmo trutta) displays extensive among-individual				
21	variation in marine migration behaviour. We studied the migration behaviour of 286 sea trout				
22	(27-89 cm) tagged with acoustic transmitters in the spring, in seven populations located in				
23	two distinct marine fjord systems in Norway. We examined whether individual nutritional				
24	state, sex, and body size influenced marine migration behaviour in terms of <i>i</i> . the decision to				

25 migrate to the sea or remain resident in freshwater and/or estuarine habitats, *ii*. seasonal

timing of sea entry, *iii*. duration of the marine residency, and *iv*. migration distance at sea 1 from the home river. Most sea trout were in a poor nutritional state in the spring prior to 2 3 migration. Sea trout with low body condition factors and low plasma triglyceride levels were 4 more likely to migrate to sea, and low triglyceride levels were also associated with earlier sea-entry. Poor body condition also increased the probability of individuals remaining at sea 5 longer and migrating further offshore compared to fish in better condition. Females were 6 7 more likely to migrate to the sea than males. Larger fish were also more likely to migrate to sea instead of remaining in freshwater and estuaries and dispersed over greater distances from 8 9 the river than smaller fish. In conclusion, this study documented general trends across multiple populations and showed that nutritional state, sex and body size influence important 10 aspects of the marine migration behaviour of sea trout. 11

12

13 1. INTRODUCTION

Migration behaviour is observed in a wide range of taxa (Dingle 2014). There are various 14 proximate explanations for why animals migrate, but the ultimate reason is for the 15 optimization of individual growth and survival in order to increase lifetime fitness (Dingle & 16 17 Drake 2007). Throughout their lifetimes, individuals must continually allocate energy to various life-history activities, while balancing the metabolic demands for somatic growth, 18 19 maturation, and reproduction (Zera & Harshman 2001). Diadromy, which refers to migrations 20 between marine and freshwater habitats, is thought to have evolved because of differences in 21 food availability between these habitats (Gross et al. 1988). Among the fish family Salmonidae, all species spawn in freshwater, but many are anadromous, which means 22 23 individuals migrate to sea at some point in their lives to exploit the richer marine food resources (Jonsson & Jonsson 1993). Salmonid populations often consist of both freshwater 24

resident and sea migrating individuals (i.e partial migration, Chapman et al. 2012). Body size
is positively correlated with fecundity within salmonids (Elliott 1995). Marine migration is
advantageous if the fitness benefits of larger body size outweighs the cost of migration,
including increased risk of mortality, disease and failure to reach spawning grounds
(Klemetsen et al. 2003, Thorstad et al. 2016).

The brown trout (Salmo trutta) is a highly adaptable salmonid species, which through 6 7 natural dispersal or human transport is found in all continents except Antarctica (MacCrimmon et al. 1970). Wide variation in environmental conditions and food availability 8 9 influences the physiology of individuals, and further determine whether they migrate to sea (anadromous brown trout, hereafter referred to as sea trout) or remain freshwater resident 10 (Forseth et al. 1999, Wysujack et al. 2009, Archer et al. 2020). After leaving freshwater, sea 11 trout display plasticity in migratory tactics, with some individuals using nearshore and 12 estuarine habitats, while others use marine areas more than 500 km away from their natal 13 watercourse (Thorstad et al. 2016, Birnie-Gauvin et al. 2019b). However, variation in 14 15 migration patterns and life history strategies is not fully understood, thus limiting our understanding of ecological and evolutionary dynamics of sea trout populations (Birnie-16 Gauvin et al. 2019b, Ferguson et al. 2019). Understanding the drivers of marine migration 17 behaviour is crucial for evaluating susceptibility of sea trout to large scale climate change, 18 19 and to human induced stressors that can vary both temporally and spatially in coastal zone 20 ecosystems (Thorstad et al. 2015, Nevoux et al. 2019). In general, migration is regarded as a 21 biological phenomenon that is particularly sensitive to environmental change and anthropogenic disturbance (Wilcove & Wikelski 2008) such that it is important to understand 22 23 how different taxa respond to such challenges (Lennox et al. 2016). Energy status is known to impact migratory strategies of individual trout 24

25 (Cucherousset et al. 2005, Boel et al. 2014, Bordeleau et al. 2018). For mature sea trout,

reproduction is energetically expensive (Lien 1978, Jonsson & Jonsson 2005), and so they 1 must recondition between spawning events. In watercourses suitable for overwintering, sea 2 3 trout can remain in the spawning river system throughout the winter after autumn spawning (Berg & Berg 1989, Östergren & Rivinoja 2008), but in these oligotrophic systems feeding 4 and growth are usually limited. Post-spawned individuals are therefore in a generally poor 5 nutritional state prior to the seaward migration in the spring (Jonsson & Jonsson 1998, 6 7 Jonsson & Jonsson 2011a). How individual variation in energy status relates to variation in marine migration behaviour of sea trout is not well understood. Body condition factor, which 8 9 is based on the relationship between body length and mass (i.e., the relative stoutness of fish), is commonly used as an index of somatic energy status in salmonids and other fishes, but 10 might not always be a precise predictor of energy status (Weatherley & Gill 1983, Simpson 11 1992, Næsje et al. 2006). In addition to body condition factor, nutritional correlates derived 12 from blood plasma samples can be used to assess the nutritional state of fishes. For 13 salmonids, low levels of plasma triglycerides, total protein, and calcium levels can indicate 14 15 poor nutritional state (Congleton & Wagner 2006), which has previously been observed to promote marine migratory decisions in brown trout and Atlantic salmon (Boel et al. 2014, 16 Bordeleau et al. 2018, Bordeleau et al. 2019). Elevated levels of cortisol possibly due to low 17 food availability, have previously been found to promote earlier seaward migration (Birnie-18 19 Gauvin et al. 2019a). These previous studies suggest that sea trout in poor nutritional state 20 will display more risk-taking behaviour than individuals in better nutritional state in order to 21 compensate for their depleted energy stores.

In this study, we tested the hypothesis that poor nutritional state promotes adopting a more high-risk ocean migratory behaviour in sea trout in terms of timing, duration and migration distance. We used both body condition factor and blood plasma metabolites as measures of individuals' nutritional state. Migration behaviour in sea trout has also been

observed to be influenced by sex (Pemberton 1976, Knutsen et al. 2004, Jensen et al. 2019) 1 and body size (Jensen et al. 2014, Jonsson & Jonsson 2014). Hence, sex determined by 2 genetic analyses and body size were also included in the analyses. We included 286 3 individual sea trout from seven populations in two fjord systems in Northern Norway in this 4 study to test the general hypotheses that poor nutritional state, females, and large size would 5 promote initiation and greater extent of the marine migration. Specifically, we examined 6 7 whether nutritional state, sex, and body size (length) influenced *i*. the tendency to migrate to the sea or remain resident in freshwater and/or estuarine habitats, *ii*. the timing of sea entry, 8 9 *iii.* the duration of the marine residency, and *iv.* the distance moved out to sea from the river where the fish were tagged. 10

11

12 2. MATERIAL & METHODS

13 2. 1 Study site

14 This study was conducted in two Norwegian fjord systems in Nordland County;

15 Skjerstadfjorden (67°N) and Tosenfjorden (65°N) (Figure 1) as part of two larger tracking studies, enabling sampling from seven river systems. In Tosenfjorden, fish were captured and 16 tagged in the period 27 March to 10 June in the Rivers Åbjøra and Urvold (Figure 1). In 17 Skjerstadfjorden, fish were captured and tagged during 28 April to 15 June in the rivers 18 Saltdalselva, Botnvassdraget, Lakselva, Laksåga, and Kosmovatnet (Figure 1). Estuaries were 19 20 defined as the transition zone between the freshwater and marine environment, where the water masses were expected to be brackish throughout the year. For all rivers, this included 21 22 receivers that were deployed less than 600 meters from the river mouth, except for River Saltdalselva where receivers deployed up to 1 km from the river mouth were categorized as 23 estuarine habitats. 24

25

In the Tosenfjorden study area, River Åbjøra has 24 km of river stretch available for

1	anadromous fish, and includes a large estuarine area influenced by the tide (about 1.6 km^2 of
2	tidal affected surface area including shoreline areas inundated at high tide and the lower
3	sections of the river including an estuarine pool), and Lake Åbjørvatnet (surface area of 4.8
4	km ² , 81 meters above the sea). River Åbjøra is regulated for hydropower production and has
5	a minimum discharge of 7 m ³ /s. River Urvold has an average water discharge of 5 m ³ /s, is not
6	developed for hydropower production, and consists of a 200 m steep river stretch from the
7	sea to Lake Urvoldvatnet (surface area of 0.6 km ² , 8.6 meters above sea level). In the inner
8	end of Lake Urvoldvatnet, River Urvold has about 1 km of river stretch available for
9	anadromous fish. The estuary of River Urvold is small (about 0.002 km ² of tidal influenced
10	area inside the littoral zone) because the steep river drains straight into the open fjord.
11	In the Skjerstadfjorden study area, River Saltdalselva is a large river with average
12	discharge of 55 m^3/s and 66 km of river stretch available for anadromous fish. Due to its large
13	size, and relatively slow-running areas in its lower part, River Saltdalselva has a relatively
14	large estuary (about 0.47 km ² tidal influenced areas inside the littoral zone). There is only one
15	lake available for anadromous fish in River Saltdalselva, Lake Vassbotnvatn, which is located
16	in a tributary. River Botnvassdraget has a 500 m steep river stretch to Lake Botnvatnet (12 m
17	above the sea), and continues upstream of the lake, making about 8 km of river stretch
18	available for anadromous fish. River Botnvassdraget's confined estuary covers about 0.002
19	km ² of tidal influenced area. River Lakselva has about 7 km of river stretch with no lakes
20	available to anadromous fish, and has a tidally influenced surface area of about 0.08 $\rm km^2$ in
21	its estuarine area. River Laksåga hasabout 6.5 km of river stretch available for anadromous
22	fish and drains into two large brackish-water lakes influenced by the sea (about 15 km^2
23	surface area). River Laksåga is regulated for hydropower purposes. River Kosmovatnet has
24	about 6 km of river stretch available to anadromous fish, and drains into a brackish-water

lake of about 8 km², separated from the sea by a 1 km narrow channel where the tide governs
 the direction of the current.

3 2.2. Capture and tagging of fish

A total of 286 sea trout, divided into 10 groups based on location and year (Table 1), were 4 captured and tagged with individually coded acoustic transmitters (Thelma Biotel AS; 9 mm 5 6 and 13 mm, expected battery life 10-24 months, tag size depended on body size) during 7 2016-2018. Fish from River Åbjøra, River Kosmovatnet and River Laksåga were caught, in the estuarine parts of the river systems. Fish from River Urvold were caught in Lake Urvold 8 9 (freshwater), except for 13 individuals during spring 2017 (Table 1) that were caught in the 10 river mouth. All fish caught in River Saltdalselva were caught in the river. The fish were captured by angling or by gillnets in the rivers, lakes and/or estuarine areas. Gill nets were 11 12 continuously monitored and quickly tended when a fish was detected to minimize the time 13 fish were entangled in the nets, and the fish were released using scissors to cut the netting to prevent damage to the skin and gills. A non-lethal blood sample was drawn shortly after 14 15 capture (max 5 ml blood per kg body mass, Lawrence et al., 2020). The fish were held in keep nets for up to four hours before they had a transmitters implanted. 16

Prior to tagging, each sea trout was anesthetized for 4 minutes using 0.5 ml⁻¹ 2-17 18 phenoxy-ethanol (EC No. 204-589-7, Sigma-Aldrich, USA). For most fish, tags were inserted through a 1.5-3 cm incision in the body cavity (Cooke et al. 2011, Eldøy et al. 2015). For the 19 fish tagged in River Åbjøra in 2017 (Table 1), the transmitters were externally attached using 20 a wire through the dorsum about 1 cm below the dorsal fin, with a silicone plate between the 21 tag and the fish and a plastic plate on the opposite side of the dorsum to prevent erosion on 22 skin and flesh. The sea trout were subsequently placed in a holding tank until recovery from 23 anaesthesia and released in a slow flowing area as close to the capture location as possible. 24

The experimental procedures were approved by the Norwegian National Animal Research
 Authority (permission number 2015/8518, 1614092 & 18/67706).

3 2.3 Fish tracking

The tagged sea trout were tracked using 74 acoustic receivers in Tosenfjorden and 82
acoustic receivers in Skjerstadfjorden (Vemco Inc. models: VR2, VR2-W and VR2-AR). The
fish in Tosenfjorden were tracked from May 2016 to December 2017, while the fish tagged in
Skjerstadfjorden were tracked from May 2017 to December 2018, although not all receivers
were operative throughout these periods (Figure 1).

9 Tracking data were filtered for false registrations generated by code collisions with 10 simultaneously transmitting tags, or by noise in the environment (Pincock 2012). After empirically assessing the frequency of false detections (i.e., a receiver reported detection of 11 an unused transmitter ID) recorded by each receiver, automated filtering was applied to 16 12 13 receivers in Tosenfjorden, and 4 receivers in Skjerstadfjorden (see Figure 1 for filtered receiver locations). The filter required that a tagged sea trout had to be registered at least two 14 15 times by a receiver within a 10-minute time period to be accepted as a true registration, and resulted in removal of 68 682 of 2 191 047 detections (3.1%) in Tosenfjorden and 2 402 of 16 17 594 345 detections (0.4%) in Skjerstadfjorden. The data were subsequently visually inspected 18 by plotting a timeline of all recordings for each fish, and registrations that did not fit with the 19 overall migration track of each fish were also removed (i.e. detections suggesting unrealistic migration speed and/or passing multiple receiver gates without detection). 20

21 2.4 Processing and analysing blood samples

Blood samples were stored in tubes and immediately placed in an ice water bath for up to 3
hours before being centrifuged at 1163g for 10 min. Blood plasma was flash-frozen in a
liquid nitrogen dry shipper and subsequently stored at -80 °C until biochemical analyses
could be carried out. Plasma triglyceride levels were assessed in duplicates using the

1	manufacturer's suggested protocols with a commercially available colorimetric kit (Cayman
2	Chemical Company, USA). Total plasma protein levels were determined using a Bradford's
3	assay (Bradford 1976, Kruger 2009) using commercial reagents (Bio-Rad Laboratories
4	(Canada) Ltd., Mississauga, ON, Canada). Plasma Ca ²⁺ concentrations were determined
5	using flame spectrophotometry (Varian Spectra AA 220FS, Varian Inc., Palo Alto, CA, USA)
6	for a single replicate. Due to the limited volume of blood that could be taken from each fish
7	(see Lawrence et al. 2020), there was not enough blood for all individuals to run all
8	biochemical assays. Therefore, 214 fish were tested for blood plasma triglycerides, 204 fish
9	were tested for blood plasma protein and 185 fish were tested for blood plasma calcium.
10	2.5 Determining the sex of individuals
11	DNA samples from adipose fin clips taken at the time of tagging and preserved in ethanol
12	were used to genetically determine the sex of the sea trout. DNA was extracted using the
13	QuickExtract kit (Epigen) using the manufacturer's protocol but with extraction volume
14	reduced to 150 μ l. Using 10 μ l reactions of the Qiagen Multiplex PCR kit and Salmo-sdY-F
15	and Samo sdYR primers, PCR amplification was applied to a 200 base pair fragment from the
16	first intron of the male-specific SDY gene (Quéméré et al. 2014). PCR steps for denaturation,
17	annealing and extension were: incubation at 95 °C for 15 min, 11 cycles of touchdown PCR,
18	held at 94 °C for 30 s, 63-52 °C for 30 s, then 72 °C for 1 min followed by 25 cycles at 94 °C
19	for 30 s, 52 °C for 30 s, 72 °C for 1 min and a final extension at 72 °C for 10 min. Sex was
20	determined by running PCR products in 1% agarose gels.
21	2.6 Migrating to the sea vs. remaining in freshwater and estuaries

22 The trout could remain resident in the habitat where they were captured and tagged (river,

23 lake and estuarine habitats), or migrate to the fjord. Individual sea trout were considered sea

24 migrants if they were recorded by any marine receiver except those categorized as estuarine

of the river where they were tagged. Fish that we lost track of shortly after tagging, or which
 showed a "permanent residency" at a particular receiver indicative of mortality or tag loss
 within the receiver's detection range were excluded from statistical analyses (n = 7).

4 2.7 Timing of sea entry and duration of the marine migration

The timing of sea entry was calculated for all individuals that were recorded leaving the 5 6 freshwater and estuarine areas of the watercourse where they were tagged. The timing of sea entry was recorded as the first detection of a tagged fish in the estuary for Rivers Urvold, 7 8 Saltdalselva and Botnvassdraget. In River Åbjøra, the timing of sea entry was defined as the 9 time of the last recorded detection in the river's estuary, provided the fish was subsequently detected by a receiver in the fjord. For the fish tagged in Laksåga, the time of sea entry was 10 defined as the time of first detection by a receiver in the fjord. The different definitions for 11 12 timing of sea entry were due to logistical and hydrologic constraints that required different approaches to receiver deployment in the estuarine areas of the different watercourses. Fish 13 14 tagged in rivers Lakselva and Kosmovatnet were excluded from these analyses due to small 15 sample sizes ($n \leq 2$).

16 Residence time at sea was calculated as the period between the time a fish entered the sea to its last detection in the sea prior to entering the river during the first year of tracking. In 17 River Åbjøra, the sea journey was considered to have ended at the last detection on a receiver 18 deployed in the river mouth, provided that detection was followed by subsequent detections 19 20 within the watercourse. In some cases, sea trout transitioned between freshwater and marine habitats multiple times within a year. Time spent in the freshwater habitat between migrations 21 22 to the sea was not included in the total marine migration time. Fish tagged in Lakselva and Kosmovatnet were excluded from the analyses of residence time at sea due to a low sample 23 size (n = 1). At these sites most tagged fish stayed in the river and estuary. 24

1 2.8 Maximum migration distance at sea

For marine migrants, the maximum migration distance at sea from the river mouth was 2 calculated for the fish from Rivers Åbjøra, Urvold, Saltdalselva, Botnvassdraget and 3 Laksåga. Because receivers were only deployed in the inner part of Skjerstadfjorden in 2016, 4 the fish tagged in Skjerstadfjorden in 2016 were excluded from the analyses of migration 5 distance. The maximum migration distance for each fish was calculated as the distance from 6 7 the receiver deployed closest to the mouth of the river in which the fish were tagged to the furthermost receiver at sea where the fish was recorded. This was done by estimating the 8 9 shortest migration route avoiding land, using the 'costDistance' function in the gdistance R package (van Etten 2018). Fish that were not detected after 20 July of the year of tagging 10 (which was the date corresponding to the upper 95 % percentile for reaching maximum 11 distance for fish observed returning to watercourses), that were not last observed returning to 12 freshwater, or were last observed at the outer arrays of receivers in the fjord were not 13 included in the maximum migration distance analysis (n = 21). 14

15 2.9 Statistical analyses

To test for effects of nutritional state, sex and body size (natural length) on migratory 16 17 behaviour, we used a set of generalized mixed effect models. The behavioural traits that were 18 used as response variables were either binomial (migrated or did not migrate) or continuous and normally distributed (timing, duration and distance of sea migration). Independent 19 variables (fixed explanatory effects) were sex, body size and nutritional state (body condition 20 21 and blood plasma triglycerides). Tagging years were nested within populations (watercourse) and used as random effects. We did not aim to investigate which of the nutritional state 22 23 variables employed were the best proxies. As such, we fitted one full model for each of the nutritional state variables (i.e. the nutritional state variables were used simultaneously in a 24 full model). As all nutritional indicators were found to be correlated, and to simplify the 25

presentation and interpretation of results in this study, modelling using blood plasma protein
 and blood plasma triglycerides was excluded from the manuscript, but can be found in
 Appendices 1 – 8. This approach allowed us to avoid issues with co-linearity among
 nutritional indicators.

All statistical analyses were conducted in R Studio version 1.2.1335 (RStudio Team 5 2019) and R version 3.5.3 (R Core Team 2019) with the 'glmer' function in the 'lme4' R 6 7 package (Bates et al. 2015) for the model with a binomial dependent variable. The 'lme' function in the nlme R package (Pinheiro et al. 2018) was used for models with normally 8 9 distributed dependent variables. Collinearity within models was checked using the 'check collinearity' function in the performance R package (Lüdecke et al. 2020), and 10 collinearity was found to be low (VIF ≤ 1.09). All variables were standardised prior to 11 modelling using the 'scale' function in the R 'base' package. Blood plasma triglycerides 12 values were log-transformed in order to stabilise variance. Body condition factor was 13 calculated from the formula $K = 100 \times \text{mass}$ (g) \times total length (cm)^{-3.028}, as the regression 14 coefficient of the mass-length relationship was 3.028 for the tagged individuals. Model 15 selection was conducted using Akaike information criterion (AIC) (Anderson et al., 2001), 16 with the 'dredge' function in the MuMIn R package (Barton, 2019). In cases when model 17 selection left us with support for multiple alternative models (Δ AIC < 2), conditional model 18 averaging was applied, using all alternative models ($\Delta AIC < 4$) to estimate the coefficients 19 of the explanatory variables. Kruskal-Wallis-tests were applied for comparisons among 20 21 groups of tagged fish (based on tagging year and population) in terms of body size, body condition factor, blood plasma triglycerides, blood plasma protein and blood plasma calcium. 22 23 Spearman correlation tests were applied to test for correlations between pairs of nutritional indicators (body condition factor, blood plasma triglycerides, blood plasma protein and blood 24 plasma calcium). For visualization purposes, linear regression lines were fitted to the 25

1	relationships among nutritional correlates in Figure 3 using the 'geom_smooth' function in				
2	the ggplot2 R package (Wickham 2016). Spearman correlation tests were applied to test for				
3	correlations between behavioural traits (timing of sea entry, marine residency and marine				
4	migration distance).				
5	The raw tracking dataset on individual fish generated and analysed during the current				
6	study is uploaded to the Ocean Tracking Network data system				
7	(www.oceantrackingnetwork.org).				
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9					
10	3. RESULTS				
11	3.1 Characteristics of tagged fish				
12	The results were based on 286 trout (165 females, 121 males, i.e. 58% females, 42% males)				
13	with a body size ranging from 270 to 890 mm (mean = 471 mm , SD = 129 mm) (Figure 2).				
14	The fish were divided into ten groups based on the river and year they were tagged. The				
15	proportion of females within these groups varied between 38% and 78% (Figure 2). There				
16	were significant differences in body condition factor among the groups (Kruskal-Wallis test n				
17	= 286, $P < 0.001$, Figure 2). Average concentrations of the nutritional metabolites (pooled				
18	samples for all fish from all rivers) derived from blood plasma sampling were 0.71 mmol l ⁻¹				
19	triglycerides (SD = 0.85, range 0.004-4.36), 25.03 mg ml ⁻¹ of protein (SD = 5.70, range 8.87-				
20	45.85) and 3.21 mmol l^{-1} of calcium (SD = 0.43, range = 1.94-5.56) among the groups				
21	(Figure 2). However, there were significant differences among the groups in concentrations				
22	of blood plasma triglycerides (Kruskal-Wallis test; $n = 214$, $P < 0.001$), plasma protein ($n =$				
23	204, <i>P</i> < 0.001) and plasma Calcium (<i>n</i> = 185, <i>P</i> < 0.001, Figure 2).				

24 3.2 Correlations between the nutritional indicators

There were significant positive correlations between all the measured variables reflecting 1 nutritional state (Appendix 1). There was a positive correlation between body condition 2 factor and 1) log transformed blood plasma triglycerides (Spearman's correlation; n = 214, r_s 3 = 0.36, P < 0.001), 2) blood plasma protein (n = 204, $r_s = 0.42$, P < 0.001), and 3) plasma 4 calcium (n = 185, $r_s = 0.23$, P < 0.001). There were positive correlations between the log 5 transformed blood plasma triglycerides and both blood plasma protein (n = 194, $r_s = 0.45$, P 6 < 0.001) and blood plasma calcium (n = 178, $r_s = 0.31$, P < 0.001). There was also a positive 7 correlation between blood plasma protein and blood plasma calcium (n = 173, $r_s = 0.60$, P < 0.608 9 0.001).

10 3.3 To migrate to the sea or stay in freshwater and estuaries

Of the 286 tagged trout, 173 individuals migrated to the sea, while 106 individuals remained 11 in freshwater and estuaries during the rest of the year. For 7 individuals, migratory decision 12 13 could not be determined due to absence of detections, or detection records suggesting mortality, tag loss or tag malfunction. Overall, the models (see below) suggested that the fish 14 migrating to the sea had greater body sizes than those remaining in freshwater, that females 15 had a greater tendency to migrate to the sea than males, and that those migrating had lower 16 body condition factors and lower blood plasma triglyceride levels than individuals remaining 17 18 resident (Figure 3 and 4, Appendix 2 and 3).

19 Using body condition factor as the nutritional indicator, a model for migratory 20 decision which included condition factor, sex and body size, and an alternative model 21 excluding sex, were of equally good (Δ AIC = 0.62, Appendix 2). Conditional model 22 averaging indicated that body size had the strongest effect on migratory decision, followed by 23 condition factor and sex, respectively (Figure 3, Appendix 3).

Using plasma triglycerides as nutritional indicator, the model on migratory decision which included sex and body size was the best model (Δ AIC 2.22 to second best model; Appendix 2). The model estimates showed that sex had the strongest effect on migratory
 decision, followed by body size and blood plasma triglycerides, respectively (Figure 4,
 Appendix 3).

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6 3.4 Timing of sea entry

7 Timing of sea entry could be determined for 161 individuals. There were four models with Δ 8 AIC < 2 (Appendix 4) for the timing of sea entry, including the null model, when using body 9 condition factor as a nutritional indicator. Here, the model averaging estimates generally 10 suggested limited effects of all explanatory variables (Figure 3, Appendix 5).

11 Three equally well supported models for timing of sea entry were identified when 12 using plasma triglycerides as the nutritional indicator (Δ AIC = 1.89, Appendix 4). The model 13 averaging estimates indicated that sea trout with higher plasma triglyceride levels entered the 14 sea later in the season, while the effect of sex and body size on timing of sea entry was 15 limited (Figure 4, Appendix 5).

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18 3.5 Marine residence time

The marine residence times could be determined for 74 individuals. Three equally well supported models for the marine residence time were identified when using body condition factor as the nutritional indicator (Δ AIC = 1.65, Appendix 6). The model estimates from conditional model averaging showed that condition factor had the strongest effect on marine residence time, showing that sea trout with lower condition factors spent longer times at sea (Figure 3, Appendix 7).

1	There were four models with Δ AIC < 2 (Appendix 6) for the marine residence time,
2	including the null model, when using body plasma triglycerides as the nutritional indicator.
3	Conditional model averaging for these models showed that the standard errors exceeded the
4	estimated effects of the explanatory variables (Figure 4, Appendix 7).
5	There was a significant negative correlation between timing of sea entry and marine
6	residence time (Spearman's correlation; $n = 80$, $P < 0.001$), where fish entering the sea earlier
7	spent more time at sea.
8	
9	3.6 Maximum migration distance in the sea
10	Maximum migration distance could be determined for 111 individuals. The full model on
11	migration distance, which included condition factor, sex and body size, and an alternative
12	model excluding sex, were equally well supported ($\Delta AIC = 1.97$, Appendix 8). Model
13	averaging showed that larger fish and fish with lower body condition factors migrated further
14	out in the marine habitat, and that sex had limited effect on the migration distance (Figure 3,
15	Appendix 9).
16	The migration distance model selection process where blood plasma triglyceride was
17	used as nutritional indicator found that a model only including body size and an alternative
18	model including body size and sex were equally well supported (Δ AIC = 1.73, Appendix 8).
19	Here, model conditional averaging showed that larger fish tended to migrate further out to sea
20	than smaller fish, while sex and blood triglycerides had limited effect on the migration
21	distance (Figure 4, Appendix 9).
22	The maximum migration distance at sea was not correlated with the timing of sea
23	entry (Spearman's correlation; $n = 167$, $P = 0.89$) or the duration of the marine residency ($n =$
24	69, <i>P</i> = 0.49).
25	

1 4 DISCUSSION

2 Overall, nutritional state, sex and body size (length) influenced the marine migration behaviour of sea trout from the seven study populations in two distinct fjord systems in 3 northern Norway. Sea trout with poor body conditions and low triglyceride levels tended to 4 leave the river and estuaries and migrate to the sea, and individuals with low triglyceride 5 6 levels migrated to the sea earlier. Fish with poor body condition prior to the migration 7 remained at sea for a longer time-period and migrated further out in the fjords than fish in 8 better condition. Although all the nutritional indicators were found to be highly correlated, 9 this study suggests that measuring both body condition and blood plasma metabolites gave a 10 better evaluation of the nutritional state of the individuals and the impacts of nutritional state on behaviour. While body condition results from the balance between energy intake vs. 11 expenditure over time frames of weeks or months, blood plasma triglycerides have previously 12 13 been observed to change in response to food intake over much shorter time scales (Sheridan 1988, Congleton & Wagner 2006). 14

15 Migrating to the marine environment is believed to provide better feeding opportunities and potentially increased growth and reproductive capacity (Klemetsen et al. 16 17 2003, Thorstad et al. 2016) because of the higher productivity in marine habitats than 18 freshwater habitats in high latitude areas (Gross et al. 1988). On the other hand, energetic costs related to migration and osmoregulation, the risk of predation, disease, or other factors 19 that could prevent a migratory individual from returning to freshwater spawning grounds are 20 21 all risk factors presumed to be higher when an individual migrates to the sea (Thorstad et al. 2015, Jensen et al. 2019). In this study, the observed effects of nutritional state on sea trout 22 23 migratory behaviour suggest that individuals in a poor nutritional state tend to engage in riskier migration behaviour than fish in a better nutritional state. The results from this study 24 also suggest that individuals in a poor nutritional state were energetically limited in 25

freshwater, tipping the cost vs. benefit trade-off in favour of migration. In a previous study, 1 Boel et al. (2014) found that sea trout with poor body condition were most likely to migrate 2 3 towards the sea. Davidsen et al. (2014) observed that starved hatchery-reared sea trout released to the wild utilized sea habitat to a greater extent than well fed hatchery fish, which 4 tended to remain in the lower parts of the river and estuarine areas to which they were 5 released. The observed relationship between nutritional state and timing of sea entry is also 6 7 consistent with previous studies on reconditioning post-spawn Atlantic salmon S. salar which exhibited earlier sea entry for individuals in poor body condition (Halttunen et al. 2018, 8 9 Bordeleau et al. 2019). Birnie-Gauvin et al. (2019a) showed that elevated baseline cortisol levels, possibly in response to nutritional need, were associated with earlier migration 10 towards the sea for post-spawned sea trout. Interestingly, we observed a stronger effect of 11 blood plasma triglycerides levels than of body condition factor on the timing of sea entry. 12 This may suggest behavioural response to nutritional need, as acute triglyceride deprivation 13 was a strong predictor for migratory initiation. Alternatively, it may suggest that 14 15 opportunistic feeding in the freshwater or estuarine habitats during early spring may recondition nutritional state and delay the initiation of marine migration. 16

Once migration has occurred, sea trout with poor body condition factors tended to 17 spend more time at sea and migrate further out in the marine habitat, possibly reflecting a 18 19 greater need to recondition compared to individuals with better body condition factors. There 20 was a significant relationship between timing of sea entry and duration of marine residency, 21 where fish that migrated early resided longer at sea. The prolonged residency at sea may enable sea trout to recondition for the next spawning and overwintering season but may also 22 23 include higher risk of mortality as marine habitats often have greater abundance of potential predators. A previous study by Haraldstad et al. (2018) showed that post-spawned sea trout in 24 poor body condition were more likely to skip spawning the following season compared to 25

individuals in better body condition. Bordeleau et al. (2018) reported that the pre-migratory 1 level of blood plasma triglycerides was negatively correlated with the duration of marine 2 residency in veteran sea trout migrants. According to previous studies, sea trout with a low 3 body condition tended to migrate further out to sea compared with individuals in a better 4 body condition (Davidsen et al. 2014, Eldøy et al. 2015, Bordeleau et al. 2018). In the present 5 study, differences in characteristics of the near marine habitats among the multiple sites we 6 7 studied probably impacted how far the sea trout from the various rivers needed to migrate to meet their metabolic demands. No correlation was found between migration distance and 8 9 timing of marine entry or marine residence time.

As expected, there were significant positive correlations among all measured 10 nutritional indicators. A previous lab experiment by Congleton et al. (2006) documented low 11 12 levels of blood plasma triglycerides, blood plasma protein and blood plasma calcium in starved juvenile salmonids. Overall nutritional state is determined by net differences over 13 periods of weeks or months between energy intake and energy expenditure (Congleton & 14 15 Wagner 2006). Poor nutritional state could likely be explained by limited feeding while overwintering, and for fish that reproduced, also by the energy expenditure during spawning 16 the previous autumn (Bordeleau et al. 2018). The energy investment in spawning can be 17 substantial for brown trout (Jonsson & Jonsson 2011b). In Lake Vangsvatnet in Norway, 18 19 Jonsson and Gravem (1985) documented that immature migrants fed little while in freshwater 20 and that mature migrants stopped feeding after the spawning season.

Although the nutritional status of most fish in the study was poor, there was large variation in nutritional state both among individuals and groups of fish. There may be several reasons for this, including differences in nutritional state after the previous growth season, energy investment in previous spawning, overwintering conditions, metabolic rate and feeding activity (Midwood et al. 2015, Auer et al. 2016). Some individuals had elevated

nutritional indicators indicative of a better nutritional status. These individuals mainly 1 belonged to the groups of fish tagged in the estuarine habitats of River Åbjøra, River 2 3 Lakselva and the Kosmovatnet watercourse. For these fish, elevated blood plasma triglyceride level was the most obvious signal. This might suggests that the fish had started 4 feeding prior to tagging, because triglycerides become elevated a few hours after feeding 5 (Sheridan 1988), and a previous laboratory experiment documented that blood plasma 6 7 triglycerides recovered quickly when refeeding began after a starvation period in rainbow trout Oncorhynchus mykiss (Congleton & Wagner 2006). Common for these groups of tagged 8 9 fish was that they were captured and tagged in lower parts of watercourses which have relatively large estuarine areas likely suitable for opportunistic feeding during early spring. 10

Females were more likely to migrate to the sea than males, instead of remaining in the 11 freshwater and estuarine areas of the river where they were tagged. Previous studies with this 12 13 species have also noted that females are more likely to migrate than males (Pemberton 1976, Knutsen et al. 2004, Jensen et al. 2019). In a study in Tosenfjorden, Bordeleau et al. (2018) 14 15 found that females in the Åbjøra watercourse were more likely to leave the estuarine areas than males. This is likely caused by a greater benefit of increased feeding opportunities for 16 17 females than males due to the strong correlation between female body size and the number of 18 eggs the female can produce (Elliott 1995). Sexual bias in migratory behaviours is a wellknown phenomenon that has previously been observed in a range of salmonid species 19 (reviewed by Dodson et al. 2013). 20

The models in the present study provided limited support for the influence of sex on migration timing, duration and distance migrated at sea. Some previous studies have reported that male sea trout tended to migrate earlier (Jensen 1968, Östergren & Rivinoja 2008), while others at different sites have suggested an earlier migration timing for females (Berg & Berg 1989). Berg & Berg (1989) also observed that females had a longer duration of marine

residency than males. Bordeleau *et al.* (2018) found that females migrated further from the
river than males and were more likely to migrate to the outer fjord areas of Tosenfjorden.
Although the reasons female and males differed in their migration patterns among these
different sites remain obscure, it suggests that a combination of local environmental
conditions and population characteristics may plays an important role for the trade-off
mechanisms shaping the migratory decisions of individuals within sea trout populations. This
plasticity is one of the reasons the species has been so successful.

Larger fish of both sexes were more likely to migrate to the sea and migrated greater 8 9 distances at sea than smaller fish. These tendencies are likely driven by the need of larger individuals to find more prey of larger size than the smaller fish, that larger fish are less 10 susceptible to predation than smaller fish, and possibly because larger fish may be more 11 powerful swimmers (Dill 1983, Klemetsen et al. 2003). Individual sea trout tend to repeat 12 their migratory patterns among successive years (Eldøy et al. 2019) although some studies 13 suggest that iteroparous salmonids may reduce their migration distances as they become 14 larger and older (Svärdson & Fagerström 1982, Bond et al. 2015). The earlier seaward 15 migration of large fish observed in the present study is similar to the timing observed in 16 previous studies (Pemberton 1976, Bohlin et al. 1996, Jonsson & Jonsson 2009). The positive 17 correlation we noted between body size and the duration of the marine migration is consistent 18 19 with previous work (Eldøy et al. 2015). However, the tendency we found for larger fish to 20 migrate further out to sea compared to smaller individuals has only been noted in a few of the previous studies on this species (e.g. Berg & Berg 1989, Jensen et al. 2014, Jonsson & 21 Jonsson 2014). 22

In conclusion, despite the large individual and among-group variation observed in both nutritional state and migratory behaviour, this study showed that sex, body size and premigratory nutritional state strongly influenced the migratory patterns of sea trout. Anadromy

is considered a quantitative threshold trait, where environmental thresholds for triggering 1 behavioural responses are genetically determined (Ferguson 2006, Ferguson et al. 2019). 2 3 Previous studies have suggested that the migratory behaviour of brown trout is a continuum of behavioural responses to the environmental cues experienced by the individuals in coastal 4 trout populations (Cucherousset et al. 2005, Boel et al. 2014, Villar-Guerra et al. 2014). 5 However, the importance of different factors affecting the pre-migratory nutritional state, and 6 7 the influence of carry-over effects are poorly understood (O'Connor et al. 2014). For example, it is unknown how the success of a previous feeding migration interacts with 8 9 spawning investment and over-wintering conditions to determine the nutritional state and subsequent marine migrations of post-spawned, veteran sea trout migrants (Bordeleau 2019). 10 As shown by Jensen et al. (2020), life history patterns or decisions adopted early in life may 11 12 persist throughout an individual's lifetime, and significantly affect the animal's lifetime fitness. Jensen et al. (2020) showed that early migrants continued to migrate early throughout 13 their life time, had better growth, and a larger lifetime fecundity. This suggests that 14 individuals developing under favourable conditions will gain fitness benefits throughout their 15 lifetime. Jensen et al. (2020) therefore concluded that individuals that experience 16 environmental conditions as juveniles in freshwater and/or with genes that contribute to a 17 large smolt size and early smolt migration may benefit preferentially from growth 18 19 opportunities in the sea, and the benefits of the early adoption of anadromy enables them to 20 continue with early and longer migrations during following years. However, the fact that sea 21 trout populations do not evolve to contain exclusively early migrants highlights again that there are costs that counterbalance the strategy. 22

While at sea, sea trout commonly reside in habitats heavily affected by human activity (Nevoux et al. 2019). Salmon lice infestation related to open cage farming of Atlantic salmon was recently evaluated as the biggest threat for Norwegian sea trout stocks (Norwegian

Scientific Advisory Committee for Atlantic Salmon 2019). Serra-Llinares et al. (2020) 1 documented that sea trout infested with salmon lice altered their migration behaviour and 2 3 experienced increased mortality. The results of the present study, where fish in poor nutritional state seemed to migrate to the sea earlier and spent more time at sea, suggest that 4 fish in poor nutritional state may display behavioural patterns that make them especially 5 vulnerable to such negative anthropogenic factors at sea. This is both because their longer 6 stay at sea increases the risk of being infested by salmon lice, and because they migrate to 7 areas with high salinity favourable for sea lice instead of remaining in brackish water areas 8 9 where the lice do not survive well. The links between migration behaviour, human induced stressors and reproductive success throughout the lifetime of sea trout remain obscure. Future 10 studies examining the link between marine migration behaviour and reproductive investment 11 over consecutive years are therefore advocated. 12

13

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24

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- 11

12	Archer LC, Hutton SA, Harman L, McCormick SD and others (2020) Food and temperature stressors						
13	have opposing effects in determining flexible migration decisions in brown trout (Salmo						
14	trutta). Global Change Biology 26:2878-2896						
15	Auer SK, Salin K, Anderson GJ, Metcalfe NB (2016) Flexibility in metabolic rate and activity level						
16	determines individual variation in overwinter performance. Oecologia 182:703-712						
17	Bates D, Maechler M, Bolker B, Walker S (2015) Fitting linear mixed-effects models using Ime4.						
18	Journal of Statistical Software 67:1-48						
19	Berg OK, Berg M (1989) The duration of sea and freshwater residence of the sea trout, Salmo trutta,						
20	from the Vardnes River in northern Norway. Environmental Biology of Fishes 24:23-32						
21	Birnie-Gauvin K, Flávio H, Kristensen ML, Walton-Rabideau S and others (2019a) Cortisol predicts						
22	migration timing and success in both Atlantic salmon and sea trout kelts. Scientific Reports						
23	9:1-9						
24	Birnie-Gauvin K, Thorstad EB, Aarestrup K (2019b) Overlooked aspects of the Salmo salar and Salmo						
25	trutta lifecycles. Reviews in Fish Biology and Fisheries 29:749-766						
26	Birnie-Gauvin K, Lennox RL, Guglielmo CG, Teffer AK and others (In Press) The value of experimental						
27	approaches in migration biology. Physiological and Biochemical Zoology 00:000-000						
28	Boel M, Aarestrup K, Baktoft H, Larsen T and others (2014) The physiological basis of the migration						
29	continuum in brown trout (Salmo trutta). Physiological and biochemical zoology 87:334-345						
30	Bohlin T, Dellefors C, Faremo U (1996) Date of smolt migration depends on body-size but not age in						
31	wild sea-run brown trout. Journal of Fish Biology 49:157-164						
32	Bond MH, Miller JA, Quinn TP (2015) Beyond dichotomous life histories in partially migrating						
33	populations: cessation of anadromy in a long-lived fish. Ecology 96:1899-1910						
34	Bordeleau X, Davidsen JG, Eldøy SH, Sjursen AD, Whoriskey FG, Crossin GT (2018) Nutritional						
35	correlates of spatiotemporal variations in the marine habitat use of brown trout (Salmo						
36	trutta) veteran migrants. Canadian Journal of Fisheries and Aquatic Sciences 75:1-11						
37	Bordeleau X (2019) The post-spawning ecology of iteroparous salmonids: Basis of variability in						
38	migratory behaviour and survival, ecological importance and conservation implications.						
39	Ph.D. Thesis. 168 p. Halifax: Department of Biology, Dalhousie University.						

1 Bordeleau X, Hatcher BG, Denny S, Whoriskey FG, Patterson DA, Crossin GT (2019) Nutritional 2 correlates of the overwintering and seaward migratory decisions, and long-term survival of 3 post-spawning Atlantic salmon. Conservation Physiology 7:1-13 4 Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of 5 protein utilizing the principle of protein-dye binding. Analytical Biochemistry 72:248-254 6 Chapman BB, Hulthén K, Brodersen J, Nilsson PA, Skov C, Hansson L-A, Brönmark C (2012) Partial 7 migration in fishes: causes and consequences. Journal of Fish Biology 81:456-478 8 Congleton JL, Wagner T (2006) Blood-chemistry indicators of nutritional status in juvenile salmonids. 9 Journal of Fish Biology 69:473-490 10 Cooke SJ, Crossin GT, Patterson DA, English KK and others (2005) Coupling non-invasive physiological 11 assessments with telemetry to understand inter-individual variation in behaviour and 12 survivorship of sockeye salmon: development and validation of a technique. Journal of Fish 13 Biology 67:1342-1358 14 Cooke SJ, Hinch SG, Farrell AP, Patterson DA and others (2008) Developing a mechanistic 15 understanding of fish migrations by linking telemetry with physiology, behavior, genomics 16 and experimental biology: An interdisciplinary case study on adult Fraser River sockeye 17 salmon. Fisheries 33:321-339 18 Cooke SJ, Murchie S, Mc Connachie S, Goldberg T (2011) Standardized surgical procedures for the 19 implantation of electronic tags in key Great Lakes fishes. Technical Report. Great Lakes 20 Fishery Commission, Ann Arbor, MI. 44 pp. 21 Crossin GT, Cooke SJ, Goldbogen JA, Phillips RA (2014) Tracking fitness in marine vertebrates: current 22 knowledge and opportunities for future research. Marine Ecology Progress Series 496:1-17 23 Crossin GT, Heupel MR, Holbrook CM, Hussey NE and others (2017) Acoustic telemetry and fisheries 24 management. Ecological Applications 27:1031-1049 25 Cucherousset J, Ombredane D, Charles K, Marchand F, Baglinière J-L (2005) A continuum of life 26 history tactics in a brown trout (Salmo trutta) population. Canadian Journal of Fisheries and 27 Aquatic Sciences 62:1600-1610 28 Davidsen JG, Daverdin M, Sjursen AD, Rønning L, Arnekleiv JV, Koksvik JI (2014) Does reduced 29 feeding prior to release improve the marine migration of hatchery brown trout Salmo trutta 30 smolts? Journal of Fish Biology 85:1992-2002 31 Dill LM (1983) Adaptive flexibility in the foraging behavior of fishes. Canadian Journal of Fisheries 32 and Aquatic Sciences 40:398-408 33 Dingle H, Drake VA (2007) What Is Migration? BioScience 57:113-121 34 Dingle H (2014) Migration: the biology of life on the move, Vol. Oxford University Press, New York 35 Dodson JJ, Aubin-Horth N, Thériault V, Páez DJ (2013) The evolutionary ecology of alternative 36 migratory tactics in salmonid fishes. Biological Reviews 88:602-625 37 Eldøy SH, Davidsen JG, Thorstad EB, Whoriskey F and others (2015) Marine migration and habitat 38 use of anadromous brown trout Salmo trutta. Canadian Journal of Fisheries and Aquatic 39 Sciences 72:1366 - 1378 40 Eldøy SH, Bordeleau X, Crossin GT, Davidsen JG (2019) Individual repeatability in marine migratory 41 behavior: A multi-population assessment of anadromous brown trout tracked through 42 consecutive feeding migrations. Frontiers in Ecology and Evolution 7:1-12 43 Elliott JM (1995) Fecundity and egg density in the redd for sea trout. Journal of Fish Biology 47:893-44 901 45 Ferguson A (2006) Genetics of sea trout, with particular reference to Britain and Ireland. In: Harris G, 46 Milner N (eds) Sea trout: biology, conservation and management. Blackwell publishing Ltd, 47 Oxford, p 155-182 48 Ferguson A, Reed TE, Cross TF, McGinnity P, Prodöhl PA (2019) Anadromy, potamodromy and 49 residency in brown trout Salmo trutta: the role of genes and the environment. Journal of 50 Fish Biology 95:692-718

1 Forseth T, Nesje TF, Jonsson B, Hårsaker K (1999) Juvenile migration in brown trout: a consequence 2 of energetic state. Journal of Animal Ecology 68:783-793 3 Gross MR, Coleman RM, McDowall RM (1988) Aquatic productivity and the evolution of diadromous 4 fish migration. Science 239:1291-1293 5 Halttunen E, Gjelland K-Ø, Hamel S, Serra-Llinares R-M and others (2018) Sea trout adapt their 6 migratory behaviour in response to high salmon lice concentrations. Journal of Fish Diseases 7 41:953-967 8 Haraldstad T, Höglund E, Kroglund F, Lamberg A, Olsen EM, Haugen TO (2018) Condition-dependent 9 skipped spawning in anadromous brown trout (Salmo trutta). Canadian Journal of Fisheries 10 and Aquatic Sciences 75:2313-2319 11 Jachowski DS, Singh NJ (2015) Toward a mechanistic understanding of animal migration: 12 incorporating physiological measurements in the study of animal movement. Conservation 13 Physiology 3:1-12 14 Jensen AJ (1968) Sea trout (Salmo trutta L.) of the River Istra, western Norway. Report: Institute of 15 Fresh-water Research, Drottningholm 48, 187-213. 16 Jensen AJ, Finstad B, Fiske P (2019) The cost of anadromy: marine and freshwater mortality rates in 17 anadromous arctic char and brown trout in the arctic region of Norway. Canadian Journal of 18 Fisheries and Aquatic Sciences 76:2408-2417 19 Jensen AJ, Finstad B, Fiske P, Diserud OH, Thorstad EB (2020) Repeatable individual variation in 20 migration timing in two anadromous salmonids and ecological consequences. Ecology and 21 Evolution 10:11727-11738 22 Jensen JLA, Rikardsen AH, Thorstad EB, Suhr AH, Davidsen JG, Primicerio R (2014) Water 23 temperatures influence the marine area use of Salvelinus alpinus and Salmo trutta. Journal 24 of Fish Biology 84:1640-1653 25 Jonsson B, Gravem FR (1985) Use of space and food by resident and migrant brown trout, Salmo 26 trutta. Environmental Biology of Fishes 14:281-293 27 Jonsson B, Jonsson N (1993) Partial migration: niche shift versus sexual maturation in fishes. Reviews 28 in Fish Biology and Fisheries 3:348-365 29 Jonsson B, Jonsson N (2005) Lipid energy reserves influence life-history decision of Atlantic salmon 30 (Salmo salar) and brown trout (S. trutta) in fresh water. Ecology of Freshwater Fish 14:296-31 301 32 Jonsson B, Jonsson N (2009) Migratory timing, marine survival and growth of anadromous brown 33 trout Salmo trutta in the River Imsa, Norway. Journal of Fish Biology 74:621-638 34 Jonsson B, Jonsson N (2011a) Habitats as template for life histories. In: Jonsson B, Jonsson N (eds) 35 Ecology of Atlantic Salmon and Brown Trout: Habitat as a template for life histories. Springer 36 Netherlands, Dordrecht, p 1-21 37 Jonsson B, Jonsson N (2011b) Maturation and Spawning. In: Ecology of Atlantic Salmon and Brown 38 Trout: Habitat as a template for life histories. Springer Netherlands, Dordrecht, p 327-414 39 Jonsson B, Jonsson N (2014) Naturally and hatchery produced European trout Salmo trutta: Do their 40 marine survival and dispersal differ? Journal of Coastal Conservation 18:79-87 41 Jonsson N, Jonsson B (1998) Body composition and energy allocation in life-history stages of brown 42 trout. Journal of Fish Biology 53:1306-1316 43 Klemetsen A, Amundsen P-A, Dempson JB, Jonsson B, Jonsson N, O'Connell MF, Mortensen E (2003) 44 Atlantic salmon Salmo salar L., brown trout Salmo trutta L., and Arctic charr Salvelinus 45 alpinus (L.): a review of aspects of their life histories. Ecology of Freshwater Fish 12:1-59 46 Knutsen JA, Knutsen H, Olsen EM, Jonsson B (2004) Marine feeding of anadromous Salmo trutta 47 during winter. Journal of Fish Biology 64:89-99 48 Kruger NJ (2009) The Bradford method for protein quantitation. In: Walker JM (ed) The Protein 49 Protocols Handbook. Humana Press, Totawa, p 17-24 50 Lawrence MJ, Raby GD, Teffer AK, Jeffries KM and others (2020) Best practices for non-lethal blood 51 sampling of fish via the caudal vasculature. Journal of Fish Biology 97:4-15

1	Lennox RJ, Chapman JM, Souliere CM, Tudorache C, Wikelski M, Metcalfe JD, Cooke SJ (2016)						
2	Conservation physiology of animal migration. Conservation Physiology 4:1-15						
3	Lien L (1978) The energy budget of the brown trout population of øvre Heimdalsvatn. Ecography						
4	1:279-300						
5	Lüdecke D, Makowski D, Waggoner P, Patil I (2020) Performance: Assessment of regression models						
6	performance. R package version 0.4.5. <u>https://CRAN.R-project.org/package=performance</u> .						
7	MacCrimmon HR, Marshall TL, Gots BL (1970) World distribution of brown trout, Salmo trutta:						
8	Further observations. Journal of the Fisheries Research Board of Canada 27:811-818						
9	Midwood JD, Larsen MH, Boel M, Aarestrup K, Cooke SJ (2015) An experimental field evaluation of						
10	winter carryover effects in semi-anadromous brown trout (Salmo trutta). Journal of						
11	Experimental Zoology Part A: Ecological Genetics and Physiology 323:645-654						
12	Nevoux M, Finstad B, Davidsen JG, Finlay R and others (2019) Environmental influences on life						
13	history strategies in partially anadromous brown trout (Salmo trutta, Salmonidae). Fish and						
14	Fisheries 20:1051-1082						
15	Norwegian Scientific Advisory Committee for Atlantic Salmon (2019) Classification of the state of 430						
16	Norwegian sea trout populations. Temarapport no. 7, Norwegian Scientific Advisory						
17	Committee for Atlantic Salmon, Trondheim						
18	Næsje TF, Thorstad EB, Forseth T, Aursand M, Saksgård R, Finstad AG (2006) Lipid class content as an						
19	indicator of critical periods for survival in juvenile Atlantic salmon (Salmo salar). Ecology of						
20	Freshwater Fish 15:572-577						
21	O'Connor CM, Norris DR, Crossin GT, Cooke SJ (2014) Biological carryover effects: linking common						
22	concepts and mechanisms in ecology and evolution. Ecosphere 5:1-11						
23	Pemberton R (1976) Sea trout in North Argyll Sea lochs, population, distribution and movements.						
24	Journal of Fish Biology 9:157-179						
25	Pincock DG (2012) False detections: what they are and how to remove them from detection data. 11						
26	p. Halifax: AMIRIX Systems Inc. DOC-004691-03.						
27	Pinheiro J, Bates D, DebRoy S, Sarkar D, Team RC (2018) nlme: Linear and nonlinear mixed effects						
28	models. R package version 3.1-137, URL: <u>https://CRAN.R-project.org/package=nlme</u> .						
29	Quéméré E, Perrier C, Besnard A-L, Evanno G, Baglinière J-L, Guiguen Y, Launey S (2014) An improved						
30	PCR-based method for faster sex determination in brown trout (Salmo trutta) and Atlantic						
31	salmon (Salmo salar). Conservation Genetics Resources 6:825-827						
32	R Core Team (2019) R: A language and environment for statistical computing. Vienna: R Foundation						
33	for Statistical Computing. URL: <u>http://www.R-project.org/</u> .						
34	RStudio Team (2019) RStudio: Integrated development for R. Boston: RStudio Inc. URL:						
35	http://www.rstudio.com/.						
36	Serra-Llinares RM, Bøhn T, Nilsen R, Karlsen Ø and others (2020) Increased mortality and altered						
37	behaviour of sea trout (Salmo trutta) post-smolts infested with salmon lice (Lepophtheirus						
38	salmonis). Marine Ecology Progress Series 635:151-168						
39	Sheridan MA (1988) Lipid dynamics in fish: aspects of absorption, transportation, deposition and						
40	mobilization. Comparative Biochemistry and Physiology Part B: Comparative Biochemistry						
41	90:679-690						
42	Simpson AL (1992) Differences in body size and lipid reserves between maturing and nonmaturing						
43	Atlantic salmon parr, Salmo salar L. Canadian Journal of Zoology 70:1737-1742						
44	Svärdson G, Fagerström Å (1982) Adaptive differences in the long-distance migration of some trout						
45	(Salmo trutta L.) stocks. Report Instute of Freshwater Research Drottningholm 60:51-80						
46	Thorstad EB, Todd CD, Uglem I, Bjørn PA and others (2015) Effects of salmon lice Lepeophtheirus						
47	salmonis on wild sea trout Salmo trutta — a literature review. Aquaculture Environment						
48	Interactions 7:91-113						
49	Thorstad EB, Todd CD, Uglem I, Bjørn PA and others (2016) Marine life of the sea trout. Marine						
50	Biology 163:1-47						

- 1 van Etten J (2018) gdistance: Distances and routes on geographical grids. R package version 1.2-2. 2 URL: https://CRAN.R-project.org/package=gdistance. 3 Villar-Guerra D, Aarestrup K, Skov C, Koed A (2014) Marine migrations in anadromous brown trout 4 (Salmo trutta). Fjord residency as a possible alternative in the continuum of migration to the 5 open sea. Ecology of Freshwater Fish 23:594-603 6 Weatherley AH, Gill HS (1983) Protein, lipid, water and caloric contents of immature rainbow trout, 7 Salmo gairdneri Richardson, growing at different rates. Journal of Fish Biology 23:653-673 8 Wickham H (2016) ggplot2: Elegant graphics for data analysis. New York: Springer Publishing. 9 Wilcove DS, Wikelski M (2008) Going, going, gone: Is animal migration disappearing. PLOS Biology 10 6:e188 11 Wysujack K, Greenberg LA, Bergman E, Olsson IC (2009) The role of the environment in partial 12 migration: food availability affects the adoption of a migratory tactic in brown trout Salmo 13 trutta. Ecology of Freshwater Fish 18:52-59 14 Zera AJ, Harshman LG (2001) The physiology of life history trade-offs in animals. Annual Review of 15 Ecology and Systematics 32:95-126 16 Östergren J, Rivinoja P (2008) Overwintering and downstream migration of sea trout (Salmo trutta L.) kelts under regulated flows—northern Sweden. River Research and Applications 24:551-17 18 563 19 20 21
- 22

- 1 Table 1: Description of fjord, watercourse, tracking year, number of individuals, date
- 2 oftagging and mean $(\pm SD)$ sea trout body size (mm) and body mass (g)
- 3

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		Tracking			Body size	Body
Site	Watercourse	year	Tagging date	n	(mm)	Mass (g)
			21/05/2016-		405±62	705±269
Tosenfjorden	Åbjøra	2016	26/05/2016	23	(290-520)	(240-1280)
			27/03/2017–		451±58	847±318
Tosenfjorden	Åbjøra	2017	29/03/2017	37	(350-560)	(400-1540)
			04/05/2016-		415±81	639±401
Tosenfjorden	Urvoll	2016	10/06/2016	24	(270-620)	(160-1940)
			19/04/2017–		463±98	949±816
Tosenfjorden	Urvoll	2017	14/05/2017	46	(330-730)	(290-4550)
			01/05/2016-		574±164	2068±1812
Skjerstadfjorden	Botnvassdraget	2016	20/05/2016	21	(300-850)	(200-5800)
			11/05/2017–		379±71	531±284
Skjerstadfjorden	Kosmovatnet	2017	15/06/2017	19	(275-470)	(180-990)
			30/05/2017–		384±78	538±447
Skjerstadfjorden	Laksåga	2017	31/05/2017	34	(290-630)	(180-2590)
			02/05/2017–		442±53	902±377
Skjerstadfjorden	Lakselva	2017	09/05/2017	13	(360-520)	(410-1580)
			28/04/2016-		520±157	1574±1611
Skjerstadfjorden	Saltdalselva	2016	30/04/2016	40	(360-860)	(350-5600)
			01/05/2018-		637±113	2459±1626
Skjerstadfjorden	Saltdalselva	2018	10/05/2018	29	(450-890)	(830-8400)



Figure 1: Map over the study areas in the two fjords Skjerstadfjorden (upper) and
Tosenfjorden (lower), with tagging sites and receiver positions indicated. "Automated
filtering" indicates receivers where automatic data filtering was applied to remove false
detections. Light blue water surface indicates watercourses. Purple to deep blue surface
colour indicate the depth of estuarine and marine habitats.



1

Figure 2: Sex (a), body size (mm) (b), body condition factor (c), blood plasma triglycerides (d), blood plasma proteins (e) and blood plasma calcium (f) for the study's groups of tagged fish (location and year of tagging for the groups indicated on the x-axes). The stacked bar plots (a) shows the proportion of males and females in each group. The box plots show the interquartile range (boxes), median (horizontal line in boxes), the 5th and 95th percentiles (whiskers) and outliers (dots), with number of individuals in each group denoted at the top of the panels.

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Figure 3: Estimated effect of body condition factor, sex (male) and body size on the decision
to migrate to sea (a), timing of sea entry (b), marine residence time (c) and migration distance

4 at sea (d). The bar plots show the estimated parameter coefficients and their standard error

5 (whiskers) for the best fitted model (Δ AIC < 2) or from conditional model averaging

6 (including models with $\Delta AIC < 4$) where model selection identified multiple models of

7 similar support.





Figure 4: Estimated effect of plasma triglycerides, sex (male) and body size on the decision to
migrate to sea (a), timing of sea entry (b), marine residence time (c) and migration distance at
sea (d). The bar plots show the estimated parameter coefficients and their standard error

- 6 (whiskers) for the best fitted model ($\Delta AIC < 2$) or from conditional model averaging
- 7 (including models with Δ AIC < 4) where model selection identified multiple models of
- 8 similar support.
- 9
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