



Relationship between lipid and protein oxidation in fish

Journal:	<i>Aquaculture Research</i>
Manuscript ID	ARE-RA-18-Aug-788.R3
Manuscript Type:	Review Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Hematyar, Nima; Jihoceska univerzita v Ceskych Budejovicich Ustav akvakultury, Faculty of Fisheries and Protection of Waters, South Bohemian Research Center of Aquaculture and Biodiversity of Hydrocenoses, Research Institute of Fish Culture and Hydrobiology Rustad , Turid ; FACULTY OF NATURAL SCIENCES, Department of Biotechnology and Food Science Sampels, Sabine; Swedish University of Agricultural Sciences, Molecular Sciences Kastrup Dalsgaard , Trine ; Faculty of Science and Technology, Food Science
Keywords:	Lipid oxidation, protein oxidation, interaction, free radicals, fish

SCHOLARONE™
Manuscripts

1 Relationship between lipid and protein oxidation in fish: A Review

2 Running head: Relationship between lipid and protein oxidation

3 Nima Hematyar ^{a,*}, Turid Rustad ^b, Sabine Sampels ^{a, c}, Trine Kastrup Dalsgaard ^d

4

5 ^a University of South Bohemia in České Budějovice, Faculty of Fisheries and Protection of
6 Waters, South Bohemian Research Center of Aquaculture and Biodiversity of Hydrocenoses,
7 Institute of Aquaculture, Husova tř. 458/102, 370 05 České Budějovice, Czech Republic

8 ^b Department of Biotechnology and Food Science, Norwegian University of Science and
9 Technology, Trondheim, Norway

10 ^c Swedish University of Agricultural Sciences, Department of Molecular Sciences, PO Box
11 7015, 75007 Uppsala, Sweden

12 ^d Department of Food Science, Faculty of Science and Technology, Aarhus University, Blichers
13 Allé 20, DK-8830 Tjele, Denmark

14

15 **ORCID** for authors:

16 Nima Hematyar: orcid.org/0000-0002-7414-2672

17 Turid Rustad: orcid.org/0000-0002-8972-6347

18 Sabine Sampels: orcid.org/0000-0003-1695-5939

19 Trine Kastrup Dalsgaard: orcid.org/0000-0002-5635-4102

20

21 *corresponding author: Nima Hematyar, Institute of Aquaculture, Husova tř. 458/102, 370 05

22 České Budějovice, Czech Republic; Hematyar@frov.jcu.cz; Tel. 00420 38 777 4660

23 Abstract

24 Reactive oxygen species (ROS) are generated in all aerobic organisms. Free radicals are highly
25 reactive ROS that cause damage to biological materials. Fish is rich in polyunsaturated fatty
26 acids, and hence, very prone to lipid peroxidation. Both lipid and protein oxidations are
27 important for quality loss during storage of fish, with high impact on taste and texture. Also,
28 there are interactions between protein and secondary lipid oxidation products (aldehydes) that
29 occur in foods because the oxidation products from one reaction can further react with both
30 lipids and proteins, respectively. This review focuses on the mechanisms and pathways of the
31 lipid and protein oxidation and their possible relationship. Additionally, the target amino acids
32 and final impacts of this relationship were considered. We propose that the products of lipid
33 oxidation promote protein oxidation in fish rather than the other way around specially, during
34 frozen storage, while during post-mortem changes protein oxidation dominate. Finally, it
35 seems that, secondary products of lipid oxidation might have more impact on the functionality
36 of proteins from both Michael addition and Schiff base reaction rather than lipid
37 hydroperoxides and lipid radical transfer.

38

39 Keywords: Lipid oxidation, protein oxidation, interaction, free radicals, fish

40 **1. Introduction**

41 Oxidation in food is a process that concerns both lipids and proteins by the effect of reactive
42 oxygen species (ROS) (Bernardini et al., 2011). Oxidative modifications can cause numerous
43 functional consequences and lead to changes in food texture, water holding capacity (WHC),
44 digestibility and juiciness in fish meat (Baron, Kjaersgard, Jessen, & Jacobsen, 2007). Also,
45 Sarma, Reddy, & Srikar (2000) found a negative correlation between lipid oxidation
46 parameters and functional properties of the protein in Indian oil sardine, indicating their
47 interdependence. However, still little is known about the kinetics of protein oxidation, the
48 interaction of protein and lipid oxidation and the subsequent impact on muscle food quality
49 (Baron et al., 2007; Kjaersgard & Jessen, 2004).

50 For many years, the focus in food was on lipid oxidation, mainly because of its effect on taste
51 and shelf life, but also because the lipid oxidation products can more easily be measured
52 (hydroperoxides, hexanal, and MDA) than protein oxidation product and therefore have been
53 detected and correlated to off-flavour in food. Especially in fish, which is rich in n-3
54 polyunsaturated fatty acids (PUFA), there is a high risk of quality loss due to oxidation
55 (Jeremiah, 2001; Medina, Gallardo, & Aubourg, 2009). Beside lipid oxidation, the oxidation
56 of proteins can also cause quality changes in the fish fillet. In general, the same factors causing
57 lipid oxidation will also cause protein oxidation. The kinetics of protein and lipid oxidation
58 with respect to the generation of hydroperoxide and carbonyls are quite similar but the
59 diversity of the protein oxidation products are more complex than those of lipid oxidation due
60 to more reactive targets in the proteins. The progress of initial oxidative reactions in foods is
61 enhanced by the interactions between the proteins and lipids due to the similarity of the
62 oxidation reactions. The development of lipid and protein oxidation can occur in parallel or
63 independently, but often there are interactions between them (Zhang, Xiao, & Ahn, 2013) .
64 Several authors reported a good correlation between oxidised products of lipid and protein in

65 fish (Soyer, & Hultin, 2000) beef (Estévez, & Cava, 2004) and chicken (Soyer, Özalp, Dalmış,
66 & Bilgin, 2010), which indicates that protein and lipid oxidation start together and can interact
67 with each other. In contrast to those conclusions, Aalhus & Dugan (2014) have suggested that
68 the products of lipid oxidation can promote protein oxidation since lipid oxidation may start
69 earlier.

70 In the initiation of lipid oxidation products can react with proteins and vice versa, but the most
71 well-described reaction is the reaction between secondary lipid oxidation products and
72 primary amino groups on proteins (Schauenstein, & Esterbauer, 1978; Burcham, & Kuhan,
73 1996).

74 As touched upon above, the reaction between secondary lipid oxidation products and amino
75 acids also takes place in fish. Saturated lipid aldehydes become covalently bound to
76 susceptible and functional groups of proteins like N-terminal groups of Tyr, Asp, Arg and Met
77 (Metz et al., 2004), the ϵ -NH₂ group of Lys, and Cys-SH (Metz et al., 2004) through a Schiff
78 base formation while unsaturated aldehydes also can react via Michael addition (Cai,
79 Bhatnagar, & Pierce, 2009). By these interactions proteins, hydrophobicity and aggregation
80 are increased in fish flesh. In addition, the progress of lipid oxidation in fish muscle strongly
81 correlates to the formation of metHb/metMb and also leads to the reduction of extractability
82 haem proteins. For example, during frozen storage of herring fillets, peroxide value (PV) and
83 thiobarbituric acid reactive substances (TBARS) were increased while the amount of haem-
84 proteins decreased (Jonsson et al., 2007).

85 This review aims to give an overview of the lipid and protein oxidation, the known pathways
86 and correlations between lipid and protein oxidation in raw and post-mortem conditions and
87 also the effects of oxidation with a special focus on fish fillet quality.

88

89 **2. Mechanisms of Lipid and Protein oxidation**

90 In order to investigate lipid-protein correlations in fish fillet, the mechanisms of lipid and
91 protein oxidation should be understood in depth. Particularly, the control of possible catalysts
92 has a key role because free radical chain reactions can be swiftly boosted by catalysts.

93 Lipid oxidation is a key factor, leading to a declined in food quality, predominantly of those
94 food products, which contain high amounts of unsaturated FAs (Secci, & Parisi, 2016).

95 Generally, the potential initiators of lipid oxidation can also initiate the protein oxidation
96 (Xiong, 2000). However, the mechanisms, pathways, and also the products of protein
97 oxidation are different (Stadtman, 2006). The functional groups, which are located in the
98 amino acid residues side chain and the peptide backbone are the targets for ROS.

99 The effect of lipid oxidation in muscle food (Estévez, & Cava, 2004) and seafood (Hultin,
100 1993; Secci, & Parisi, 2016; Mariutti, & Bragagnolo, 2017) and protein oxidation (Estévez,
101 2015; Soladoye, Juarez, Aalhus, Shand, & Estevez, 2015) has been reviewed extensively
102 whereas correlation between the lipid and protein oxidation has not really been touched upon
103 in a review.

104

105 **2.1. Lipid oxidation**

106 Unpleasant off-flavours and formation of volatiles can develop due to the oxidation of PUFA.

107 The most important factors which can affect rancidity in fish muscle are the high content of
108 PUFA and also the presence of pro-oxidants, especially the parts containing heme groups
109 (Richard, & Hultin, 2002).

110 Autoxidation enzymatic catalysed oxidation and photosensitized oxidation are three major
111 mechanisms in lipid oxidation.

112 Autoxidation in meat and fish can be initiated by light, heat, and the presence of metal ions
113 and radicals (Sampels, 2013). Autoxidation leads to the formation of the primary oxidation
114 products, hydroperoxides (ROOH). Once the oxidation process has started, a cascade of

115 reactions will occur with each new molecule increasing the reaction speed and variability
116 (Heinonen, Meyer, & Frankel, 1998). ROOH from lipid oxidation decompose easily at high
117 temperature or in the presence of metals to secondary products such aldehydes, short chain
118 hydrocarbons, alcohols, esters, acids, and ketones (Choe, & Min, 2006). Finally, termination
119 products can be crosslinking products e.g. where two radicals react with each other, thereby
120 terminating the chain reaction caused by radical reaction.

121 The atmospheric triplet oxygen is the most common oxygen species involved in the oxidation
122 of lipids but in the photosensitized reaction, both singlet and triple oxygen are two types of
123 oxygen have more interactions with lipids (Foote, 1976). Singlet oxygen has been suggested
124 to react 1,450 faster with linoleic acids than triplet oxygen (Rawls, & Santen, 1970). Singlet
125 oxygen is needed to start the so called type II photosensitized oxidation process, which can
126 react with unsaturated lipids (Foote, 1976) but compared to type I reaction the type II reaction
127 generate different types of hydroperoxide products. The greater importance is the energy of
128 oxygen, which has a significant impact on the initiation of the oxidation reaction.

129 Metal catalysis can be considered as an important lipid oxidation reaction in fish muscle, this
130 means that after post-slaughter processes, the released haem iron (ferrous (+2)) is converted
131 to the ferric (+3) and start the autoxidation progress. Both haem proteins (haemoglobin (Hb)
132 and myoglobin) can increase the lipid oxidation in fish fillet and other muscle foods (Kanner,
133 1994). Richards & Hultin (2002) reported that the blood residue in fish fillet, catalyse lipid
134 oxidation during storage of fatty fish and bleeding was also shown to retard lipid oxidation of
135 minced trout muscle during storage at +2°C. Maqsood & Benjakul (2011) showed that the
136 initiation and propagation of lipid oxidation in the un-bled samples compared with the bled
137 samples were more marked.

138 In addition, there are several enzymes in fish, which are capable to catalyse lipid oxidation
139 such as, lipoxygenases and myeloperoxidases. The first one exists in fish skin and gills and

140 can catalyse the incorporation O₂ into an unsaturated fatty acid and generate ROOH, and the
141 second one initiate lipid oxidation in the presence of halides and hydrogen peroxide. During
142 the processing, this can be critical because the lipid, oxygen and blood interaction will
143 increased (Mozuraityte, Kristinova, Rustad, & Storro, 2016).

144

145 **2.2. Protein oxidation**

146 Foods are constantly exposed to ROS and this will not only cause lipid oxidation but also
147 protein oxidation. However, for several decades proteins were ignored as a target for ROS, in
148 the opposed of lipid oxidation, which was investigated very deeply. In contrast to lipids,
149 proteins are complex macromolecules arranged in 3D-structures, and when oxidation occurs
150 it leads to various changes in the proteins, both chemical changes on individual amino acids
151 (Davies, Delsignore, & Lin, 1987) such as Met (Dalsgaard et al., 2010) Tyr (Dalsgaard,
152 Nielsen, Brown, Stadler, & Davies, 2011) His and Trp (Dalsgaard, Nielsen, & Larsen, 2007)
153 Lys and Arg (Dalsgaard et al., 2007; Lund, Heinonen, Baron, & Estevez, 2011) but also
154 structural changes may occur (Davies, & Delsignore, 1987). Because protein's functions are
155 very specific, oxidative modifications can cause numerous functional consequences and lead
156 to changes in e food texture, water holding capacity (WHC), digestibility and juiciness (Baron
157 et al., 2007; Sarma et al., 2000). Protein oxidation may be caused directly by ROS and reactive
158 nitrogen species (RNS) or indirectly as the result of reactions with products from lipid
159 oxidation with reducing sugars or carbohydrate (Lund et al., 2011). On the other hand,
160 nucleophilic reaction on the carbonyl groups of free sugars and aldehyde on the side chain of
161 amino acids can lead to production of Schiff base products (Dalsgaard, Nielsen, & Larsen,
162 2006, Dalsgaard et al., 2007). Generally, the pathways of protein carbonylation can be divided
163 to direct oxidation, metal-catalyzed oxidation, reaction with free sugars and also lipid
164 peroxidation products (Michael adducts) (Fedorova, Bollineni, & Hoffmann, 2014). Due to

165 side chain oxidation of some amino acids (Arg, Lys, His, and Pro) or backbone oxidation of
166 Asp, Pro and Glu residues carbonylation can be considered as a permanent and destructive
167 (Hawkins, & Davies, 2001) indicator of protein oxidation (Nystrom, 2005).

168 Protein oxidation is initiated when a hydrogen atom is abstracted from the protein to generate
169 a carbon-centered radical (C•) and in the presence of oxygen is converted to an alkylperoxy
170 radical (COO•). The following reaction of (COO•) with hydrogen atom abstraction from
171 another molecule leads to alkyl peroxide (COOH) formation. Subsequent reactions lead to the
172 formation of the alkoxy radical (CO•) and hydroxyl compounds (COH). In addition, two
173 carbon-centered radicals (alkyl-radical such as ethane, methane, and propane) can react with
174 each other in the absence of oxygen to generate carbon–carbon cross-linked derivatives
175 (Papuc, Goran, Predescu, & Nicorescu, 2017). The termination reaction, which in relation to
176 fish and meat quality may affect tenderness (Soladoye et al., 2015).

177 Additionally, aromatic amino acids like Tyr, Trp, His, and Phe are very susceptible to
178 oxidation (Hawkins, & Davies, 2001). During oxidation of aromatic amino acids, phenoxy
179 radicals will be formed from tyrosine and their metabolites, dityrosine and other products, are
180 generated. This occurs especially when tyrosine is close to tyrosol radicals and also tyrosyl
181 radicals which cannot be repaired because they are not reductants (Aeschbach, Amado, &
182 Neukom, 1976).

183 On the other hand, the amino acids, which are aliphatic but do not contain sulfur like Pro or
184 Arg are oxidized via another way. In this group, oxidation takes place by hydrogen abstraction
185 at the α -carbon generating a carbon centered radical (Stadtman, 1993). This reaction occurs at
186 the terminal amine of the Lys side-chain and sites distance from deactivate α -amino group.
187 The generated product will be different depending on the presence or absence of oxygen
188 (Stadtman, 1993).

189 Protein oxidation via metal-catalysed cleavages main reason of oxidative damage *in vivo*
190 systems like fish (Moller, Rogowska-Wrzesinska, & Rao, 2011). Also, in the post mortem
191 fish muscle.

192 In some proteins, metal ion-catalysed oxidation systems can easily oxidize the side-chains of
193 amino acid residues. Lys, Pro, and Arg are the most important targets of metal catalysed
194 oxidation, (Amici, Levine, Tsai, & Stadtman, 1989). Requena, Chao, Levine, & Stadtman,
195 (2001) reported glutamic and then amino adipic semialdehydes are respectively very important
196 carbonyl products in metal catalysed oxidation systems. For example, Lys can be a target for
197 Fe (II)-catalysed oxidation system. In the mentioned mechanism, the chelate form of Fe (II)
198 and amino group of Lys can generate hydroxyl radical by a reaction between Lys and hydrogen
199 peroxide. Hydroxyl radical preferentially attacks to the Lys moiety to convert Lys to a 2-
200 amino-adipic-semialdehyde residue. The similar reaction with other amino acids by Fe (II)
201 can produce carbonyl derivatives. The site-specific mechanism is supported by the
202 confirmation that the metal-catalysed reactions are prevented by catalase but not by $\cdot\text{OH}$
203 scavengers, maybe because the scavengers are not be able to compete with the “caged”
204 reaction at the metal binding site between amino acids and $\cdot\text{OH}$.

205 Amadori products, which have been generated via Schiff base products rearrangement are
206 highly susceptible to the degradation and metal-catalysed oxidation reaction. Consequences
207 of the reactions lead to protein cross-linked adducts and Maillard reaction products (Lund et
208 al., 2011).

209 The major mechanisms in protein oxidation are still unclear because there are only a few
210 methods available to evaluate protein oxidation mechanisms, but the number of reaction
211 products is large. Detection of carbonyl groups, the formation of dityrosine and changes in
212 sulfhydryl groups are the most common methods to detect and quantify protein oxidation. In
213 addition, there are some advanced methods such as fluorescence spectroscopy, and electron

214 spin resonance, immune-spin trapping in combination with mass spectrometry used to
215 investigate the mechanism of protein oxidation (Dalsgaard et al., 2014)

216

217 **2.2.1 Function of Proteins as Antioxidants**

218 On the other hand, in some cases, proteins besides being the target for oxidation, are also
219 known to be able to function as antioxidants. The mechanisms of proteins and amino acids as
220 antioxidants in food have been related to their ability to chelate pro-oxidative metals;
221 sulfhydryl groups, which exist in the proteins and amino acids can inactivate free radicals
222 (Viljanen, Kylli, Hubbermann, Schwarz, & Heinonen, 2005). In some cases, the proteins may
223 act as a shield between ROS and the lipid (Dalsgaard et al., 2011). Carnosine can prevent lipid
224 oxidation in various model systems by free radical scavenging or metal ion chelating (Liu,
225 Xing, Fu, Zhou, & Zhang, 2016). His residues and carnosine can postpone oxidation by
226 removing metal ions from the surface of other macromolecules (Guiotto, Calderan, Ruzza, &
227 Borin, 2005). Furthermore, some peptides, which contain Trp or Tyr at the C-terminus have
228 shown to possess a high radical scavenging ability (Saito et al., 2003) and as mentioned above
229 e.g. dityrosine formation as a consequence of two tyrosine radicals reaction with each other
230 will terminate the radical (Dalsgaard et al., 2011). The amount of riboflavin, which is
231 responsible for photo-oxidation is variable in different fish species (Brjekkan, 1959), may play
232 an important role as well. It seems that fish with higher amount of proteins contain more
233 riboflavin. Therefore, Trp and Tyr can be considered as main target of photo-oxidation in fish
234 fillet, which are the only amino acids that can compete with oxygen or unsaturated fatty acids
235 in the quenching of triplet state riboflavin (Cardoso, Franco, Olsen, Andersen, & Skibsted,
236 2004; Dalsgaard et al., 2011). However, the sequence and categories of amino acid might have
237 a key role in the peptides antioxidant activity (Liu et al., 2016). Imidazole ring in the R group
238 of His has the ability of metal ion-chelating, lipid peroxy radical trapping and hydrogen

239 donating (Chan, & Decker, 1994). In addition, Pro-His-His sequence showed higher
240 antioxidant ability in the linoleic acid system compare to other synthetic peptides (Liu et al.,
241 2016), while, the mechanism of hydrophobic amino acids such as Trp and Try might be differ.
242 The presence of peptides are enhanced at the water-lipid interface which can approach to the
243 lipid phase and scavenge free radicals (Ranathunga, Rajapakse, & Kim, 2006).
244 Hence lipid oxidation in fish could also be retarded due to the simultaneous presence of certain
245 proteins and amino acids. However, this connection is also still widely unexplored.

246

247 **3. Impact of lipid and protein oxidation on fish fillet traits**

248 **3.1 Lipid oxidation and quality parameters**

249 Sensory, nutritional value and colour are the major quality characteristics, which can be
250 affected in muscle foods by lipid oxidation. Several authors reported the negative effects of
251 lipid oxidation on the sensory aspect of fish fillet (Baron et al., 2007; Yin, Luo, Fan, Wu, &
252 Feng, 2014). Yin et al. (2014) studied effects of frozen storage on grass carp
253 (*Ctenopharyngodon idellus*) fillet and reported that the sensory parameters (colour, odour,
254 morphology and muscle elasticity) decreased significantly d by the effect of lipid oxidation.
255 Estévez, Ventanas, & Cava, (2005) reported a significant correlation between lipid oxidation
256 and fat content in liver of pâté. Fat content might have a key role in the development of lipid
257 and protein oxidation in the muscle foods (Estevez, Morcuende, & Cava, 2003; Stadtman,
258 1990). In order to find a correlation between fat content and the development of lipid oxidation
259 we compared PV results of two different kind of fish species (lean and fatty fish). Comparing
260 the studies by Saeed & Howell (2002) and Baron et al. (2007) on Atlantic mackerel (*Scomber*
261 *scombrus*) rainbow trout (*Oncorhynchus mykiss*), respectively, reveals that in fatty fish lipid
262 oxidation started earlier compared to lean fish and PV values were higher during storage at -
263 20 °C in fatty fish.

264 In most fish, the major contributor to the colour of muscle is myoglobin. During storage,
265 ferrous oxymyoglobin (Fe^{2+}) oxidized to ferric metmyoglobin (Fe^{3+}), which is responsible for
266 a discoloration reaction in fish fillets (Papuc et al., 2017). Secondary lipid oxidation products
267 (aldehydes) can, furthermore, alter the myoglobin structure via covalent bonds and change the
268 fillet colour during the storage time (Lynch, & Faustman, 2000). In addition, brown pigments
269 in fish can be generated via lipid-protein interaction. In this case, lipid peroxide can interact
270 with active types of proteins and leads to the transformation of the light coloured or colourless
271 precursor to brown pigments (Hidalgo, & Zamora, 2000).

272

273 **3.2 Consequences of protein oxidation**

274 The hydrophobicity, solubility, WHC tenderness and texture are the most important quality
275 parameters, which depend directly on protein oxidation in fish fillet (Lund et al., 2011).
276 Oxidation in a side chains of amino acids can produce carbonyl groups that eventually result
277 in a loss of solubility and protein aggregation (Rowe, Maddock, Lonergan, & Huff-Lonergan,
278 2004).

279

280 3.2.1 Hydrophobicity

281 Hydrophobic residues, in the native form of proteins, are hidden. In a semi oxidised protein,
282 the alteration of secondary and tertiary of protein structure expose the hydrophobic residues
283 to the proteases enzyme substrates follows by protein degradation (Jung, Hohn, & Grune,
284 2014), while heavily oxidation resulted in protein aggregation via cross-linked proteins (Hohn
285 et al., 2011). Aggregated proteins are stable against enzymatic degradation (Reeg, & Grune,
286 2015).

287 3.2.2 Water Holding Capacity (WHC)

288 Protein oxidation has a negative affect on the WHC. As the accessibility of the polar groups
289 to pro-oxidants, which are present in fish muscle, is very high, they are more prone to oxidative
290 reactions (Standal et al., 2018). Protein carbonylation leads to the loss of amino groups, which
291 in turn results in the alteration of the distribution of the electrical charges and the overall
292 arrangement of myofibril protein. It appears that one result of intensive oxidative protein
293 modification is a change in the isoelectric points of proteins. Therefore, the oppositely charged
294 groups are more attracted to each other and thereby decrease the amount of water that is held
295 by the protein. Moreover, the repulsion of the myofibril protein structures is reduced due to
296 the isoelectric point therefore, the protein structures can be more compact and decrease WHC
297 (Huff-Lonergan, & Lonergan, 2005).

298

299 3.2.3 Tenderness and Firmness

300 The tenderness of fish fillets can change as a result of protein oxidation. There are two
301 hypotheses for a decrease in tenderness in relation to protein oxidation:

- 302 a) Amplification of the myofibrillar structure through the formation of MP cross-linking
303 (Kim, Huff-Lonergan, Sebranek, & Lonergan, 2010) particularly, in the presence of
304 oxygen. Myosin proteins especially, MHC is susceptible to oxidation due to the
305 presence of Cys in the root tail of myosin, which is responsible for protein cross-link.
306 The results of this cross-link may decrease the fish muscle tenderness.
307 Glutathionylation of particular Cys in myosin binding protein C and troponin I might
308 be responsible to change the sensitivity of myofilament to calcium (Patel, Wilder, &
309 Solaro, 2013). In several studies, myosin has been reported as a target of
310 glutathionylation and also protein oxidation (Passarelli et al., 2010) but still there is a
311 lack of knowledge about the functional and structural alteration in myosin by the effect
312 of specific redox sites.

313 b) μ -Calpain inactivation (Rowe et al., 2004). For peptide bonds hydrolysis by calpains,
314 electron transfer is needed on the active side chain of His and Cys residues. In this
315 case, carbonyl derivatives of some amino acids, such as His, lead to the formation of
316 inter and/or intra-protein disulfide cross-links (Martinaud et al., 1997). Because SH-
317 containing Cys and His residues are present at the active sites of both μ -calpain and
318 m-calpain enzymes, these products might be inactivated by oxidation and lead to
319 reduce fish muscle tenderness (Carlin, Huff-Lonergan, Rowe, & Lonergan, 2006).

320 On the other hand, it has been proved that due to protein oxidation firmness is reduced during
321 frozen storage by the impact of ice crystal formation and cell disruption (Hematyar, Masilko,
322 Mraz, & Sampels, 2018; Subbaiah, 2015).

323 It seems that myoglobin and myofibril are the most important proteins to impact on fish
324 quality. Probably, the formation of covalent bonds and cross-links, by the impact of lipids and
325 proteins oxidation are responsible for quality changes in the fish fillet. Therefore, extensive
326 oxidation in fish muscle often leads to higher firmness while moderate protein oxidation can
327 decline the firmness due to more unfold protein structures. Additionally, degradation and
328 aggregation of proteins in the post mortem fish muscle lead to higher firmness and lower
329 tenderness. On the other hand, during frozen storage formation of unfolded proteins due to
330 oxidation or lipid-protein interaction probably are responsible for reduction of firmness.

331

332 **4. Correlation and interaction of lipid and protein oxidation**

333 During the storage of fish fillet, proteins are exposed to oxidised lipids or secondary
334 breakdown products that may cause some changes in protein functionality including
335 insolubilisation, polymerisation, loss of enzymatic activity and formation of lipid-protein
336 complexes (Howell, Herman, & Li-Chan, 2001).

337 **4.1 Mechanisms of interaction**

338 Two mechanisms can be considered for the interaction of lipids-proteins:

339 a) First, hydroperoxide decomposes to secondary lipid oxidation products e.g. malonaldehyde
340 that may react further with amino groups (Schaich, 1976) or for unsaturated lipid aldehydes
341 reacting with Cys, His, or Lys through Michael addition (Stadtman, & Levine, 2000).

342 b) Some products of lipid oxidation (hydroperoxides, lipid free radicals, and volatile secondary
343 oxidation products) can react with proteins to generate protein-centred free radicals (Saeed,
344 Fawthrop, & Howell, 1999).

345 Secondary lipid oxidation products can bind to proteins in two main ways by either binding to
346 an active site or cavity or at less well defined hydrophobic patches at positions close to the
347 surface of the protein (Fillery-Travis, Mills, & Wilde, 2000). Aldehydes as a product of metal
348 catalysed lipid oxidation cause, protein-lipid aggregation via crosslink bonds (Gardner, 1979).

349 The pathways for the interaction of lipid oxidation products (carbonyls) with proteins are
350 formation of Schiff base and Michael-type (Refsgaard, Tsai, & Stadtman, 2000). In the
351 carbonyl amine reaction, aldehydes can bind directly to amino groups in proteins through
352 covalent bonds. Interaction of dimethylamine and formaldehyde from trimethylamine-N-
353 oxide may result in decreased WHC as formaldehyde may form crosslinks between proteins,
354 which have been shown to decrease WHC and protein solubility in the Gadiform Fish fillet
355 (Nielsen, & Jorgensen, 2004).

356 Also, the products of lipid oxidation might have interaction with proteins on the hydrophobic
357 groups site, which lead to generate lipid-soluble fluorescence products. The consequence of
358 this interaction in the defatted samples can be resulted to decline in soluble protein
359 hydrophobicity (Liang, 1999) upon when the secondary and tertiary structure of proteins are
360 changed (Meng, Chan, Rousseau, & Li-Chan, 2005). This reaction mostly happens in the
361 presence of some prone amino acids such as His, Lys, Pro, Arg, Tyr, Trp, Cys, and Met via
362 side chain reaction with lipid oxidation products. As soon as protein unfolding occurs at the

363 water-lipid interface, the hydrophobic groups are absorbed and interact with the lipid phase.
364 While, the negatively charged groups of protein can remain in contact with water molecules
365 and hence increase the solubility or reduce the risk of protein-protein aggregation (Gitlin,
366 Carbeck, & Whitesides, 2006; Kramer, Shende, Motl, Pace, & Scholtz, 2012).
367 Thus, it appears that the covalent bonds are a very important type of bonds between free amino
368 groups of Cys, His, and Lys with MDA via side chain pathway (Pizzimenti et al., 2013). For
369 a better understanding of protein-lipid interaction, it is necessary to know more about the
370 water-lipid bonds and also the lipid-protein relationship, as it seems that there is a close
371 relationship between them (Alzagat, & Alli, 2002). In the frozen fish muscle due to the
372 absence of water molecules, lipid-protein or protein-protein interactions are dominated.
373 Additionally, the interactions depend on the secondary products or radicals predominate that
374 can be specified by which proteins get involved in the lipid oxidation reaction chain (Ladikos,
375 & Lougovois, 1990). Furthermore, the stability of the proteins and volatiles may increase via
376 non-covalent bonds during the storage time of muscle foods.

377

378 **4.2 Role of haemoglobin and myoglobin on the progress of lipid oxidation**

379 It has also been demonstrated in model systems that lipid oxidation can increase in the
380 presence of protein radicals (Østdal, Davies, & Andersen, 2002). A good example for this kind
381 of interaction in fish muscle has been reported by Richard & Hultin (2002) that revealed
382 oxidised haemoglobin (deoxyhemoglobin) is a powerful catalyst of lipid oxidation.

383 Haemoglobin (Hb) autoxidation rate in different fish species, might be influenced by their
384 residue (Powers, 1972). Richards & Hultin (2003) revealed that Hb from mackerel (*Scomber*
385 *scombrus*) and herring (*Clupea harengus*) showed more pro-oxidative activity than from trout
386 (*Onchorhynchus mykiss*) Hb. This would be related to the frequent migration, which made Hb
387 more susceptible to autoxidation. Probably, mackerel blood contains more lipid oxidation

388 promoters or less powerful inhibitors (Richards, & Hultin, 2003). However, Maqsood &
389 Benjakul (2011); Maqsood et al. (2011) reported that active migratory fish like seabass (*Lates*
390 *calcarifer*) had less pro-oxidative Hb. In addition, Hb autoxidation in the fish from cold water
391 was 10-fold faster than warm water fish (Maqsood, & Benjakul, 2011). Might be His residue
392 is situated away from the centre and makes Hb more resistance to oxidise (Jensen, 2001).
393 Probably, when His and Phe residue are located on the distal part of the haem group the
394 accessibility of iron to the mentioned amino acids is decreased that leads to less interaction
395 between Hbs and lipid oxidation products. The rate of lipid oxidation can be affected by the
396 different Hbs formations in fish flesh (Maqsood, Benjakul, & Kamal-Eldin, 2012). Richards
397 & Hultin (2002) reported that deoxyHb is a stronger catalyst for lipid oxidation compare to
398 oxyHb. Formation of deoxyHb release the iron from the inside of the porphyrin group, which
399 is a catalyst for lipid oxidation and lipid-protein interaction. Immediate bleeding would keep
400 the freshness of muscle for a longer time and maintenance the fillet firmness during the storage
401 time. It has been demonstrated that inadequately bled or non-bled fish show the lower overall
402 quality in the fish fillet as bleeding decrease the total haemoglobin in the muscle (Richards, &
403 Hultin, 2002).

404 In fish, myoglobin oxidation and lipid oxidation are associated and influence each other
405 (Chaijan, 2008). Secondary lipid oxidation products (aldehydes) can modify the stability of
406 myoglobin and generate adducts through a covalent modification with myoglobin but in the
407 absence of lipid oxidation products metal catalyse via side chain reaction can unfold the
408 proteins. In line with this, Lynch & Faustman (2000) proposed that pro-oxidant activity of
409 metmyoglobin and oxymyoglobin oxidation will be increased by aldehydes. Furthermore,
410 metmyoglobin and H₂O₂, resulting from oxymyoglobin oxidation, can provoke lipid oxidation.
411 In one way the products of oxymyoglobin (metmyoglobin and H₂O₂) are necessary to start

412 lipid oxidation and on the other way aldehydes can change myoglobin stability and promote
413 oxymyoglobin oxidation

414

415 **4.3 Myosin- lipid interaction**

416 Myosin is another protein in fish that has a weak tendency to form a complex with lipids, but
417 the form of myosin can change during the storage due to unfolding. However, linoleic acid
418 hydroperoxides are highly destructive for myofibrillar structure that can precipitate and
419 denature the A-band (predominantly myosin). Chopin, Kone & Serot (2007) investigated the
420 interaction between fish myosin and secondary of lipid oxidation products and found a
421 correlation between myosin solution and aldehydes, which leads to lead to a decline of protein
422 solubility, due to aggregations, also supported by (Buttkus, 1966). It seems that the interaction
423 is taking place at some uncovered and new opened sites of unfolded myosin chain. During this
424 interaction myosin solubility, sulfhydryl groups and free amino groups significantly
425 decreased, thus supporting the idea of Schiff base and/or Michael addition reaction taking
426 place.

427

428 In order to make more visual, we showed some mechanisms and target proteins in the
429 relationship between lipid and protein oxidation products in a schematically graph (Figure 1
430 and 2).

431 During post mortem of fish muscle the lack of ATP and anaerobic conditions leading to the
432 antioxidants consumption which are resulted to the oxidation development. Probably during
433 post-mortem changes, first metal catalysed oxidation in proteins are dominated follow with
434 lipid oxidation development. We would say maybe protein oxidation is started earlier than
435 lipid oxidation or both are in parallel. Therefore, bleeding or washing the muscle after stunning
436 can be considered as a main factor to reduce this interaction. On the other hand, it appears that

437 during frozen storage formation of secondary products of lipid oxidation have key roll on
438 lipid-protein interaction via Schiff base reaction on the side chain of amino acids.

439 Secondary lipid oxidation products might have more impact on the functionality of proteins
440 from both Michael addition and Schiff base reaction rather than lipid hydroperoxides.

441

442 **6. Conclusions**

443 The present review summarizes the main actions of protein oxidation and the possible
444 connection to lipid oxidation with a focus on fish quality. Until now, both oxidation processes
445 have been investigated more or less separate from each other. However, both processes occur
446 in parallel in fish and fish products. The reaction products can then react further with each
447 other and form either volatile or non-volatile stable products. Formation of 'induced' protein-
448 lipid complexes is the result of this interaction.

449 We propose that the products of lipid oxidation promote protein oxidation in fish rather than
450 the other way around specially, during frozen storage. While during post-mortem changes,
451 protein oxidation is dominating. This hypothesis needs to be explored in model systems as
452 well as on real fish samples. In addition, a comparison of lean and fatty fish should be made.

453 There is the possibility, that retarding lipid oxidation could also slow down protein oxidation
454 and hence increase shelf life of fish. Furthermore, there is a strong need to investigate how
455 oxidation products influence the kinetics of the ongoing lipid and protein oxidation processes
456 to better prevent oxidation and food spoilage. Finally, it seems that, secondary products of
457 lipid oxidation might have more impact on the functionality of proteins from both Michael
458 addition and Schiff base reaction rather than lipid hydroperoxides.

459

460 **7. Acknowledgements**

461 The study was financially supported by the Ministry of Education, Youth and Sports of the Czech
462 Republic - projects „CENAKVA“ (No. CZ.1.05/2.1.00/01.0024), “CENAKVA II“ (No. LO1205 under
463 the NPU I program) and GAJU 060/2016/Z.

464

465

466 References

- 467 Aeschbach, R., Amadoò, R., & Neukom, H. (1976). Formation of dityrosine cross-links in proteins by
468 oxidation of tyrosine residues. *Biochimica et Biophysica Acta (BBA) - Protein Structure*,
469 439(2), 292-301. doi: [https://doi.org/10.1016/0005-2795\(76\)90064-7](https://doi.org/10.1016/0005-2795(76)90064-7)
- 470 Alzagat, A. A., & Alli, I. (2002). Protein-lipid interactions in food systems: a review. *International*
471 *Journal of Food Sciences and Nutrition*, 53(3), 249-260. doi: 10.1080/09637480220132850
- 472 Adolfo Amici, R. L. L., Lin Tsai, and Earl R. Stadtman. (1989). Conversion of Amino Acid Residues in
473 Proteins and Amino Acid Homopolymers to Carbonyl Derivatives by Metal-catalyzed Oxidation
474 Reactions. *Journal of Biological Chemistry*. 264(6), 3341-3346.
- 475 Baron, C. P., Kjaersgard, I. V. H., Jessen, F., & Jacobsen, C. (2007). Protein and lipid oxidation during
476 frozen storage of rainbow trout (*Oncorhynchus mykiss*). *Journal of Agricultural and Food*
477 *Chemistry*, 55, 8118-8125.
- 478 Brjekkan, O. R. (1959). A Comparative Study of Vitamins in the Trunk Muscles of Fishes *Director of*
479 *Fisheries*, 3(8).
- 480 Buttkus, H. (1966). Preparation and Properties of Trout Myosin. *Journal of the Fisheries Research Board*
481 *of Canada*, 23(4), 563-573.
- 482 Cai, J., Bhatnagar, A., & Pierce, W. M. (2009). Protein Modification by Acrolein: Formation and Stability
483 of Cysteine Adducts. *Chemical Research in Toxicology*, 22(4), 708-716. doi:
484 10.1021/tx800465m
- 485 Cardoso, D. R., Franco, D. W., Olsen, K., Andersen, M. L., & Skibsted, L. H. (2004). Reactivity of bovine
486 whey proteins, peptides, and amino acids toward triplet riboflavin as studied by laser flash
487 photolysis. *Journal of Agricultural and Food Chemistry*, 52(21), 6602-6606. doi:
488 10.1021/jf0401165
- 489 Carlin, K. R. M., Huff-Lonergan, E., Rowe, L. J., & Lonergan, S. M. (2006). Effect of oxidation, pH, and
490 ionic strength on calpastatin inhibition of μ - and m-calpain. *Journal Of Animal Science*, 84(4),
491 925-937. doi: 10.2527/2006.844925x
- 492 Chaijan, M. (2008). Lipid and myoglobin oxidations in muscle foods. *Songklanakarin Journal of Science*
493 *and Technology (SJST)*, 30, 47-53.
- 494 Chan, K. M., & Decker, E. A. (1994). Endogenous Skeletal-Muscle Antioxidants. *Critical Reviews in Food*
495 *Science and Nutrition*, 34(4), 403-426. doi: 10.1080/10408399409527669
- 496 Choe, E., & Min, D. B. (2006). Mechanisms of Antioxidants in the Oxidation of Foods. *Comprehensive*
497 *Reviews in Food Science and Food Safety*, 8(4), 345-358. doi: 10.1111/j.1541-
498 4337.2009.00085.x
- 499 Chopin, C., Kone, M., & Serot, T. (2007). Study of the interaction of fish myosin with the products of
500 lipid oxidation: The case of aldehydes. *Food Chemistry*, 105(1), 126-132. doi:
501 10.1016/j.foodchem.2007.03.058
- 502 Dalsgaard, T. K., Nielsen, J. H., Brown, B. E., Stadler, N., & Davies, M. J. (2011). Dityrosine, 3,4-
503 Dihydroxyphenylalanine (DOPA), and Radical Formation from Tyrosine Residues on Milk
504 Proteins with Globular and Flexible Structures as a Result of Riboflavin-Mediated Photo-

- 505 oxidation. *Journal Of Agricultural And Food Chemistry*, 59(14), 7939-7947. doi:
506 10.1021/jf200277r
- 507 Dalsgaard, T. K., Nielsen, J. H., & Larsen, L. B. (2007). Proteolysis of milk proteins lactosylated in model
508 systems. *Molecular Nutrition & Food Research*, 51(4), 404-414. doi: 10.1002/mnfr.200600112
- 509 Dalsgaard, T. K., Otzen, D., Nielsen, J. H., & Larsen, L. B. (2007). Changes in structures of milk proteins
510 upon photo-oxidation. *Journal Of Agricultural And Food Chemistry*, 55(26), 10968-10976. doi:
511 10.1021/jf071948g
- 512 Dalsgaard, T. K., Sorensen, J., Bakman, M., Nebel, C., Albrechtsen, R., Vognsen, L., & Nielsen, J. H.
513 (2011). Light-induced protein and lipid oxidation in low-fat cheeses: whey proteins as
514 antioxidants. *Dairy Science & Technology*, 91(2), 171-183. doi: 10.1007/s13594-011-0001-1
- 515 Dalsgaard, T. K., Sorensen, J., Bakman, M., Vognsen, L., Nebel, C., Albrechtsen, R., & Nielsen, J. H.
516 (2010). Light-induced protein and lipid oxidation in cheese: Dependence on fat content and
517 packaging conditions. *Dairy Science & Technology*, 90(5), 565-577. doi: 10.1051/dst/2010019
- 518 Dalsgaard, T. K., Triquigneaux, M., Deterding, L., Summers, F. A., Mortensen, G., & Mason, R. P. (2014).
519 Oxidation of α -lactalbumin after a lactoperoxidase-catalysed reaction: An oxidomics approach
520 applying immuno-spin trapping and mass spectrometry. *International Dairy Journal*, 38(2),
521 154-159. doi: <https://doi.org/10.1016/j.idairyj.2013.11.005>
- 522 Davies, K. J. A., & Delsignore, M. E. (1987). Protein Damage and Degradation by Oxygen Radicals .3.
523 Modification Of Secondary And Tertiary Structure. *Journal of Biological Chemistry*, 262(20),
524 9908-9913.
- 525 Davies, K. J. A., Delsignore, M. E., & Lin, S. W. (1987). Protein Damage and Degradation by Oxygen
526 Radicals .2. Modification Of Amino-Acids. *Journal of Biological Chemistry*, 262(20), 9902-9907.
- 527 Di Bernardini, R., Harnedy, P., Bolton, D., Kerry, J., O'Neill, E., Maria Mullen, A., & Hayes, M. (2011).
528 *Antioxidant and antimicrobial peptidic hydrolysates from muscle protein sources and by-*
529 *products* (Vol. 124).
- 530 Estévez, M. (2015). Oxidative damage to poultry: from farm to fork. *Poultry Science*, 94(6), 1368-1378.
531 doi: 10.3382/ps/pev094
- 532 Estévez, M., & Cava, R. (2004). Lipid and protein oxidation, release of iron from heme molecule and
533 colour deterioration during refrigerated storage of liver pâté. *Meat Science*, 68(4), 551-558.
534 doi: <http://dx.doi.org/10.1016/j.meatsci.2004.05.007>
- 535 Estevez, M., Morcuende, D., Ventanas, S., & Cava, R. (2003). Analysis of volatiles in meat from Iberian
536 pigs and lean pigs after refrigeration and cooking by using SPME-GC-MS. *Journal Of Agricultural*
537 *And Food Chemistry*, 51(11), 3429-3435.
- 538 Estévez, M., Ventanas, S., & Cava, R. (2005). Physicochemical properties and oxidative stability of liver
539 pâté as affected by fat content. *Food Chemistry*, 92(3), 449-457. doi:
540 <http://dx.doi.org/10.1016/j.foodchem.2004.08.014>
- 541 Fedorova, M., Bollineni, R. C., & Hoffmann, R. (2014). Protein Carbonylation as a Major Hallmark of
542 Oxidative Damage: Update of Analytical Strategies. *Mass Spectrometry Reviews*, 33(2), 79-97.
543 doi: 10.1002/mas.21381
- 544 Fillery-Travis, A., Mills, E. N. C., & Wilde, P. (2000). Protein-lipid interactions at interfaces. *Grasas Y*
545 *Aceites*, 51(1-2), 50-55.
- 546 Foote, C. S. (1976). Photosensitized oxidation and singlet oxygen: consequences in biological systems.
547 *Free radicals in biology*, 2, 85-133.
- 548 Gardner, H. W. (1979). Lipid hydroperoxide reactivity with proteins and amino acids: a review. *Journal*
549 *Of Agricultural And Food Chemistry*, 27(2), 220-229. doi: 10.1021/jf60222a034
- 550 Gitlin, I., Carbeck, J. D., & Whitesides, G. M. (2006). Why Are Proteins Charged? Networks of Charge-
551 Charge Interactions in Proteins Measured by Charge Ladders and Capillary Electrophoresis.
552 *Angewandte Chemie International Edition*, 45(19), 3022-3060. doi: 10.1002/anie.200502530
- 553 Guiotto, A., Calderan, A., Ruzza, P., & Borin, G. (2005). Carnosine and carnosine-related antioxidants:
554 A review. *Current Medicinal Chemistry*, 12(20), 2293-2315. doi: 10.2174/0929867054864796

- 555 Hawkins, C. L., & Davies, M. J. (2001). Generation and propagation of radical reactions on proteins.
556 *Biochimica Et Biophysica Acta-Bioenergetics*, 1504(2-3), 196-219. doi: 10.1016/s0005-
557 2728(00)00252-8
- 558 Heinonen, I. M., Meyer, A. S., & Frankel, E. N. (1998). Antioxidant Activity of Berry Phenolics on Human
559 Low-Density Lipoprotein and Liposome Oxidation. *J. Agric. Food Chem.*, 46(10), 4107-4112.
- 560 Hematyar, N., Masilko, J., Mraz, J., & Sampels, S. (2018). Nutritional quality, oxidation, and sensory
561 parameters in fillets of common carp (*Cyprinus carpio* L.) influenced by frozen storage (-20 °C).
562 *Journal of Food Processing and Preservation*. doi: 10.1111/jfpp.13589
- 563 Hidalgo, F. J., & Zamora, R. (2000). The role of lipids in nonenzymatic browning. [Amino-carbonyl
564 reactions; Maillard reaction; Nonenzymatic browning; Oxidized lipid-protein interactions].
565 2000, 51(1-2), 15. doi: 10.3989/gya.2000.v51.i1-2.405
- 566 Hohn, A., Jung, T., Grimm, S., Catalgol, B., Weber, D., & Grune, T. (2011). Lipofuscin inhibits the
567 proteasome by binding to surface motifs. *Free Radical Biology and Medicine*, 50(5), 585-591.
568 doi: 10.1016/j.freeradbiomed.2010.12.011
- 569 Howell, N. K., Herman, H., & Li-Chan, E. C. Y. (2001). Elucidation of protein-lipid interactions in a
570 lysozyme-corn oil system by Fourier transform Raman spectroscopy. *Journal of Agricultural
571 and Food Chemistry*, 49(3), 1529-1533. doi: 10.1021/jf001115p
- 572 Huff-Lonergan, E., & Lonergan, S. M. (2005). Mechanisms of water-holding capacity of meat: The role
573 of postmortem biochemical and structural changes. *Meat Science*, 71(1), 194-204. doi:
574 10.1016/j.meatsci.2005.04.022
- 575 Jensen, F. B. (2001). Comparative analysis of autoxidation of haemoglobin. *Journal of Experimental
576 Biology*, 204(11), 2029-2033.
- 577 Jeremiah, L. E. (2001). Packaging alternatives to deliver fresh meats using short- or long-term
578 distribution. *Food Research International*, 34(9), 749-772.
- 579 Jonsson, A., Olsen, R. E., Hyldig, G., Nielsen, H. H., Jorgensen, B., Larsson, K., Undeland, I., &
580 Hafssteinsson, H. (2007). Improved quality of herring for humans *Nordic Innovation Centre
581 (NiCe) project number: 02106 78*.
- 582 Jung, T., Hohn, A., & Grune, T. (2014). The proteasome and the degradation of oxidized proteins: Part
583 II - protein oxidation and proteasomal degradation. *Redox Biology*, 2, 99-104. doi:
584 10.1016/j.redox.2013.12.008
- 585 Kim, Y. H., Huff-Lonergan, E., Sebranek, J. G., & Lonergan, S. M. (2010). High-oxygen modified
586 atmosphere packaging system induces lipid and myoglobin oxidation and protein
587 polymerization. *Meat Science*, 85(4), 759-767. doi: 10.1016/j.meatsci.2010.04.001
- 588 Kjaersgard, I. V. H., & Jessen, F. (2004). Two-dimensional gel electrophoresis detection of protein
589 oxidation in fresh and tainted rainbow trout muscle. [Article]. *Journal of Agricultural and Food
590 Chemistry*, 52(23), 7101-7107.
- 591 Kramer, R. M., Shende, V. R., Motl, N., Pace, C. N., & Scholtz, J. M. (2012). Toward a molecular
592 understanding of protein solubility: increased negative surface charge correlates with
593 increased solubility. *Biophysical journal*, 102(8), 1907-1915. doi: 10.1016/j.bpj.2012.01.060
- 594 Kubow, S. (1992). Routes of Formation and Toxic Consequences of Lipid Oxidation-Products In Foods.
595 *Free Radical Biology and Medicine*, 12(1), 63-81. doi: 10.1016/0891-5849(92)90059-p
- 596 Ladikos, D., & Lougovois, V. (1990). Lipid Oxidation in Muscle Foods - a Review. *Food Chemistry*, 35(4),
597 295-314.
- 598 Liang, J. H. (1999). Fluorescence due to interactions of oxidizing soybean oil and soy proteins. *Food
599 Chemistry*, 66(1), 103-108. doi: 10.1016/s0308-8146(98)00250-7
- 600 Liu, R., Xing, L. J., Fu, Q. Q., Zhou, G. H., & Zhang, W. G. (2016). A Review of Antioxidant Peptides
601 Derived from Meat Muscle and By-Products. *Antioxidants*, 5(3). doi: Unsp
602 3210.3390/antiox5030032
- 603 Lund, M. N., Heinonen, M., Baron, C. P., & Estevez, M. (2011). Protein oxidation in muscle foods: A
604 review. *Molecular Nutrition & Food Research*, 55(1), 83-95. doi: 10.1002/mnfr.201000453

- 605 Lynch, M. P., & Faustman, C. (2000). Effect of aldehyde lipid oxidation products on myoglobin. *Journal*
606 *Of Agricultural And Food Chemistry*, 48(3), 600-604. doi: 10.1021/jf990732e
- 607 Maqsood, S., & Benjakul, S. (2011). Comparative studies on molecular changes and pro-oxidative
608 activity of haemoglobin from different fish species as influenced by pH. *Food Chemistry*,
609 124(3), 875-883. doi: 10.1016/j.foodchem.2010.07.01
- 610 Maqsood, S., Benjakul, S., & Kamal-Eldin, A. (2012). Haemoglobin-mediated lipid oxidation in the fish
611 muscle: A review. *Trends in Food Science & Technology*, 28(1), 33-43. doi:
612 <http://dx.doi.org/10.1016/j.tifs.2012.06.009>
- 613 Maqsood, S., Singh, P., Samoon, M. H., & Munir, K. (2011). Emerging role of immunostimulants in
614 combating the disease outbreak in aquaculture. *International Aquatic Research*, 3(3), 147-163.
- 615 Martinaud, A., Mercier, Y., Marinova, P., Tassy, C., Gatellier, P., & Renerre, M. (1997). Comparison of
616 oxidative processes on myofibrillar proteins from beef during maturation and by different
617 model oxidation systems. *Journal of Agricultural and Food Chemistry*, 45(7), 2481-2487.
- 618 Medina, I., Gallardo, J. M., & Aubourg, S. P. (2009). Quality preservation in chilled and frozen fish
619 products by employment of slurry ice and natural antioxidants. [Review]. *International Journal*
620 *of Food Science and Technology*, 44(8), 1467-1479. doi: 10.1111/j.1365-2621.2009.02016.x
- 621 Meng, G. T., Chan, J. C. K., Rousseau, D., & Li-Chan, E. C. Y. (2005). Study of protein - Lipid interactions
622 at the bovine serum albumin/oil interface by Raman microspectroscopy. *Journal Of*
623 *Agricultural And Food Chemistry*, 53(4), 845-852. doi: 10.1021/jf040259r
- 624 Metz, B., Kersten, G. F. A., Hoogerhout, P., Brugghe, H. F., Timmermans, H. A. M., de Jong, A., . . .
625 Jiskoot, W. (2004). Identification of formaldehyde-induced modifications in proteins -
626 Reactions with model peptides. *Journal of Biological Chemistry*, 279(8), 6235-6243. doi:
627 10.1074/jbc.M310752200
- 628 Moller, I. M., Rogowska-Wrzesinska, A., & Rao, R. S. P. (2011). Protein carbonylation and metal-
629 catalyzed protein oxidation in a cellular perspective. *Journal of Proteomics*, 74(11), 2228-2242.
630 doi: 10.1016/j.jprot.2011.05.004
- 631 Mozuraityte, R., Kristinova, V., Rustad, T., & Storro, I. (2016). The role of iron in peroxidation of PUFA:
632 Effect of pH and chelators. *European Journal of Lipid Science and Technology*, 118(4), 658-668.
633 doi: 10.1002/ejlt.201400590
- 634 Nielsen, M. K., & Jorgensen, B. M. (2004). Quantitative relationship between trimethylamine oxide
635 aldolase activity and formaldehyde accumulation in white muscle from gadiform fish during
636 frozen storage. *Journal Of Agricultural And Food Chemistry*, 52(12), 3814-3822. doi:
637 10.1021/jf035169l
- 638 Nystrom, T. (2005). Role of oxidative carbonylation in protein quality control and senescence. *Embo*
639 *Journal*, 24(7), 1311-1317. doi: 10.1038/sj.emboj.7600599
- 640 Østdal, H., Davies, M. J., & Andersen, H. J. (2002). Reaction between protein radicals and other
641 biomolecules. *Free Radical Biology and Medicine*, 33(2), 201-209. doi:
642 [http://dx.doi.org/10.1016/S0891-5849\(02\)00785-2](http://dx.doi.org/10.1016/S0891-5849(02)00785-2)
- 643 Papuc, C., Goran, G. V., Predescu, C. N., & Nicorescu, V. (2017). Mechanisms of Oxidative Processes in
644 Meat and Toxicity Induced by Postprandial Degradation Products: A Review. *Comprehensive*
645 *Reviews in Food Science and Food Safety*, 16(1), 96-123. doi: 10.1111/1541-4337.12241
- 646 Passarelli, C., Di Venere, A., Piroddi, N., Pastore, A., Scellini, B., Tesi, C., . . . Piemonte, F. (2010).
647 Susceptibility of Isolated Myofibrils to In Vitro Glutathionylation: Potential Relevance to
648 Muscle Functions. *Cytoskeleton*, 67(2), 81-89. doi: 10.1002/cm.20425
- 649 Patel, B. G., Wilder, T., & Solaro, R. J. (2013). Novel control of cardiac myofilament response to calcium
650 by S-glutathionylation at specific sites of myosin binding protein C. *Frontiers in Physiology*, 4.
651 doi: 33610.3389/fphys.2013.00336
- 652 Pizzimenti, S., Ciamporcero, E., Daga, M., Pettazzoni, P., Arcaro, A., Cetrangolo, G., . . . Barrera, G.
653 (2013). Interaction of aldehydes derived from lipid peroxidation and membrane proteins.
654 *Frontiers in Physiology*, 4. doi: 24210.3389/fphys.2013.00242

- 655 Powers, D. A. (1972). Hemoglobin Adaptation for Fast and Slow Water Habitats In Sympatric
656 Catostomid Fishes. *Science*, 177(4046), 360-&. doi: 10.1126/science.177.4046.360
- 657 Ranathunga, S., Rajapakse, N., & Kim, S. K. (2006). Purification and characterization of antioxidative
658 peptide derived from muscle of conger eel (*Conger myriaster*). *European Food Research and*
659 *Technology*, 222(3-4), 310-315. doi: 10.1007/s00217-005-0079-x
- 660 Reeg, S., & Grune, T. (2015). Protein Oxidation in Toxicology. In S. M. Roberts, J. P. Kehrer & L.-O. Klotz
661 (Eds.), *Studies on Experimental Toxicology and Pharmacology* (pp. 81-102). Cham: Springer
662 International Publishing.
- 663 Refsgaard, H. H. F., Tsai, L., & Stadtman, E. R. (2000). Modifications of proteins by polyunsaturated
664 fatty acid peroxidation products. *Proceedings of the National Academy of Sciences of the*
665 *United States of America*, 97(2), 611-616. doi: 10.1073/pnas.97.2.611
- 666 Requena, J. R., Chao, C. C., Levine, R. L., & Stadtman, E. R. (2001). Glutamic and amino adipic
667 semialdehydes are the main carbonyl products of metal-catalyzed oxidation of proteins.
668 *Proceedings of the National Academy of Sciences of the United States of America*, 98(1), 69-
669 74. doi: 10.1073/pnas.011526698
- 670 Richards, M. P., & Hultin, H. O. (2002). Contributions of blood and blood components to lipid oxidation
671 in fish muscle. *Journal of Agricultural and Food Chemistry*, 50(3), 555-564. doi:
672 10.1021/jf010562h
- 673 Richards, M. P., & Hultin, H. O. (2003). Effects of added hemolysate from mackerel, herring and
674 rainbow trout on lipid oxidation of washed cod muscle. *Fisheries Science*, 69(6), 1298-1300.
675 doi: 10.1111/j.0919-9268.2003.00758.x
- 676 Richards, M. P., Ostdal, H., & Andersen, H. J. (2002). Deoxyhemoglobin-mediated lipid oxidation in
677 washed fish muscle. *Journal Of Agricultural And Food Chemistry*, 50(5), 1278-1283. doi:
678 10.1021/jf011093m
- 679 Rowe, L. J., Maddock, K. R., Lonergan, S. M., & Huff-Lonergan, E. (2004). Influence of early postmortem
680 protein oxidation on beef quality. *J. Anim Sci.*, 82(3), 785-793.
- 681 Saeed, S., Fawthrop, S. A., & Howell, N. K. (1999). Electron spin resonance (ESR) study on free radical
682 transfer in fish lipid-protein interaction. *Journal of the Science of Food and Agriculture*, 79(13),
683 1809-1816. doi: 10.1002/(sici)1097-0010(199910)79:13<1809::aid-jsfa440>3.0.co;2-v
- 684 Saeed, S., & Howell, N. K. (2002). Effect of lipid oxidation and frozen storage on muscle proteins of
685 Atlantic mackerel (*Scomber scombrus*). *Journal of the Science of Food and Agriculture*, 82(5),
686 579-586. doi: 10.1002/jsfa.1080
- 687 Saito, K., Jin, D. H., Ogawa, T., Muramoto, K., Hatakeyama, E., Yasuhara, T., & Nokihara, K. (2003).
688 Antioxidative properties of tripeptide libraries prepared by the combinatorial chemistry.
689 *Journal Of Agricultural And Food Chemistry*, 51(12), 3668-3674. doi: 10.1021/jf021191n
- 690 Sampels, S. (2013). *Oxidation and Antioxidants in Fish and Meat from Farm to Fork*.
- 691 Sarma, J., Vidya Sagar Reddy, G., & Srikar, L. N. (2000). Effect of frozen storage on lipids and functional
692 properties of proteins of dressed Indian oil sardine (*Sardinella longiceps*). *Food Research*
693 *International*, 33(10), 815-820. doi: [http://dx.doi.org/10.1016/S0963-9969\(00\)00077-6](http://dx.doi.org/10.1016/S0963-9969(00)00077-6)
- 694 Schaich KM, K. M. (1976). Free radical reactions of peroxidizing lipids with amino acids and proteins:
695 an ESR study. *Lipids*, 11(5), 392-400.
- 696 Secci, G., & Parisi, G. (2016). From farm to fork: lipid oxidation in fish products. A review. *Italian Journal*
697 *of Animal Science*, 15(1), 124-136. doi: 10.1080/1828051x.2015.1128687
- 698 Soladoye, O. P., Juarez, M. L., Aalhus, J. L., Shand, P., & Estevez, M. (2015). Protein Oxidation in
699 Processed Meat: Mechanisms and Potential Implications on Human Health. *Comprehensive*
700 *Reviews in Food Science and Food Safety*, 14(2), 106-122. doi: 10.1111/1541-4337.12127
- 701 Soyer, A., Hultin, H. O. (2000). Kinetics of oxidation of the lipids and proteins of cod sarcoplasmic
702 reticulum. *Journal of Agriculture and Food Chemistry*. 48 (6), pp 2127–2134
- 703 Soyer, A., Özalp, B., Dalmış, Ü., & Bilgin, V. (2010). Effects of freezing temperature and duration of
704 frozen storage on lipid and protein oxidation in chicken meat. *Food Chemistry*, 120(4), 1025-
705 1030. doi: <http://dx.doi.org/10.1016/j.foodchem.2009.11.042>

- 706 Stadtman, E. R. (1990). Metal Ion-Catalyzed Oxidation of Proteins - Biochemical-Mechanism and
707 Biological Consequences. *Free Radical Biology and Medicine*, 9(4), 315-325. doi: 10.1016/0891-
708 5849(90)90006-5
- 709 Stadtman, E. R. (1993). Oxidation of Free Amino-Acids and Amino-Acid-Residues In Proteins by
710 Radiolysis and by Metal-Catalyzed Reactions. *Annual Review of Biochemistry*, 62, 797-821. doi:
711 10.1146/annurev.bi.62.070193.004053
- 712 Stadtman, E. R. (2006). Protein oxidation and aging. *Free Radical Research*, 40(12), 1250-1258. doi:
713 10.1080/10715760600918142
- 714 Stadtman, E. R., & Levine, R. L. (2000). Protein oxidation. *Ann N Y Acad Sci*, 899, 191-208.
- 715 Stadtman, E. R., & Levine, R. L. (2003). Free radical-mediated oxidation of free amino acids and amino
716 acid residues in proteins. *Amino Acids*, 25(3-4), 207-218. doi: 10.1007/s00726-003-0011-2
- 717 Standal, I. B., Mozuraityte, R., Rustad, T., Alinasabhematabadi, L., Carlsson, N. G., & Undeland, I. (2018).
718 Quality of Filleted Atlantic Mackerel (*Scomber Scombrus*) During Chilled and Frozen Storage:
719 Changes in Lipids, Vitamin D, Proteins, and Small Metabolites, including Biogenic Amines.
720 *Journal of Aquatic Food Product Technology*, 27(3), 338-357. doi:
721 10.1080/10498850.2018.1436107
- 722 Subbaiah, K., Majumdar, R. K., Choudhury, J., Priyadarshini, B. M., Dhar, B., Roy, D., Saha, A., & Maurya,
723 P. (2015). Protein Degradation and Instrumental Textural Changes in Fresh Nile Tilapia
724 (*Oreochromis niloticus*) during Frozen Storage. *Journal of Food Processing and Preservation*,
725 39(6), 2206-2214. doi: 10.1111/jfpp.12465
- 726 Viljanen, K., Kylli, P., Hubbermann, E. M., Schwarz, K., & Heinonen, M. (2005). Anthocyanin antioxidant
727 activity and partition behavior in whey protein emulsion. *Journal of Agricultural and Food*
728 *Chemistry*, 53(6), 2022-2027. doi: 10.1021/jf047975d
- 729 Xiong, Y. L. (2000). Protein oxidation and implications for muscle foods quality. *Book chapter. Editors :*
730 *Decker, E.; Faustman, C.; Lopez-Bote, C. J.*, pp.85-111 ref.163.
- 731 Yin, X., Luo, Y., Fan, H., Wu, H., & Feng, L. (2014). Effect of previous frozen storage on quality changes
732 of grass carp (*Ctenopharyngodon idellus*) fillets during short-term chilled storage. *International*
733 *Journal of Food Science & Technology*, 49(6), 1449-1460. doi: 10.1111/ijfs.12431
- 734 Zhang, W. G., Xiao, S., & Ahn, D. U. (2013). Protein Oxidation: Basic Principles and Implications for Meat
735 Quality. *Critical Reviews in Food Science and Nutrition*, 53(11), 1191-1201. doi:
736 10.1080/10408398.2011.577540

737

738

739

740

741

742

743

744

745

746

747

748 **X. Figure legends:**

749 **Figure. 1.** Main factors that can influence fish flesh quality before and after storage time and
750 prefer primary and secondary targets.

751 The efficiency of parameters which affect on fish flesh quality in the time period is indicated
752 by dotted lines.

753 **Figure. 2.** The main possible interaction pathways with the target amino acids and involved
754 products of lipid and protein oxidation with respect to the ideal condition. Main impact of the
755 mentioned interaction on the final flesh quality

For Review Only

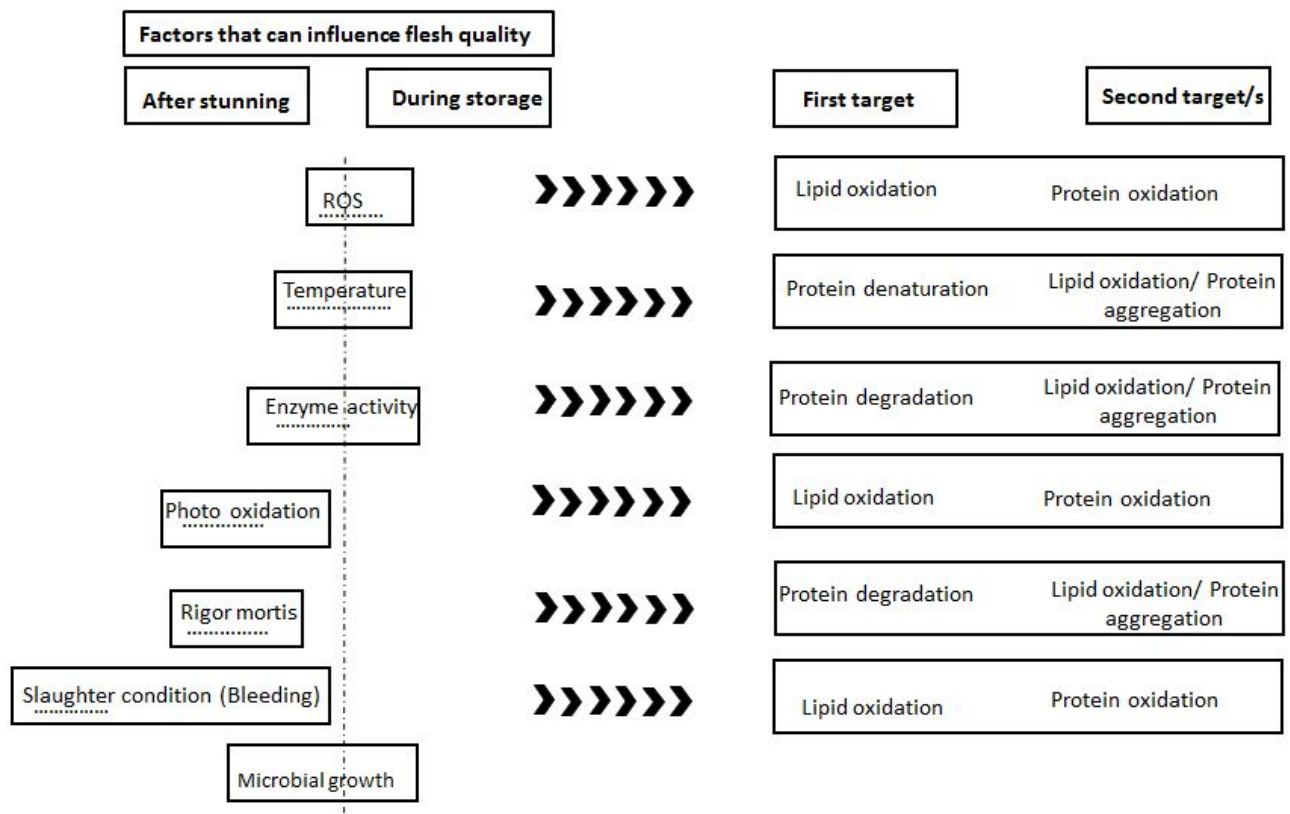


Figure. 1. Main factors that can influence fish flesh quality before and after storage time and prefer primary and secondary targets.

The efficiency of parameters, which affect fish flesh quality in the time period is indicated by dotted lines.

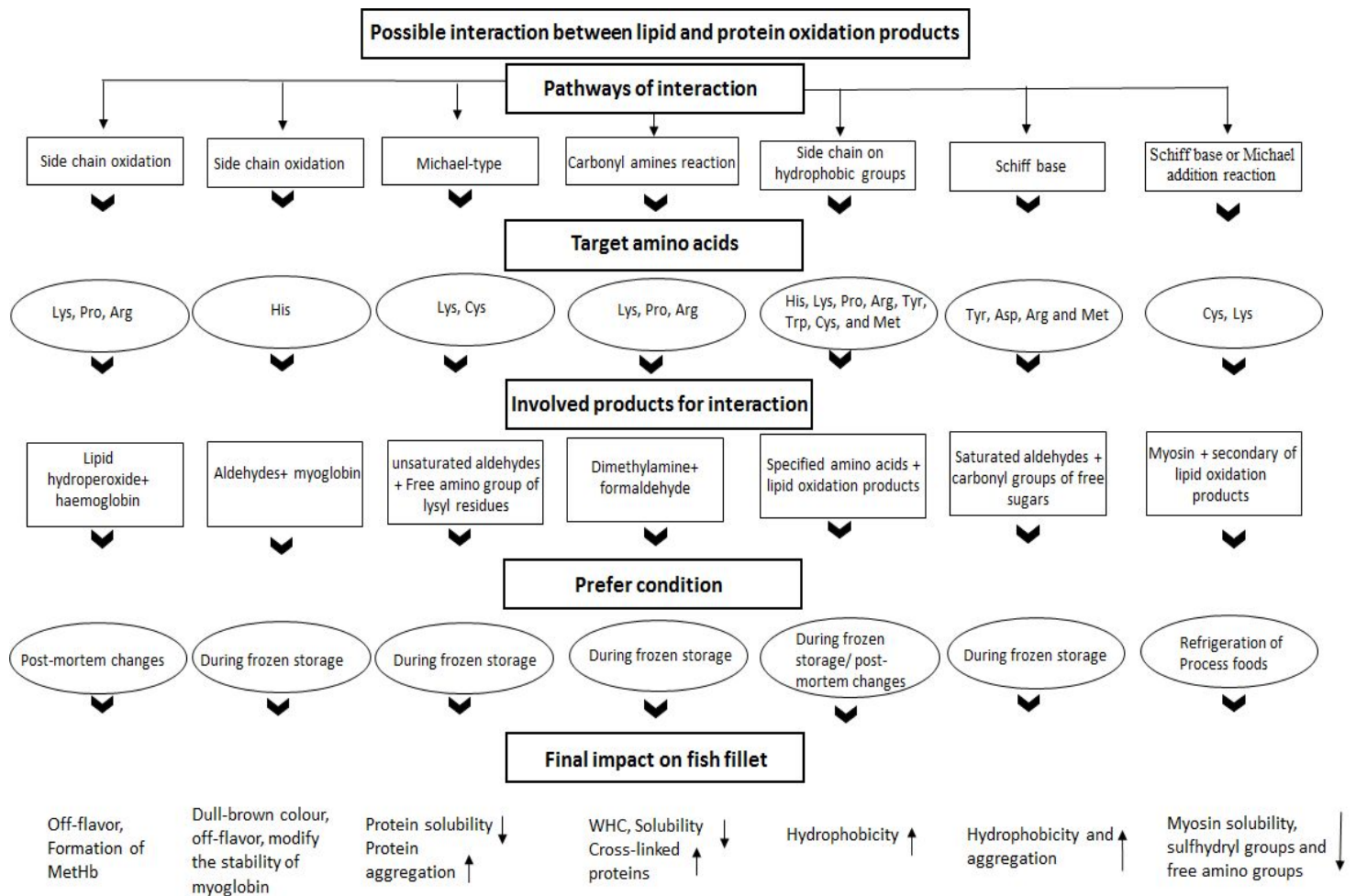


Figure. 2. The main possible interaction pathways with the target amino acids and involved products of lipid and protein oxidation with respect to the ideal condition. Main impact of the mentioned interaction on the final flesh quality

For Review Only