1 Mamiellophyceae shift in seasonal predominance in the Baltic Sea

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12 ABSTRACT: The green algae Mamiellophyceae are a group of marine picoeukaryotes. We

13 studied the succession of Mamiellophyceae in the Baltic Sea water column and ice from

14 autumn to summer, using the hypervariable V4 region of the 18S ribosomal RNA gene. We

15 show that species of Mamiellophyceae shift in seasonal predominance and that different

16 species characterize sea ice, under-ice water and the water column in the Baltic Sea.

17 KEY WORDS: 18S rRNA gene · Hypervariable V4 region · Green algae · Picoeukaryotes ·
 18 Succession

19 **1. Introduction**

Mamiellophyceae, a class of green algae, includes 3 described orders: Mamiellales, 20 Dolichomastigales and Monomastigales (Marin & Melkonian 2010). Dolichomastigales and 21 Monomastigales host many lineages, but they are not abundant in marine waters (Monier et 22 al. 2016). In contrast, the most common Mamiellophyceae in coastal waters—species from 23 the Mamiellales genera Ostreococcus, Bathycoccus and Micromonas-may contribute 24 significantly to the primary production of picoeukaryotes (Worden et al. 2004, Tragin & 25 Vaulot 2018). This has raised research interest in Mamiellophyceae recently, and with the 26 help of molecular methods, the contributions and seasonal patterns of different lineages have 27 become possible to investigate in more detail (e.g. Foulon et al. 2008, Demir-Hilton et al. 28 29 2011).

30 Mamiellophyceae as a class does not show preferences to environmental conditions globally (Tragin & Vaulot 2018), and Mamiellophyceae biogeography is driven largely by 31 geographical location rather than water depth (Monier et al. 2016). For example, species of 32 Ostreococcus are globally distributed but are not always found together with usually co-33 occurring Bathycoccus and Micromonas (Demir-Hilton et al. 2011), and Ostreococcus species 34 are absent in Arctic waters (Tragin & Vaulot 2019). Ostreococcus is, however, present in 35 adjacent seasonally sea ice covered areas, for example the Baltic Sea and the White Sea 36 37 (Majaneva et al. 2012, Belevich et al. 2018).

At the species and genetic strain level, Mamiellophyceae occupy different ecological niches. For example, within the morphospecies *Micromonas pusilla*, different genetic lineages recently divided into several species (Simon et al. 2017) show shifts in abundance along local and basin-wide environmental gradients (Foulon et al. 2008). Similarly, *Ostreococcus* strains show differences in distribution, and co-occurrence at the same geographical location is rare 43 (Demir-Hilton et al. 2011). Metagenomes of the genetically less variable *Bathycoccus* 44 *prasinos* indicate the same (Vannier et al. 2016).

In the Baltic Sea, species of Monomastix, Dolichomastix, Mamiella, Mantoniella and 45 Micromonas are reported (Hällfors 2004, Majaneva et al. 2012). Mantoniella sp. appears 46 characteristic for ice, while *B. prasinos* is characteristic for under-ice water (Majaneva et al. 47 2017). However, there are no studies on seasonal patterns or environmental preferences of 48 Mamiellophyceae in the Baltic Sea. Here, we take advantage of our bi-weekly sampling of 49 50 cold water season protists and describe Mamiellophyceae succession based on V4 reads of the 18S ribosomal RNA (18S rRNA) gene in Baltic Sea water and sea ice during the period from 51 autumn to summer. 52

53 2. Materials and Methods

54 Samples were collected from 2 coastal brackish water (salinity 3.5–6) sites in the northwestern Gulf of Finland, Baltic Sea. Sampling was carried out from 8 October 2012 to 8 55 May 2013, with extra samples on 3 September 2012 and 1 July 2013. The sampling protocol, 56 DNA extraction, PCR setup and sequencing are described in Enberg et al. (2018) and in 57 Supplement 1 at www.int-res.com/articles/suppl/mXXXpXXX_supp/. Enberg et al. (2018) 58 outlined the eukaryotic communities using microscopy and a 310 bp long fragment of the V4 59 region of the 18S rRNA gene (hereafter short V4). Here, we focus on Mamiellophyceae short 60 V4 reads and a longer fragment of the V4 region that was amplified using 574*F and 1132R 61 (Hugerth et al. 2014) primers (hereafter long V4). This long V4 was approximately 540 bp 62 long. The raw reads are available at the ENA SRA repository with the study names 63 64 PRJEB21047, PRJEB23628 and PRJEB25089. The paired-end sequenced short and long V4 reads were merged using the -65

fastq_mergepairs command in vsearch v.2.6.2 (Rognes et al. 2016). The quality filtering using 66 the -fastq_filter command discarded reads shorter than 320 bases (short V4) and 550 bases 67 (long V4), reads longer than 380 bases (short V4) and 610 bases (long V4), reads with 68 69 ambiguous bases and reads with over 1 maximum expected error. Primer sequences were removed using the command trim.seqs in mothur v.1.39.5 (Schloss et al. 2009). Exact 70 duplicates were removed using the command -derep_fulllength in vsearch. Chimeric reads 71 were searched using the command -uchime denovo in vsearch. A read was treated as 72 chimeric and removed if its abundance was 5 times lower than its assumed parental reads. 73 Operational taxonomic units (OTUs) were searched using the option fastidious in swarm 74 75 v.2.1.6 (Mahé et al. 2015). The abundance of each OTU in each sample was searched using the -usearch_global command (-id 0.0) in vsearch. 76

The OTUs were assigned taxonomically searching all OTUs first against the NCBI non-redundant nucleotide database (short V4: 9 February 2018, long V4: 10 February 2018) and second against the SILVA_132_SSURef database (Quast et al. 2013), using BLAST v.2.6.0+ (Zhang et al. 2000). All OTUs affiliated with Mamiellophyceae were selected for the subsequent analyses. Read abundance was normalized to the total number of

82 Mamiellophyceae reads in each sample.

To place the OTUs phylogenetically, we took all available sequences of described Mamiellophyceae species together with some relevant uncultured sequences (e.g. Mamiellophyceae DSGM-81) and 6 Pyrenomonadaceae sequences as an outgroup from the SILVA and NCBI Nucleotide databases (23 January 2019). The datasets were combined and aligned with the MAFFT online service (Katoh et al. 2017), and the alignment was cut to the length of the long V4. Bayesian phylogenetic analysis was performed with MrBayes v.3.2.6

(Ronquist et al. 2012). Two independent runs with 4 Markov chains and 1500000 generations

were carried out. We did not choose the model prior to the analysis but sampled across the
general time-reversible model space with gamma-distributed rate variation across sites and a
proportion of invariable sites. The resulting estimates (e.g. tree topology) were posterior
probability weighted averages of the models. The scripts for methods are provided in
Supplement 2. The reads and read abundance are provided in Table S1 in Supplement 3.

95 3. Results and Discussion

In the dataset, 59 short and 46 long V4 Mamiellophyceae OTUs were present, and 96 after manually checking the alignment and combining identical short and long V4 OTUs, 59 97 OTUs were used in the analyses. Mamiellophyceae OTU richness was significantly lower in 98 sea ice than in under-ice water and the water column (Kruskal-Wallis $\chi^2 = 13.7$, p = 0.003, 99 followed by pairwise comparisons using the Mann-Whitney U-test). Mamiellophyceae 100 contributed 3.3% of the total abundance of taxonomically assigned OTUs, and they were 101 present in all samples. The lowest Mamiellophyceae read abundance was in the Krogarviken 102 103 April ice sample (<0.1% of total abundance), and the highest contribution was in the Storfjärden March bottom water sample (17% of total abundance). Mamiellophyceae read 104 abundance was significantly lower in sea ice and under-ice water than in the water column 105 (Kruskal-Wallis $\chi^2 = 26.3$, p < 0.001, followed by pairwise comparisons using the Mann-106 107 Whitney U-test).

The Bayesian phylogenetic tree (Fig. 1) resembled the latest phylogenies of 108 Mamiellophyceae (Marin & Melkonian 2010, Monier et al. 2016, Simon et al. 2017, Yau et 109 al. unpubl.; preprint doi:10.1101/506915), and the OTUs represented all 3 described orders 110 with 35 Mamiellales OTUs, 23 Dolichomastigales OTUs and 1 Monomastigales OTU (Fig. 111 1). OTUs that referred to the genus *Mamiella* were not found. In accordance with an earlier 112 global study (Monier et al. 2016), Dolichomastigales were diverse but rare in the Baltic Sea, 113 114 and the abundant OTUs (>100 reads in total, 22 OTUs) were mainly from Mamiellales. The abundant OTUs were associated with Bathycoccus (86923 reads in total, 3 OTUs), 115 Ostreococcus (59313 reads, 3 OTUs), Micromonas (36640 reads, 4 OTUs) and Mantoniella 116 (25982 reads, 7 OTUs). The rest of the abundant OTUs were affiliated with DSGM-81 117

Mamiellophyceae (1147 reads, 3 OTUs) and with *Crustomastix* (362 reads, 2 OTUs) from
 Dolichomastigales.

- Different Mamiellophyceae OTUs characterized sea ice, under-ice water and the water column (generalized discriminant analysis of 4 first principal coordinates analysis axes, analysis based on $\ln[x + 1]$ transformed Bray-Curtis dissimilarities, squared correlations 0.80
- and 0.58, p < 0.001, misclassification error 43.9%).

The Mamiellophyceae OTU associated with an uncultured Baltic Sea ice clone FN690723 characterized sea ice (**Fig. 2a,d**, Otu07). This species is clearly sea ice associated, and it is present in the sea ice of the Gulf of Bothnia and the Gulf of Finland in the Baltic Sea (Majaneva et al. 2012) and in the White Sea (Belevich et al. 2018). Otu07 and the uncultured

Baltic Sea ice clones form a clade of their own, basal to the *Mantoniella* and *Micromonas*

- 129 clade in our Bayesian phylogenetic tree (Fig. 1) but which are classified as *Mantoniella* in
- 130 Tragin & Vaulot (2019). *Mantoniella squamata* (Otu14) was the most abundant *Mantoniella*
- species in the water column in September and October, as was *M. beaufortii* (Otu04) in
- 132 November to May and OTUs close to *M. antarctica* (Otu21, Otu22) in July (**Fig. 3c**). *M.*

beaufortii and several *Crustomastix* OTUs characterized under-ice water. The presence of

134 freshwater *Crustomastix* species in low abundance in under-ice water is an indication of river

discharge under ice in the area (Kaartokallio et al. 2007).

- 136 The abundant *Bathycoccus*, *Ostreococcus* and *Micromonas* species characterized the
- 137 water column but shifted in predominance during the sampling season (Fig. 2c,f,g).
- 138 Ostreococcus mediterraneus (Otu02) was the most abundant species in October to December,
- although *Bathycoccus* and *Micromonas* species had a relatively high abundance as well.
- 140 Otu01, closely related to *Bathycoccus prasinos*, predominated distinctly in January to April,
- 141 *Micromonas polaris* (Otu06) in May and *M. commoda* (Otu03) in July (Fig. 2c,f).

The 4 species of *Micromonas* alternated in dominance in the water column: the predominant species was *M. pusilla* (Otu09) in October surface water and *M. bravo* (Otu10) in bottom water, *M. polaris* (Otu06) in January to May and *M. commoda* (Otu03) in summer and early autumn water (Fig. 3a,b). These shifts in predominance are in line with the thermal niches of *Micromonas* species (Demory et al. 2019).

The 2 abundant Ostreococcus OTUs—O. mediterraneus (Otu02) and the Otu05 basal 147 to the clade of O. tauri and O. lucimarinus—co-occurred in autumn, and both OTUs were 148 practically absent after March (Fig. 2). The co-occurrence is in accordance with the earlier 149 findings of co-occurring coastal Ostreococcus strains (Demir-Hilton et al. 2011) and the 150 absence of Ostreococcus in cold polar waters (Tragin & Vaulot 2019). B. prasinos had a 151 similar occurrence in the Baltic Sea, being present and abundant only in the autumn (e.g. Fig. 152 2c). The 2 Ostreococcus OTUs and B. prasinos appear to be characteristic species of the 153 autumn water community (Enberg et al. 2018). 154

The most abundant OTU in our dataset (Otu01, 1 base difference to clone FN690721 155 from Majaneva et al. 2012) was present in all samples. It is closely related to *B. prasinos* (Fig. 156 1), which is the only described species within the genus currently and whose cultures show 157 identical 18S rRNA gene sequences but differing genomes (Vannier et al. 2016, Tragin & 158 Vaulot 2019). It is beyond this note, but the uncorrected interspecific 18S rRNA sequence 159 divergence within the other Mamiellales genera is 1.1 to 5.6%, and the 1.1% uncorrected 160 interspecific divergence of Otu01 and B. prasinos implies that Otu01 could be an uncultured, 161 undescribed species of *Bathycoccus*. 162

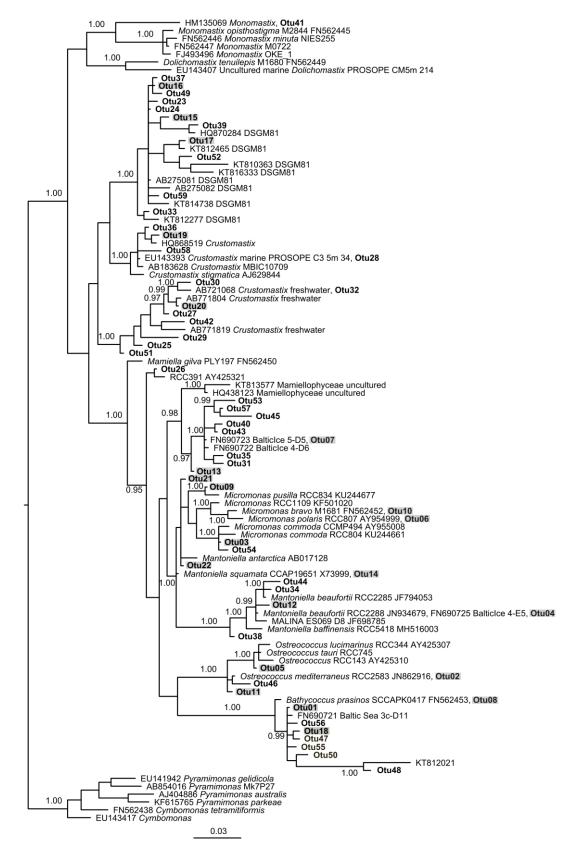
Here, we have shown that species of Mamiellophyceae shift in seasonal predominance in a coastal brackish water locality in the Baltic Sea and that different species characterize sea ice, under-ice water and the water column during the sea ice covered season. Our findings support the earlier research showing geographic niche partition within Mamiellophyceae (Foulon et al. 2008, Demir-Hilton et al. 2011, Monier et al. 2016) and show that the niche partition holds also for populations in the water and sea ice of the Baltic Sea.

Acknowledgements. Walter and Andrée de Nottbeck Foundation and University of Helsinki 3
 yr research grants provided funding to carry out the field and laboratory analysis for this
 study.

LITERATURE CITED 172 <irn>Belevich TA, Ilyash LV, Milyutina IA, Logacheva MD, Goryunov DV, Troitsky AV 173 (2018) Photosynthetic picoeukaryotes in the land-fast ice of the White Sea, Russia. 174 Microb Ecol 75:582-597 doi:10.1007/s00248-017-1076-x</jrn> 175 <irn>Demir-Hilton E, Sudek S, Cuvelier ML, Gentemann CL, Zehr JP, Worden AZ (2011) 176 Global distribution patterns of distinct clades of the photosynthetic picoeukaryote 177 Ostreococcus. ISME J 5:1095-1107 doi:10.1038/ismej.2010.209</jrn> 178 <irn>Demory D, Baudoux AC, Monier A, Simon N and others (2019) Picoeukaryotes of the 179 *Micromonas* genus: sentinels of a warming ocean. ISME J 13:132–146 180 doi:10.1038/s41396-018-0248-0</jrn> 181

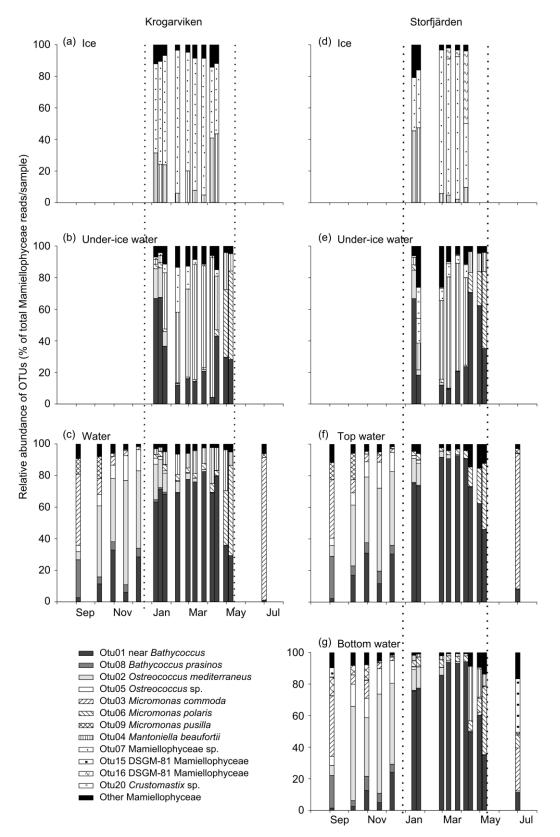
182	<jrn>Enberg S, Majaneva M, Autio R, Blomster J, Rintala JM (2018) Phases of microalgal</jrn>
183	succession in sea ice and the water column in the Baltic Sea from autumn to spring. Mar
184	Ecol Prog Ser 599:19–34 doi:10.3354/meps12645
185	<jrn>Foulon E, Not F, Jalabert F, Cariou T, Massana R, Simon N (2008) Ecological niche</jrn>
186	partitioning in the picoplanktonic green alga <i>Micromonas pusilla</i> : evidence from
187	environmental surveys using phylogenetic probes. Environ Microbiol 10:2433–2443
188	doi:10.1111/j.1462-2920.2008.01673.x
189	<jrn>Hällfors G (2004) Checklist of Baltic Sea phytoplankton species (including some</jrn>
190	heterotrophic protistan groups). Baltic Sea Environ Proc 95. Baltic Marine Environment
191	Protection Commission, Helsinki Commission, Helsinki
192	<jrn>Hugerth LW, Muller EEL, Hu YOO, Lebrun LAM and others (2014) Systematic design</jrn>
193	of 18S rRNA gene primers for determining eukaryotic diversity in microbial consortia.
194	PLOS ONE 9:e95567 doi:10.1371/journal.pone.0095567
195	<jrn>Kaartokallio H, Kuosa H, Thomas DN, Granskog MA, Kivi K (2007) Biomass,</jrn>
196	composition and activity of organism assemblages along a salinity gradient in sea ice
197	subjected to river discharge in the Baltic Sea. Polar Biol 30:183–197 <u>doi:10.1007/s00300-</u>
198	006-0172-z
199	<jrn>Katoh K, Rozewicki J, Yamada KD (2017) MAFFT online service: multiple sequence</jrn>
200	alignment, interactive sequence choice and visualization. Brief Bioinform
201	30:bbx108doi:10.1093/bib/bbx108
202	<jrn>Mahé F, Rognes T, Quince C, de Vargas C, Dunthorn M (2015) Swarm v2: highly-</jrn>
203	scalable and high-resolution amplicon clustering. PeerJ 3:e1420
204	doi:10.7717/peerj.1420
205	<jrn>Majaneva M, Rintala JM, Piisilä M, Fewer PD, Blomster J (2012) Comparison of</jrn>
206	wintertime nano-sized eukaryotic communities in the Baltic Sea ice and water, based on
207	sequencing of the 18S rRNA gene. Polar Biol 35:875–889 doi:10.1007/s00300-011-1132-
208	9
209 210 211	<jrn>Majaneva M, Blomster J, Müller S, Autio R and others (2017) Sea-ice eukaryotes of the Gulf of Finland, Baltic Sea, and evidence for herbivory on weakly shade-adapted ice algae. Eur J Protistol 57:1–15 doi:10.1016/j.ejop.2016.10.005</jrn>
212	<jrn>Marin B, Melkonian M (2010) Molecular phylogeny and classification of the</jrn>
213	Mamiellophyceae class. nov. (Chlorophyta) based on sequence comparisons of the
214	nuclear- and plastid-encoded rRNA operons. Protist 161:304–336
215	doi:10.1016/j.protis.2009.10.002
216	<jrn>Monier A, Worden AZ, Richards TA (2016) Phylogenetic diversity and biogeography of</jrn>
217	the Mamiellophyceae lineage of eukaryotic phytoplankton across the oceans. Environ
218	Microbiol Rep 8:461–469 doi:10.1111/1758-2229.12390
219 220 221	<jrn>Quast C, Pruesse E, Yilmaz P, Gerken J and others (2013) The SILVA ribosomal RNA gene database project: improved data processing and web-based tools. Nucleic Acids Res 41:D590–D596 doi:10.1093/nar/gks1219</jrn>
222	<jrn>Rognes T, Flouri T, Nichols B, Quince C, Mahé F (2016) VSEARCH: a versatile open</jrn>
223	source tool for metagenomics. PeerJ 4:e2584 doi:10.7717/peerj.2584
224	<jrn>Ronquist F, Teslenko M, van der Mark P, Ayres DL and others (2012) MrBayes 3.2:</jrn>
225	efficient Bayesian phylogenetic inference and model choice across a large model space.
226	Syst Biol 61:539–542 doi:10.1093/sysbio/sys029

- <jrn>Schloss PD, Westcott SL, Ryabin T, Hall JR and others (2009) Introducing mothur:
 open-source, platform-independent, community-supported software for describing and
 comparing microbial communities. Appl Environ Microbiol 75:7537–7541
 doi:10.1128/AEM.01541-09</jrn>
- <jrn>Simon N, Foulon E, Grulois D, Six C and others (2017) Revision of the genus
 Micromonas Manton et Parke (Chlorophyta, Mamiellophyceae), of the type species M.
 pusilla (Butcher) Manton & Parke and of the species M. commoda van Baren, Bachy and
- Worden and description of two new species based on the genetic and phenotypic characterization of cultured isolates. Protist 168:612–635
- 236 doi:10.1016/j.protis.2017.09.002</jrn>
- </l
- <jrn>Tragin M, Vaulot D (2019) Novel diversity within marine Mamiellophyceae
 (Chlorophyta) unveiled by metabarcoding. Sci Rep 9:5190 doi:10.1038/s41598-019-41680-6</jrn>
- <jrn>Vannier T, Leconte J, Seeleuthner Y, Mondy S and others (2016) Survey of the green
 picoalga *Bathycoccus* genomes in the global ocean. Sci Rep 6:37900
 doi:10.1038/srep37900</jrn>
- <jrn>Worden AZ, Nolan JK, Palenik B (2004) Assessing the dynamics and ecology of marine
 picophytoplankton: the importance of the eukaryotic component. Limnol Oceanogr
 49:168–179 doi:10.4319/lo.2004.49.1.0168</jrn>
- <jrn>Zhang Z, Schwartz S, Wagner L, Miller W (2000) A greedy algorithm for aligning DNA sequences. J Comput Biol 7:203–214 doi:10.1089/10665270050081478</jrn>



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Fig. 1. Bayesian phylogenetic tree of Mamiellophyceae. Posterior probabilities >0.95 are shown on the branches. The operational taxonomic units (OTUs) of the current research are named Otu01, Otu02, etc., in decreasing order by their abundance in the total dataset, and are in **bold**. OTUs with >100 reads in total have a grey background



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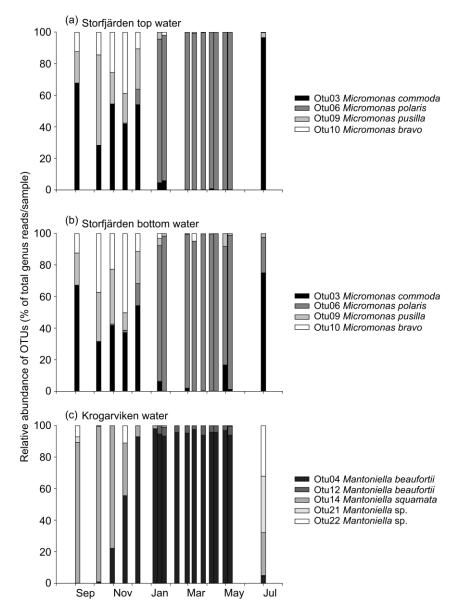
Fig. 2. Relative read abundance of the most abundant Mamiellophyceae operational

taxonomic units in sea ice, under-ice water and in the water column at Krogarviken and

258 Storfjärden throughout the sampling season. The water depth was 3 m at Krogarviken and ca.

30 m at Storfjärden. The top water was 0 to 15 m and the bottom water 15 to 30 m at

260 Storfjärden. The time between dotted lines represents the ice-covered season



262 Fig. 3. Relative read abundance of *Micromonas* and *Mantoniella* operational taxonomic units

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in the water column