

1.TITLE PAGE

1.1.Title:

Gammadelta T cells in Crohn's disease: a new player in the disease pathogenesis?

1.2.Authors:

1. Catalan-Serra, Ignacio

ignaciocatalan76@hotmail.com

Hospital Arnau de Vilanova de Valencia, Aparato Digestivo
Valencia, Comunidad Valenciana, ES +4798135353
Norges teknisk-naturvitenskapelige universitet, Department of Cancer
Research and Molecular Medicine Trondheim, NO
Centre of Molecular Inflammation Research (CEMIR), Norwegian
University of Science and Technology (NTNU), CEMIR, Trondheim, NO
Sykehuset Levanger, Helse Nord-Trøndelag.
Medicine. Gastroenterology, Levanger, NO

2. Sandvik, Arne Kristian

arne.sandvik@ntnu.no

Norwegian University of Science and Technology, Department of
Cancer Research and Molecular Medicine
Trondheim, NO
Centre of Molecular Inflammation Research, Norwegian University of
Science and Technology, Trondheim, Norway, CEMIR
Trondheim, NO
St. Olav's University Hospital, Gastroenetrology
Trondheim, NO

3. Bruland, Torunn

torunn.bruland@ntnu.no

Norwegian University of Science and Technology, Department of
Cancer Research and Molecular Medicine
Trondheim, NO

4. Andreu-Ballester, Juan Carlos

jcandreu@ono.com

Arnau de Vilanova Hospital, Research Unit
Valencia, ES

1.3.Short Title:

Gammadelta T cells in Crohn's disease.

1.4.Adress of correspondence:

- Ignacio Catalan-Serra
- Mail adress: ignaciocatalan76@hotmail.com
- Levanger Hospital, Levanger, Norway. Kirkegata 2, 7600, Levanger, Norway
- Phone number: +47 98135353 / +47 74098000

2.ABSTRACT

Crohn's disease (CD) is a chronic relapsing systemic disease affecting the gastrointestinal tract. An altered immune response to commensal intestinal bacteria takes place in genetically predisposed individuals, resulting in chronic inflammation in the gut. CD can be considered an immune deficiency condition and several alterations in the innate immunity mechanisms have been described in the recent years. Thus, the study of the immunological aspects of CD, specifically the role of lymphocytes, is a key element for understanding the pathogenesis of the disease.

Gammadelta T cells constitute only a small proportion of the lymphocytes that circulate in the blood and peripheral organs and they are present mainly in the epithelia, where they can constitute up to 50% of intraepithelial T cells (IELs) in the mucosa. Due to their lack of MHC restriction and their unique plasticity and immune regulating properties they are considered key cells in the first line of defense against infections and in wound healing. However, their clinical role in IBD, including CD, is largely unknown.

In this review, we attempt to address the possible involvement of gamamdelta T cells in the pathogenesis of CD, reviewing their role against infections and in inflammation, the current evidence in animal and human studies and hypothesis for their involvement in the disease.

Key words: Gammadelta T cells, Crohn's disease, pathogenesis.

3.MAIN TEXT

Crohn's Disease (CD) is a chronic recurrent inflammatory process that can affect any part of the gastrointestinal tract and may present associated extraintestinal manifestations(1, 2), leading to a significantly compromised quality of life for patients and high associated healthcare costs(3).

Despite the sharp increase in its incidence in recent decades(4) and the important advances made in understanding the mechanisms underlying CD, the etiopathogenesis of the disease remains largely unknown. It is currently considered a polygenic immune disorder due to multiple causes in which the following are involved: 1) individual genetic factors; 2) environmental factors; 3) intestinal flora (microbiome); and 4) immune response. A combination of these factors triggers an inadequate excessive immune response against the commensal flora in genetically predisposed subjects(5-8).

CD can be considered an immune deficiency condition(9-12). Several alterations in the innate immunity mechanisms have been reported in recent years, leading to the presence of micro-infections that are not efficiently solved by the immune system, thus perpetuating an exaggerated inflammatory immune response(13-21). Hence, the study of the immunological aspects of CD, and more specifically the role of lymphocytes, is a key element for understanding the pathogenesis of the disease.

$\gamma\delta$ T cells constitute only a small proportion of the lymphocytes that circulate in the blood and peripheral organs and they are present mainly in the epithelia, where they can account for up to 50% of intraepithelial T cells (IELs) in the mucosa(22, 23). Due to their lack of MHC restriction and their singular plasticity and immunoregulatory properties they are considered key cells in the first line of defence against infections and in wound healing(22)(24-26). These unique features, their known participation in the pathogenesis of other autoinflammatory conditions(27-31) and the results of animal and human studies published

to date point to an important role of $\gamma\delta$ T cells in the pathogenesis of CD(32-37), a condition characterised by the presence of recurrent infections and ulcerations in the gut (1) (2).

However, their clinical role in IBD, including CD, is still unclear. Our aim in this review is to examine the existing evidence on the role played by alterations in **gammadelta T lymphocytes ($\gamma\delta$ T cells)** as a possible new pathogenic mechanism in CD, as these cells are essential to innate immunity in the first line of defence against infection in the mucosa, and play a key role in immunoregulation and tissue repair.

1. Innate immunity alterations in CD

There is a growing body of evidence from both basic and clinical research that suggests that CD would be essentially a primary immunodeficiency (PI) rather than an autoimmune process in the proper sense of the term. That is to say, patients with CD present a genetically determined flaw in their immune system (basically in their innate or unspecific immunity), with an identifiable phenotype which, as a result, can suffer unresolved recurrent infections and chronic "autoinflammation" with the secondary appearance of autoimmune phenomena or neoplasia. This aspect has been analysed in several recent reviews (9-12).

In fact, genetic alterations have been reported in mechanisms directly involved in the recognition (like the nucleotide-binding oligomerisation domain-containing protein 2, NOD2) (38, 39) and clearance of intracellular organisms and certain bacteria (like the autophagy related protein 16L -ATG16L- (40, 41) or the immunity-related GTPase M -IRGM-)(42, 43).

Moreover, the fact that the typical lesions occur in the areas of greater bacterial density (ileum and rectum), the improvement of the inflammation in areas without faecal transit or the presence of a greater number of intramucosal bacteria and granulomas, make the presence of recurrent infections a plausible mechanism for maintaining the inflammatory response (44, 45). Proof of the presence of adherent-invasive *Escherichia coli* (AIEC) in the colon and ileum(46) and in the granulomas of patients(47) would support this possibility.

In addition, there are other disorders in innate immune mechanisms that contribute to the appearance of recurrent infections and colonic inflammation in CD, such as: 1) alterations in the mucosal layer(13, 14); 2) alterations in the intestinal permeability(15-17); 3) dysfunction of Paneth cells (and the production of defensins)(18, 19) and of macrophage functioning(20, 21); or 4) alterations in the stress mechanisms of the endoplasmic reticulum(48, 49).

2.Gammadelta T cells

2.1.General aspects of $\gamma\delta$ T cells

T cells can be divided into two large sub-populations according to the antigen receptor that they express on the surface membrane (T-cell receptor or TCR): alphabeta T cells ($\alpha\beta$ T cells) and gammadelta T cells ($\gamma\delta$ T cells) (50). TCR is a heterodimer that can be composed of two chains: α and β or γ and δ . Its expression is exclusive, that is, a T cell can only express one of these two phenotypic variants ($\alpha\beta$ T cells or $\gamma\delta$ T cells)(22, 51). This heterodimer is associated with the CD3 complex (T-cell marker) in the cell membrane(50).

Since the discovery of $\gamma\delta$ T cells in 1984 by Saito et al.(52), there has been a growing interest in revealing the biological functions of this lymphocytes. Today, they are considered a key element in the first line of defence against invasive pathogens in the epithelia, as well as in the homeostasis of the immune response(22, 24).

$\alpha\beta$ T cells are the largest population in peripheral blood, where they account for up to 95% of the circulating T cells, and respond exclusively against antigens that are processed and presented by antigen-presenting cells (APC) in molecules of the major histocompatibility complex (MHC). Their response is therefore restricted and delayed(22).

There are three fundamental characteristics that differentiate $\gamma\delta$ T cells from $\alpha\beta$ T cells (*Figure 1*):

1) $\gamma\delta$ T cells are mainly located in the mucosa – between the epithelial cells – accounting for up to 50% of the intraepithelial lymphocytes (IEL) and acting as the first line of mucosal immune defence; they are scarce in peripheral blood (PB) and the secondary lymphoid organs(22, 23). During infections, however, the proportion of $\gamma\delta$ T cells in PB may increase considerably, to the extent where they can make up to 40-60% of the circulating T cells(53, 54). This increase can remain for up to four months following some infections(55).

2) $\gamma\delta$ T cells recognise proteins directly without the need for prior antigenic processing by APC or their presentation in the MHC molecules(56, 57). They are also able to recognise a wide range of non-peptide ligands such as viral proteins(58), bacterial superantigens - such as staphylococcal enterotoxin A -(59), lipid antigens(60), heat shock proteins (HSP)(61), or MICA/B molecules (MHC-class I related molecules) inducible by cell stress(62).

3) $\gamma\delta$ T cells are powerfully stimulated directly by phosphorylated microbial metabolites or phosphoantigens(63, 64). Among these, their most powerful activator capable of stimulating $\gamma\delta$ T cells at nanomolecular concentrations is HMBPP (E-4-hydroxy-3-methyl-but-2-enyl-pyrophosphate), a sub-product of the synthesis of isoprenoid by the non-mevalonate pathway synthesised by bacteria and protozoa(60). They are also activated at much higher concentrations by isopentenyl pyrophosphate (IPP), a derivative produced by eukaryotic cells via the mevalonate metabolic pathway(60, 65). Aminobisphosphonates (ABP) such as alendronate, pamidronate or zoledronate (currently used in clinical practice for the treatment and prevention of osteoporosis) as well as alkylamines have been employed for the in vivo and in vitro activation of these lymphocytes, since they increase the intracellular concentration of secondary PA by inhibiting the enzyme of the mevalonate pathway farnesyl-diphosphate synthase(66-68). Some synthetic phosphoantigen derivatives such as bromohydrin pyrophosphate BrHPP (Phosphostim®), have been successfully used to stimulate $\gamma\delta$ T cells ex vivo(69).

These specific characteristics and their great plasticity enable them to offer a swift response against a wide range of antigens, making $\gamma\delta$ T cells a key element in the defence against infections, tumors, and in the regulation of the immune response in many chronic and autoimmune inflammatory diseases(70, 71). They are thus considered to be "bridge" cells between innate (or unspecific) and acquired (or specific) immunity(72).

In fact, and most important for clinical practice, several lines of research are currently using these cells in antineoplastic and anti-infectious immunotherapy with promising results.(67, 73).

2.2.Subtypes and phenotypic classification

Gammadelta T cells are the first T cells to develop in vertebrates and the first to appear in the foetal thymus(22). They make up the greater sub-population of T cells during the first year of life, which is suggestive of their key role in neonatal protection while the IgA protective system is still not fully developed(71, 74).

Gammadelta T cells can be classified into two main populations, according to TCR expression: $V\delta 1+$ and $V\delta 2+$ (also known as $V\gamma 9V\delta 2$ because this is its predominant phenotype) (22).

$V\delta 1+$ are predominant in epithelia: skin ($V\gamma 5\delta 1$), intestine ($V\gamma 8\delta 1$) and genitourinary tract ($V\gamma 6\delta 1$)(28, 75, 76). They usually express the marker $CD8+$ (77), present characteristics of motility and migration to the mucous membranes (adherence, emission of long filopodia, etc.)(78, 79) and play a crucial role in epithelial regeneration(80).

$V\delta 2+$ ($V\gamma 9V\delta 2$) are the predominant subtype in peripheral blood(81) and possess a greater cytotoxic capacity of the natural killer (NK) and antibody-mediated types(82, 83). These features made them most suitable for immunotherapy against cancer (67, 84). Recent studies reveal a significant presence of a third subtype, $V\delta 3+$, in the intestinal mucosa.(85).

Most of the peripheral $\gamma\delta$ T cells are double negative for CD4 and CD8 ($CD4-CD8-$)(23).

However, intestinal $\gamma\delta$ T cells ($\gamma\delta$ IELi) frequently express the marker CD8+ (50% of $\gamma\delta$ intraepithelial lymphocytes), and the homodimer CD8 $\alpha\alpha$, an extrathymic differentiation marker, is commonly expressed(86).

A variable percentage of peripheral $\gamma\delta$ T cells express CD28 (co-stimulator of $\alpha\beta$ T cells)(87), CD40L (which entails its capacity to interact with B lymphocytes)(88), or NK cell receptors such as NKG2D (a mediator of cytolytic activity)(89).

In recent years other populations of $\gamma\delta$ T cells have also been reported, such as IL-17-producing $\gamma\delta$ T cells, which seem to play an essential role in the pathogenesis of certain autoimmune diseases(28, 90), and $\delta 2$ CD56+ TL, with a greater cytolytic and anti-infectious capacity(91).

2.3. Specific actions: cytotoxicity, immunoregulation and tissue repair.

In recent years other specific functions of $\gamma\delta$ T cells that reinforce their key role in immunoregulation and tissue regeneration have been reported(71, 92) (*Figure 2*):

1) Direct cytotoxic action against infected or tumorous cells via the secretion of perforins and granzyme B(93), independent of antibodies(94). They are even capable of opsonising and engulfing infected cells directly(95).

2) Secretion of proinflammatory cytokines, above all Th1 type, which are essential for controlling intracellular viruses and bacteria (such as IFN- γ or TNF- α), or extracellular bacteria and fungi (such as IL-17)(96, 97).

3) Activation of the immune response locally, thereby promoting the maturation of dendritic cells and the anti-infection response of macrophages and NK cells(98, 99) and increasing the capacity to resist invasion of epithelial cells(100).

4) Stimulation and regulation of innate and acquired immunity. $\gamma\delta$ T cells can regulate other cells involved in the innate immunity response by producing immunosuppressant cytokines like TGF- β or IL-10(70). Furthermore, they can migrate to the secondary lymphoid organs and collaborate with B lymphocytes in the production of antibodies via the production of CXCR5 (CXC chemokine receptor 5)(101, 102); regulate the activity of $\alpha\beta$ T cells(103) (in fact mice with $\gamma\delta$ T-cell deficiency present an exaggerated response to $\alpha\beta$ T cells); or eliminate regulatory T cells (Treg)(104).

5) They can act as antigen-presenting cells. They can process and present antigens of pathogens (infected cells, viruses) to other immune cells, such as $\alpha\beta$ T cells(105, 106).

6) Epithelial regeneration and wound healing: by stimulating the production of hyaluronic acid by epithelial cells (which is deposited in the extracellular matrix and attracts monocytes and macrophages)(25) or by means of the production of epithelial growth factors – such as insulin-like growth factor (IGF-1) or keratinocyte growth factor 1 and 2 (KGF-1, KGF-2)(26).

2.4.The role of $\gamma\delta$ T cells in defence against infections

There is strong evidence for the fundamental role of $\gamma\delta$ T cells in the immune response against pathogens (excellent reviews elsewhere) (24).

The antiviral effects of $\gamma\delta$ T cells has been confirmed in several acute and chronic viral infections, both by attack on infected cells and by the production of antiviral cytokines, especially interferon gamma (IFN- γ)(107, 108). A number of studies have confirmed the protective role of $\gamma\delta$ T cells in infection by the influenza virus(109), human immunodeficiency virus (HIV) (110, 111) cytomegalovirus (CMV) (112), or West Nile virus(113). They also play a role in the control of infection due to *Epstein Barr* virus (EBV) (110) and in the suppression of tumour growth induced by human polyomavirus (HPV)(114).

Also, although there is a decrease in the population of V γ 9V δ 2 in peripheral blood in patients with chronic infection by hepatitis C virus (HCV)(115), it has been suggested that the infiltration of activated V γ 1 lymphocytes into hepatic tissue could accelerate disease progression(116).

$\gamma\delta$ T cells are capable of recognising a number of bacterial antigens and of triggering a rapid immune response(76). There is evidence of their participation in the regulation of the immune response in a number of bacterial infections, including salmonellosis, brucellosis, legionellosis, tularemia, listeria and infections by *Escherichia coli*(24). Infection by *Mycobacterium tuberculosis*, produces an expansion and activation of V γ 9V δ 2 T cells(117) and mice lacking $\gamma\delta$ T cells undergo an increased and sustained granulomatous inflammatory response following tuberculosis infection via aerosol(118) reinforcing their importance in the pathogenesis of the disease. A recent study showed a deficit of $\gamma\delta$ T cells in the PB of patients with sepsis, with a significant correlation between low levels and mortality(119).

These cells have a protective role against other pathogens like malaria (120), *Toxoplasma Gondii* ileitis (121), *Trypanosoma cruzi* hepatic infection via IFN- γ (122) or against *Cryptosporidium parvum* (123). Their action against fungi has also been demonstrated by an early and significant increase in $\gamma\delta$ T cells following infection by *Encephalitozoon cuniculi*(124).

3.The role of $\gamma\delta$ T cells in intestinal inflammation:

3.1.Preclinical studies

There is a large body of evidence of the protective role of $\gamma\delta$ T cells in intestinal inflammation in several murine models (32-34). This is consistent with their role against mucosal infections, tissue repair, and regulation of the immune response. They play a role in oral tolerance(125) and can enhance IgA-mediated responses (126) or contribute to tissue healing by means of the production of KGF(127) (128). In addition they can exert protective

immunoregulatory effects – such as producing TGF- β (32), decreasing the expression of MHC II molecules(128) or suppressing $\alpha\beta$ T-cell activity (129) (*Table 1*).

Tsuchiya et al. and Chen et al. used dextrane sulfate sodium DSS-induced colitis models to test the anti-inflammatory effects of $\gamma\delta$ T cells, showing that large numbers of $\gamma\delta$ T cells (but not $\alpha\beta$ T cells) were found in the intestinal damaged areas, and were able to protect from the colitis by stimulating tissue repair by means of KGF expression and controlling infiltration by neutrophils(32, 130). Other studies used colitis induced by intrarectal TNBS (2,4,6-trinitrobenzene sulphonic acid) to show that the passive transfer of $\alpha\beta$ T cells (and not $\gamma\delta$ T cells) was capable of inducing colitis(131).

Studies by Hoffmann and Kühl et al. demonstrated the protective role of the inflammation of $\gamma\delta$ T cells in different murine models of IBD. The same researchers also used a TNBS-induced colitis model to prove that depletion of $\gamma\delta$ T cells with monoclonal antibodies (and not $\alpha\beta$ T cells or T-helper cells) increased the severity of colitis and even mortality(36). This aggravation of the lesions with increased mortality was corroborated in later work(132). They also used TNF Δ ARE/+ mice that present a transmural ileitis similar to CD with extraintestinal manifestations (arthritis) showing that $\gamma\delta$ T cells depletion caused a histological aggravation – without statistical significance -and lower levels of TGF- β – (132).

A later study confirmed their role as a protector against colitis in two murine models of ulcerative colitis (DSS and IL-2 k.o. mice), and a distinct (although non-significant) histological improvement in a model similar to CD (TNF Δ ARE/+). The authors showed that $\gamma\delta$ T cells are also capable of controlling the production of IFN- γ by $\alpha\beta$ T cells and of stimulating epithelial regeneration(37).

Two key studies by Inagaki-Ohara et al. and Hoffmann et al. opened up the possibility of using $\gamma\delta$ T cells as immunological therapy in IBD after showing that their selective transfer improves inflammation and even decreases mortality in mice, through the diminished production of TNF- α and the increase in IL-10 and TGF- β (33) (35).

Nevertheless, although several preclinical studies have revealed the importance of $\gamma\delta$ T cells as agents protecting against inflammation, there is no common agreement on their pro- or anti-inflammatory role, as inflammation-inducing effects have also been reported in some murine models of colitis(133).

Simpson et al. were the first to suggest that $\gamma\delta$ T cells might contribute to intestinal inflammation, showing that their infusion in mice lacking these cells (tg Σ 26 mice) was capable of producing colitis via a Th1 response and the production of IFN- γ (134). In the same line, others showed proinflammatory effects of these cells in TCR α -/- mice (135).

Recent reports have offered evidence of the existence of a specific subtype of IL-17A-producing $\gamma\delta$ T cells with proinflammatory actions in immunocompromised mice (136). This phenomenon had already been confirmed earlier in models of autoimmune encephalomyelitis(97) and in collagen-induced arthritis. This subtype of $\gamma\delta$ T cells could aggravate IL-17-mediated colitis by lack of suppression by Treg(137).

The main preclinical studies of $\gamma\delta$ T cells in animal models are shown in *Table 2*.

3.2.Role of $\gamma\delta$ T cells in Crohn's disease: studies in humans

Several groups have conducted studies on $\gamma\delta$ T cells in peripheral blood (PB) and in the intestinal mucosa of patients with CD in an attempt to explain their role "in situ" in humans (*Table 3*).

3.2.1.Serum values of lymphocytes and $\gamma\delta$ T cells in Crohn's Disease

The vast majority of studies of lymphocytes in PB in patients with CD show the presence of significant lymphopenia, with heterogeneous results as regards the circulating $\gamma\delta$ T cells.

The first evidence of the presence of lymphopenia in patients with CD was reported in the 1970s. Using antiserum against human thymocytes and a rosette assay utilising ram erythrocytes, respectively, Strickland et al. and Sorensen et al. were the first to report a decrease of T cells in the PB of patients with CD and normal levels in patients with UC(138)

(139). The authors point to differences in the pathogenesis of the two diseases as a possible explanation for this. Yet these differences have not been confirmed in later studies(140).

Selby et al. employed monoclonal antibodies for the first time to identify lymphocyte sub-populations in 54 patients with IBD (28 UC and 26 CD), showing a significant decrease of T cells both in CD and in active UC (with lower levels of both CD4+ T cells and CD8+ T cells in CD with respect to controls) (140). This decrease in circulating T cells in patients with CD was confirmed in a later study that included 43 patients(141). A recent study of circulating lymphocytes in a broad sample of patients with CD confirmed the presence of lymphopenia regardless of the clinical activity and the use of treatment(142).

The impact of the presence of lymphopenia in CD and its possible effect on the appearance of secondary autoimmune phenomena (which might explain, for example, the appearance of extraintestinal manifestations) must be taken into account(143). The association between lymphopenia and autoimmunity is well known in other autoimmune diseases -such as rheumatoid arthritis (RA), systemic lupus erythematosus (SLE) or Sjogren's disease-, and the induction of lymphopenia is used in several animal models of autoimmunity (143) (144) (145). Sustained lymphopenia can give rise to a peripheral homeostatic lymphocyte proliferation (PHLP) to maintain the peripheral T cell population stable, with the subsequent loss of diversity of the TCR repertoire and the appearance of autoreactive effector T cells(143). Two animal studies conducted in mice, on autoimmune diabetes(146) and autoimmune pancreatitis models(147) support this hypothesis. It has recently been suggested that states of lymphopenia could also alter the balance between regulatory T cells (Treg) and effector T cells (Tef), thereby inducing states of autoreactivity via IL-2 and IL-21(148) (149). However, two studies were including a total of 40 patients with CD did not find any significant lymphopenia(150) (151).

Several previous studies analyse the serum values of $\gamma\delta$ T cells in CD. Giacomelli et al. observed a significant increase in $\gamma\delta$ T cells in serum in CD only in activity, especially by

expansion of the subtype V δ 1+ in a small sample of nine patients(151). Bucht et al. also found higher values of $\gamma\delta$ T cells (with a relative increase in the subtype V δ 1+) in three patients with CD with respect to controls(152), a finding that was confirmed in a later study carried out by the same group(153).

We published recently a study comparing the serum levels of different lymphocyte subpopulations in 40 patients with CD and 40 controls. Results showed the presence of lymphopenia and a pronounced deficit of $\gamma\delta$ T cells in the patients in all the subtypes studied (CD4+ $\gamma\delta$ T cells, CD8+ $\gamma\delta$ T cells and CD56+ $\gamma\delta$ T cells), especially significant in the CD8+ $\gamma\delta$ T cells subset, which was independent of disease activity (remission or active disease) and of the treatments used(142). These data established for the first time the possible existence of an immune disorder in CD that affects this sub-population and contributes to the appearance of recurrent infections, thus corroborating the data obtained in experimental studies in mice(32-34). This deficit of $\gamma\delta$ T cells in PB has recently been confirmed in a later study conducted by our group involving a larger sample of 102 patients with CD with respect to controls (186).

A recent study showed similar results, with significantly lower levels of V δ 2+ T cells in 12 paediatric patients with moderate CD without any immunosuppressant treatment in comparison with controls with irritable bowel syndrome (IBS) (154). Despite presenting fewer circulating V δ 2+, a selective depletion of the subtype CD27+ (related with Th1 response and TNF- α producers) was observed, which was found to be increased at the same time in the biopsies of colon tissue(154). In addition, the same study showed the selective ablation of V δ 2+ cells in azathioprine treated patients.

Similar results have been reported in patients with CD undergoing treatment with infliximab, who presented lower levels of $\gamma\delta$ T cells in blood than the controls (1.6% of the total number of T cells vs. 2.6% in controls), although without reaching any statistically significant differences(155).

It is striking that none of the studies published reported an increase in the total number of $\gamma\delta$ T cells (or of the subtype $V\delta 2+$) in PB in patients in activity. It is known that this population increases significantly following infections, with values that can reach up to 40-50% of the total T cells(156). Hence, a secondary expansion could be expected in the active phases of the disease (in which there is an increase in permeability and recurrent micro-infections). This lack of expansion of the $\gamma\delta$ T cells in active CD is consistent with the deficiency observed in these studies and would support the notion of an altered immune response of the $\gamma\delta$ T cells in CD(155) (157). In fact, preliminary data from our series suggest an inverse relationship between the $\gamma\delta$ T cell values and the clinical and endoscopic disease activity(186).

3.2.2. Study of $\gamma\delta$ T cells in the intestinal mucosa in Crohn's Disease

Three studies have confirmed a decrease of $\gamma\delta$ T cells in the intestinal mucosa of patients with CD(158) (152) (159).

Fukushima et al. were the first to observe a significant decrease of $\gamma\delta$ T cells in the mucosa in CD. The intraepithelial lymphocytes (IEL)/ $\gamma\delta$ T cell ratio was 13% in CD vs. 36% in controls, and that of the lamina propria lymphocytes (LPL) was 4% in CD vs. 15% in controls. Most of the LPL were $CD4(-)CD8(-)$ (158).

Another study showed as well a decrease in $\gamma\delta$ T cells (sub-population $V\delta 1+$) in the inflamed mucosa with respect to the healthy mucosa in four patients. The authors interpreted that these findings may be due to the destruction of the epithelial barrier or to a local expansion of $\gamma\delta$ T cells in the inflamed mucosa(152).

Lee et al. reported a significant decrease in the sub-populations $CD3+$, $\alpha\beta$ TCR+, and $\gamma\delta$ TCR+ in the intestinal mucosa of patients with CD with respect to controls. A significant decrease in $\gamma\delta$ T cells was also observed in patients with CD compared to patients with UC, which would suggest a different role of this sub-population in the two diseases(159). Another study found no significant differences in the proportion of $\gamma\delta$ T cells and the LPL in 14 patients with ileal CD(160). Conversely, other authors found an increase in $V\delta 1+$ $\gamma\delta$ T cells

capable of producing IFN- γ and interacting with fibroblasts in inflamed areas with respect to non-inflamed areas in a broad series of surgical samples of CD, UC and diverticulitis(161).

Another recent study explored for the first time the molecules responsible for the intestinal migration of $\gamma\delta$ T cells in the PBoF patients with CD and active UC. It was shown that both the $\gamma\delta$ T cells of healthy subjects and active IBD express the molecule $\beta 7$, responsible for migration to the intestine, and that patients with IBD present an increased expression of CCR9 (corresponding chemokine receptor-9), a specific marker of migration to the small intestine(162). These data agree with those of the studies carried out by McCarthy et al. that showed that activated V $\delta 2^+$ $\gamma\delta$ T cells display enhanced gut-homing potential and can populate the human intestinal mucosa, thereby confirming that recruitment does indeed take place from PB to the inflamed areas(162) (157).

4.Gammadelta T cells in clinical therapy

$\gamma\delta$ T cells have been used in several clinical trials, especially as anti-tumour immunotherapy. $\gamma\delta$ T cells possess a powerful cytotoxic capacity independent of MHC that enables them to act against haematopoietic and solid tumours(67). In fact, $\gamma\delta$ T cells have been isolated from the intratumoral lymphocytes (ITL) of patients with colorectal, kidney, prostate, ovarian and lung tumours(163).

Ligands expressed in tumour and non-healthy cells (such as NKG2D or MIC/B)(164, 165) would bind the TCR activating thereby their anti-tumour action by means of the secretion of IFN- γ and TNF- α , cytotoxins such as granzyme B and perforins(93, 94), or by stimulating antibody-dependent cytotoxic activity(166, 167).

Two strategies have been tried to use $\gamma\delta$ T cells as anti-tumour immunotherapy: 1) transfer of *ex vivo* activated autologous V $\gamma 9$ V $\delta 2$; and 2) induction of the expansion of V $\gamma 9$ V $\delta 2$ *in vivo*(67, 168).

The anti-tumour action of *ex vivo* activated autologous V γ 9V δ 2 transfer has been shown to be well tolerated and effective, especially in advanced tumours, such as renal cancer(169) (170). The other alternative with promising results is to induce the expansion of V γ 9V δ 2 in vivo. The use of zoledronate and low doses of IL-2 in patients with metastatic breast cancer and metastatic prostate cancer was well tolerated and produced a sustained activation and maturation of $\gamma\delta$ T cells, which also correlated with better clinical responses(171) (165).

Research is currently being conducted to explore the activation of $\gamma\delta$ T cells through the use of dendritic cells (DC), since these are able to increase their cytotoxic and anti-tumour capacity(172).

Although there is less clinical experience in this regard, $\gamma\delta$ T cells have also been used successfully in the treatment of some infectious diseases. Activated $\gamma\delta$ T cells can improve immunocompetence in antiretroviral-naive HIV-infected patients, thus opening up new therapeutic possibilities in the infection(173). In another phase II study, repeated treatment with phosphoantigens reduced the viral load by up to 50% in patients with chronic infection by the hepatitis C virus (HCV), probably due to the increased production of IFN- γ .

Currently there are still not any clinical studies testing these cells in intestinal inflammation diseases in humans, like IBD.

5.The role of $\gamma\delta$ T cells in CD: friend or foe?

The singular characteristics of $\gamma\delta$ T cells, their great plasticity and the possibility of their use in immunotherapy has stimulated their study in the pathogenesis of CD. Yet, their role as a protector or inducer of intestinal inflammation in the disease today remains insufficiently clear.

There is good evidence on the protective role of $\gamma\delta$ T cells in intestinal inflammation in several murine models, in line with their protective capacity against infections, in the regeneration of the mucosa tissue and as a regulator of immunity (32-36). The fact that the

depletion of $\gamma\delta$ T cells in induced colitis aggravates the lesions and that their transfer produces a histological improvement and increased survival in mice is suggestive of their antiinflammatory role(35) (33).

Other studies, however, point to their possible proinflammatory role(134) (135), especially of the IL-17-producing subtype of $\gamma\delta$ T cells in CD(136). This phenomenon has previously been reported in models of autoimmune encephalomyelitis(97) and in collagen-induced arthritis(174).

Apparently contradictory phenomena, such as the capacity of murine $\gamma\delta$ T cells to produce IFN- γ (134) and to inhibit the production of IFN- γ by $\alpha\beta$ T cells(37), need to be investigated further in order to understand the plasticity of these cells in the different contexts of infection or inflammation.

Another aspect to be taken into account is the heterogeneity of the murine models used in the experimental studies of IBD and their reproducibility in such a heterogeneous disease as CD. In this sense, Kühl et al. suggest the possibility that the mice models utilised in many studies were not valid for studying the behaviour of $\gamma\delta$ T cells. They argue that α -chain deficient mice perhaps produce $\gamma\delta$ T cells that are altered or which have a greater capacity for cytotoxicity or IFN- γ production(132) (175). This would explain their proinflammatory effect in some studies, since, as has been shown, the development and correct functioning of $\gamma\delta$ T cells is partly dependent upon $\alpha\beta$ T cells(176).

Some data suggest that there may be a primary deficiency of this cell type in patients with CD, in line with other previously reported defects in the innate immune defence mechanisms in the disease(13-19) (21) (48) (177). In fact, several studies have reported lower values of $\gamma\delta$ T cells in the PB of these patients(142) (155) (154), independently of the clinical activity and the use or not of specific treatment(142). The presence of low levels of IL-7 (one of their most powerful stimulants) in patients with CD both in remission and with active disease(119) with a tendency to return to normal values in remission(178) would support the hypothesis

that this $\gamma\delta$ T deficiency is characteristic of the disease and could have clinical consequences, favouring the appearance of recurrent infections that could perpetuate the intestinal inflammation.

In this regard, it is to note the recent report of an opportunistic infection by an intracellular micro-organism of the fungi family, Microsporidia, in patients with CD(119), which may take advantage of this defect in cell immunity and, specifically, in $\gamma\delta$ T cells(179) (124). In fact, this study also showed an inverse correlation between IgE anti-Encephalitozoon (a subspecies of microsporidia) antibodies and $\gamma\delta$ T cell levels, especially the subtype CD8+ $\gamma\delta$ T cells(142).

In contrast, other authors noted an increase in $\gamma\delta$ T cells in patients with CD, especially in activity, and of the subtype V δ 1+ (151-153). In line with this, McCarthy et al. provided evidence of a selective depletion of the subtype CD27+ (related with Th1 response, and with high expression of intestinal migration β 7 integrins), which was simultaneously increased in colon biopsies(157). Thus, they propose the attractive hypothesis that, despite having a $\gamma\delta$ T-cell deficiency in PB (at least in paediatric CD patients), some cells with a high degree of activation (capable of producing TNF- α , IL-17A and stimulating the production of IFN- β by $\alpha\beta$ T cells) managed to migrate to the inflamed areas in the intestine(157).

Although no clear data are available about the clinical correlation of these alterations of $\gamma\delta$ T cells in PB, a recent study conducted by our group in a comprehensive sample of patients with CD reveals an inverse correlation between the serum values of $\gamma\delta$ T cells (and not $\alpha\beta$ T cells) and clinical and endoscopic activity (own data, not published). This would reinforce the relation between $\gamma\delta$ T-cell deficiency and the more severe forms of the disease, and a possible predicting role of its determination.

There is currently no general agreement about the location and the role of $\gamma\delta$ T cells in the intestinal mucosa in CD, which makes it more difficult to interpret their role on a local level. Some authors detected a decrease in the $\gamma\delta$ T-cell population in the biopsies of patients compared to controls, especially in the ileum(158) (159). Others, however, found no

significant differences in the percentage of IEL or of $\gamma\delta^+$ TCR LPL(160), or even observed an increase of them in colon biopsies, although without reaching statistical significance(180). Nevertheless, most of the studies showed a decrease in $\gamma\delta$ T cells in the inflamed vs. non-inflamed mucosa, although in a very limited number of patients(152) (161).

This contradiction between the pro- and anti-inflammatory functions of $\gamma\delta$ T cells is not exclusive to CD, since similar phenomena have been reported in other chronic inflammatory diseases such as multiple sclerosis(27) (28), Behcet's disease(29) or systemic lupus erythematosus(30) (31).

For example, IL-17-producing $\gamma\delta$ T cells can aggravate and induce inflammation in models of rheumatoid arthritis(181), psoriasis or ankylosing spondylitis(182), whereas, in contrast, this sub-population has been shown to play a protective role in the development of diabetes in NOD mice via the production of TGF- β (183). Likewise, it has been shown that $\gamma\delta$ T cells can perform an anti-tumour function or favour tumoral growth in different immunological contexts(73).

Another possible explanation for these discrepancies is the possibility that the different subtypes of $\gamma\delta$ T cells may have distinct actions and effects. The subtype V δ 2 $^+$ is predominant in PB(81), and has a greater cytotoxic capacity of both the natural killer (NK) and antibody-mediated types(82) (83). The V δ 1 $^+$ subtype predominates in the epithelia(75), has a lower cytotoxic capacity and plays a crucial role in epithelial regeneration(78-80).

A relative increase in V δ 1 $^+$ $\gamma\delta$ T cells has been demonstrated not only in patients with CD(180) (152), but also in the inflamed mucosa of patients with UC, with a direct relationship between their number and the severity of the inflammation(184). It appears that the V δ 1 $^+$ subtype may play an important role in chronic inflammation in the epithelia, since similar findings have been reported in active coeliac disease(185), in the skin lesions of leprosy(187) or in the synovial tissue of patients with rheumatoid arthritis(75).

Some authors also point to the possibility that exposure of the different types of $\gamma\delta$ T cells in different cytokine micro-environments in inflamed tissues would produce activation of distinct genes, with opposing effects. That is to say, the diverse interactions of $\gamma\delta$ T cells with the microbiota, epithelial cells and other cells in the immune system (such as macrophages) would result in a different immune response.

6. Conclusions and future directions.

Crohn's disease is a chronic recurrent systemic disease affecting any part of the gastrointestinal tract, typically the ileocecal area. An altered immune response to commensal intestinal bacteria takes place in genetically predisposed individuals, resulting in chronic inflammation. Several alterations in the innate immunity mechanisms have been reported in recent years, leading to the presence of micro-infections that are not efficiently solved by the immune system, leading to chronic inflammation.

$\gamma\delta$ T cells constitute only a small proportion of the lymphocytes that circulate in the blood and peripheral organs and they are present mainly in the epithelia, where they can account for up to 50% of intraepithelial T cells (IELs) in the mucosa. Their lack of MHC restriction and their singular plasticity and immunoregulatory properties makes them key cells in the first line of defence against infections and in wound healing. These unique features, their known participation in the pathogenesis of other autoinflammatory conditions and the results of animal and human studies published to date point to an important role of $\gamma\delta$ T cells in the pathogenesis of CD. However, their clinical role in IBD is still not fully understood. Some animal studies indicate that they may play a protective role against colitis, while others have shown a possible deleterious role. In humans, there is evidence that patients with CD present a $\gamma\delta$ T-cell deficiency in PB, with possible clinical consequences. Other studies, however, have identified proinflammatory phenotypes that could contribute to intestinal inflammation.

These contradictory effects must be studied in depth in order to know the biology of this type of lymphocyte, its interaction with the microbiota, the epithelial cells and its relation with other innate and acquired immunity cells. A systematic study of the different sub-populations of $\gamma\delta$ T cells ($V\delta 1+$, $V\delta 2+$, $V\delta 3+$), as well as their different phenotypes (IL-17+ $\gamma\delta$, $\gamma\delta$ Tregs, etc.), within the different clinical contexts of a heterogeneous disease like CD will allow some of these key issues to be solved.

The study of the relations between the values in serum and in the mucosa (both inflamed and healthy), together with the migration mechanisms, their possible prognostic value, their correlation with opportunistic infections, the effect of treatments and their role in the different phenotypic forms of CD (including the extraintestinal manifestations) will help to define better their true importance in the pathogenesis and clinical symptoms.

Furthermore, we could also speculate on the possibility of using immunoregulatory treatments to modulate their action and improve the propensity to present infections, by optimising the immune response in the intestinal mucosa in CD. In fact, clinical trials have already been conducted that show that manipulating these cells in immunotherapy against cancer is effective and well tolerated, thereby opening up a new possible therapeutic alternative in the future for CD patients.

5.FUNDING

No fundig has contributed to the preparation of this manuscript.

6.ACKNOWLEDGEMENTS AND AUTHORS STATEMENTS

Catalan-Serra, Ignacio has contributed in the conception and design of the study, acquisition and analysis of data, drafting the article or revising it critically for important intellectual content and final approval of the version to be submitted.

Sandvik, Arne Kristian has contributed in the analysis and interpretation of data, drafting the article and revising it critically for important intellectual content and final approval of the version to be submitted.

Bruland, Torunn has contributed in the analysis and interpretation of data, drafting the article and revising it critically for important intellectual content and final approval of the version to be submitted.

Andreu-Ballester, Juan Carlos has contributed in the conception and design of the study, acquisition and analysis of data, drafting the article or revising it critically for important intellectual content and final approval of the version to be submitted.

7.REFERENCES

1. Abraham C, Cho JH. Inflammatory bowel disease. *N Engl J Med* 2009;361:2066-78.
2. Baumgart DC, Sandborn WJ. Crohn's disease. *Lancet* 2012;380:1590-605.
3. Rocchi A, Benchimol EI, Bernstein CN, Bitton A, Feagan B, Panaccione R, et al. Inflammatory bowel disease: a Canadian burden of illness review. *Can J Gastroenterol* 2012;26:811-7.
4. Lakatos PL. Recent trends in the epidemiology of inflammatory bowel diseases: up or down? *World J Gastroenterol* 2006;12:6102-8.
5. Boyapati R, Satsangi J, Ho GT. Pathogenesis of Crohn's disease. *F1000Prime Rep* 2015;7:44.
6. Xavier RJ, Podolsky DK. Unravelling the pathogenesis of inflammatory bowel disease. *Nature* 2007;448:427-34.
7. Butto LF, Schaubek M, Haller D. Mechanisms of Microbe-Host interaction in Crohn's Disease: Dysbiosis vs. Pathobiont Selection. *Front Immunol* 2015;6.
8. de Souza HS, Fiocchi C. Immunopathogenesis of IBD: current state of the art. *Nat Rev Gastroenterol Hepatol* 2016;13:13-27.
9. Huang Y, Chen Z. Inflammatory bowel disease related innate immunity and adaptive immunity. *Am J Transl Res* 2016;8:2490-7.
10. Vinh DC, Behr MA. Crohn's as an immune deficiency: from apparent paradox to evolving paradigm. *Expert Rev Clin Immunol* 2013;9:17-30.
11. Glocker E, Grimbacher B. Inflammatory bowel disease: is it a primary immunodeficiency? *Cell Mol Life Sci* 2012;69:41-8.
12. Marks DJ. Defective innate immunity in inflammatory bowel disease: a Crohn's disease exclusivity? *Curr Opin Gastroenterol* 2011;27:328-34.
13. Buisine MP, Desreumaux P, Debailleul V, Gambiez L, Geboes K, Ectors N, et al. Abnormalities in mucin gene expression in Crohn's disease. *Inflammatory Bowel Diseases* 1999;5:24-32.

14. Franke A, McGovern DP, Barrett JC, Wang K, Radford-Smith GL, Ahmad T, et al. Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nat Genet* 2010;42:1118-25.
15. Pearson AD, Eastham EJ, Laker MF, Craft AW, Nelson R. Intestinal permeability in children with Crohn's disease and coeliac disease. *Br Med J (Clin Res Ed)* 1982;285:20-1.
16. Zeissig S, Burgel N, Gunzel D, Richter J, Mankertz J, Wahnschaffe U, et al. Changes in expression and distribution of claudin 2, 5 and 8 lead to discontinuous tight junctions and barrier dysfunction in active Crohn's disease. *Gut* 2007;56:61-72.
17. Goetz M, Kiesslich R. Advances of endomicroscopy for gastrointestinal physiology and diseases. *Am J Physiol Gastrointest Liver Physiol* 2010;298:G797-806.
18. Wehkamp J, Salzman NH, Porter E, Nuding S, Weichenthal M, Petras RE, et al. Reduced Paneth cell alpha-defensins in ileal Crohn's disease. *Proc Natl Acad Sci U S A* 2005;102:18129-34.
19. Peyrin-Biroulet L, Beisner J, Wang G, Nuding S, Oommen ST, Kelly D, et al. Peroxisome proliferator-activated receptor gamma activation is required for maintenance of innate antimicrobial immunity in the colon. *Proc Natl Acad Sci U S A* 2010;107:8772-7.
20. Smith AM, Rahman FZ, Hayee B, Graham SJ, Marks DJ, Sewell GW, et al. Disordered macrophage cytokine secretion underlies impaired acute inflammation and bacterial clearance in Crohn's disease. *J Exp Med* 2009;206:1883-97.
21. Levine AP, Segal AW. What is wrong with granulocytes in inflammatory bowel diseases? *Dig Dis* 2013;31:321-7.
22. Chien YH, Meyer C, Bonneville M. gammadelta T cells: first line of defense and beyond. *Annu Rev Immunol* 2014;32:121-55.
23. Andreu-Ballester JC, Garcia-Ballesteros C, Benet-Campos C, Amigo V, Almela-Quilis A, Mayans J, et al. Values for alphabeta and gammadelta T-lymphocytes and CD4+, CD8+, and CD56+ subsets in healthy adult subjects: assessment by age and gender. *Cytometry B Clin Cytom* 2012;82:238-44.
24. Zheng J, Liu Y, Lau YL, Tu W. gammadelta-T cells: an unpolished sword in human anti-infection immunity. *Cell Mol Immunol* 2013;10:50-7.
25. Jameson JM, Cauvi G, Sharp LL, Witherden DA, Havran WL. Gammadelta T cell-induced hyaluronan production by epithelial cells regulates inflammation. *J Exp Med* 2005;201:1269-79.

26. Jameson JM, Sharp LL, Witherden DA, Havran WL. Regulation of skin cell homeostasis by gamma delta T cells. *Front Biosci* 2004;9:2640-51.
27. Blink SE, Caldis MW, Goings GE, Harp CT, Malissen B, Prinz I, et al. gammadelta T cell subsets play opposing roles in regulating experimental autoimmune encephalomyelitis. *Cell Immunol* 2014;290:39-51.
28. Paul S, Shilpi, Lal G. Role of gamma-delta (gammadelta) T cells in autoimmunity. *J Leukoc Biol* 2015;97:259-71.
29. Hasan MS, Bergmeier LA, Petrushkin H, Fortune F. Gamma Delta (gammadelta) T Cells and Their Involvement in Behcet's Disease. *J Immunol Res* 2015;2015:705831.
30. Li X, Kang N, Zhang X, Dong X, Wei W, Cui L, et al. Generation of human regulatory gammadelta T cells by TCRgammadelta stimulation in the presence of TGF-beta and their involvement in the pathogenesis of systemic lupus erythematosus. *J Immunol* 2011;186:6693-700.
31. Ponomarev ED, Dittel BN. Gamma delta T cells regulate the extent and duration of inflammation in the central nervous system by a Fas ligand-dependent mechanism. *J Immunol* 2005;174:4678-87.
32. Chen YP, Chou K, Fuchs E, Havran WL, Boismenu R. Protection of the intestinal mucosa by intraepithelial gamma delta T cells. *P Natl Acad Sci USA* 2002;99:14338-43.
33. Inagaki-Ohara K, Chinen T, Matsuzaki G, Sasaki A, Sakamoto Y, Hiromatsu K, et al. Mucosal T cells bearing TCRgammadelta play a protective role in intestinal inflammation. *J Immunol* 2004;173:1390-8.
34. Egan CE, Maurer KJ, Cohen SB, Mack M, Simpson KW, Denkers EY. Synergy between intraepithelial lymphocytes and lamina propria T cells drives intestinal inflammation during infection. *Mucosal Immunol* 2011;4:658-70.
35. Hoffmann JC, Pawlowski NN, Grollich K, Loddenkemper C, Zeitz M, Kuhl AA. Gammadelta T lymphocytes: a new type of regulatory T cells suppressing murine 2,4,6-trinitrobenzene sulphonic acid (TNBS)-induced colitis. *Int J Colorectal Dis* 2008;23:909-20.
36. Hoffmann JC, Peters K, Henschke S, Herrmann B, Pfister K, Westermann J, et al. Role of T lymphocytes in rat 2,4,6-trinitrobenzene sulphonic acid (TNBS) induced colitis: increased mortality after gammadelta T cell depletion and no effect of alphabeta T cell depletion. *Gut* 2001;48:489-95.

37. Kuhl AA, Pawlowski NN, Grollich K, Loddenkemper C, Zeitz M, Hoffmann JC. Aggravation of intestinal inflammation by depletion/deficiency of gammadelta T cells in different types of IBD animal models. *J Leukoc Biol* 2007;81:168-75.
38. Hugot JP, Chamaillard M, Zouali H, Lesage S, Cezard JP, Belaiche J, et al. Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 2001;411:599-603.
39. Ogura Y, Bonen DK, Inohara N, Nicolae DL, Chen FF, Ramos R, et al. A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. *Nature* 2001;411:603-6.
40. Hampe J, Franke A, Rosenstiel P, Till A, Teuber M, Huse K, et al. A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1. *Nat Genet* 2007;39:207-11.
41. Rioux JD, Xavier RJ, Taylor KD, Silverberg MS, Goyette P, Huett A, et al. Genome-wide association study identifies new susceptibility loci for Crohn disease and implicates autophagy in disease pathogenesis. *Nat Genet* 2007;39:596-604.
42. Singh SB, Davis AS, Taylor GA, Deretic V. Human IRGM induces autophagy to eliminate intracellular mycobacteria. *Science*.2006;313:1438-41.
43. Brest P, Lapaquette P, Souidi M, Lebrigand K, Cesaro A, Vouret-Craviari V, et al. A synonymous variant in IRGM alters a binding site for miR-196 and causes deregulation of IRGM-dependent xenophagy in Crohn's disease. *Nat Genet* 2011;43:242-5.
44. Bosca-Watts MM, Tosca J, Anton R, Mora M, Minguez M, Mora F. Pathogenesis of Crohn's disease: Bug or no bug. *World J Gastrointest Pathophysiol* 2015;6:1-12.
45. Haag LM, Siegmund B. Intestinal Microbiota and the Innate Immune System - A Crosstalk in Crohn's Disease Pathogenesis. *Front Immunol* 2015;6:489.
46. Masseret E, Boudeau J, Colombel JF, Neut C, Desreumaux P, Joly B, et al. Genetically related *Escherichia coli* strains associated with Crohn's disease. *Gut* 2001;48:320-5.
47. Ryan P, Kelly RG, Lee G, Collins JK, O'Sullivan GC, O'Connell J, et al. Bacterial DNA within granulomas of patients with Crohn's disease--detection by laser capture microdissection and PCR. *Am J Gastroenterol* 2004;99:1539-43.
48. Kaser A, Lee AH, Franke A, Glickman JN, Zeissig S, Tilg H, et al. XBP1 links ER stress to intestinal inflammation and confers genetic risk for human inflammatory bowel disease. *Cell* 2008;134:743-56.

49. Kaser A, Martinez-Naves E, Blumberg RS. Endoplasmic reticulum stress: implications for inflammatory bowel disease pathogenesis. *Curr Opin Gastroenterol* 2010;26:318-26.
50. Allison TJ, Winter CC, Fournie JJ, Bonneville M, Garboczi DN. Structure of a human gammadelta T-cell antigen receptor. *Nature* 2001;411:820-4.
51. Wu YL, Ding YP, Tanaka Y, Shen LW, Wei CH, Minato N, et al. gammadelta T cells and their potential for immunotherapy. *Int J Biol Sci* 2014;10:119-35.
52. Saito H, Kranz DM, Takagaki Y, Hayday AC, Eisen HN, Tonegawa S. Complete primary structure of a heterodimeric T-cell receptor deduced from cDNA sequences. *Nature* 1984;309:757-62.
53. Hara T, Mizuno Y, Takaki K, Takada H, Akeda H, Aoki T, et al. Predominant activation and expansion of V gamma 9-bearing gamma delta T cells in vivo as well as in vitro in Salmonella infection. *J Clin Invest* 1992;90:204-10.
54. Perera MK, Carter R, Goonewardene R, Mendis KN. Transient increase in circulating gamma/delta T cells during Plasmodium vivax malarial paroxysms. *J Exp Med* 1994;179:311-5.
55. Bertotto A, Gerli R, Spinozzi F, Muscat C, Scalise F, Castellucci G, et al. Lymphocytes bearing the gamma delta T cell receptor in acute Brucella melitensis infection. *Eur J Immunol* 1993;23:1177-80.
56. Schild H, Mavaddat N, Litzemberger C, Ehrich EW, Davis MM, Bluestone JA, et al. The nature of major histocompatibility complex recognition by gamma delta T cells. *Cell* 1994;76:29-37.
57. Chien YH, Jores R, Crowley MP. Recognition by gamma/delta T cells. *Annu Rev Immunol* 1996;14:511-32.
58. Bukowski JF, Morita CT, Brenner MB. Recognition and destruction of virus-infected cells by human gamma delta CTL. *Gamma-Delta-Ctl* . *J Immunol* 1994;153:5133-40
59. Tanaka Y, Sano S, Nieves E, De Libero G, Rosa D, Modlin RL, et al. Nonpeptide ligands for human gamma delta T cells. *Proc Natl Acad Sci U S A* 1994;91:8175-9.
60. Tanaka Y, Morita CT, Tanaka Y, Nieves E, Brenner MB, Bloom BR. Natural and synthetic non-peptide antigens recognized by human gamma delta T cells. *Nature* 1995;375:155-8.
61. Multhoff G, Botzler C. Heat-shock proteins and the immune response. *Ann N Y Acad Sci* 1998;851:86-93.
62. Groh V, Steinle A, Bauer S, Spies T. Recognition of stress-induced MHC molecules by intestinal epithelial gammadelta T cells. *Science* 1998;279:1737-40.

63. Pfeffer K, Schoel B, Gulle H, Kaufmann SH, Wagner H. Primary responses of human T cells to mycobacteria: a frequent set of gamma/delta T cells are stimulated by protease-resistant ligands. *Eur J Immunol* 1990;20:1175-9.
64. Gomes AQ, Martins DS, Silva-Santos B. Targeting gammadelta T lymphocytes for cancer immunotherapy: from novel mechanistic insight to clinical application. *Cancer Res* 2010;70:10024-7.
65. Nedellec S, Bonneville M, Scotet E. Human Vgamma9Vdelta2 T cells: from signals to functions. *Semin Immunol* 2010;22:199-206.
66. Thompson K, Rojas-Navea J, Rogers MJ. Alkylamines cause Vgamma9Vdelta2 T-cell activation and proliferation by inhibiting the mevalonate pathway. *Blood* 2006;107:651-4.
67. Chiplunkar S, Dhar S, Wesch D, Kabelitz D. gammadelta T cells in cancer immunotherapy: current status and future prospects. *Immunotherapy* 2009;1:663-78.
68. Kunzmann V, Bauer E, Wilhelm M. Gamma/delta T-cell stimulation by pamidronate. *N Engl J Med* 1999;340:737-8.
69. Bennouna J, Bompas E, Neidhardt EM, Rolland F, Philip I, Galea C, et al. Phase-I study of Innacell gammadelta, an autologous cell-therapy product highly enriched in gamma9delta2 T lymphocytes, in combination with IL-2, in patients with metastatic renal cell carcinoma. *Cancer Immunol Immunother* 2008;57:1599-609.
70. Bonneville M, O'Brien RL, Born WK. Gammadelta T cell effector functions: a blend of innate programming and acquired plasticity. *Nat Rev Immunol* 2010;10:467-78.
71. Vantourout P, Hayday A. Six-of-the-best: unique contributions of gammadelta T cells to immunology. *Nat Rev Immunol* 2013;13:88-100.
72. Kabelitz D. gammadelta T-cells: cross-talk between innate and adaptive immunity. *Cell Mol Life Sci* 2011;68:2331-3.
73. Silva-Santos B, Serre K, Norell H. gammadelta T cells in cancer. *Nat Rev Immunol* 2015;15:683-91.
74. Kagnoff MF. Current concepts in mucosal immunity. III. Ontogeny and function of gamma delta T cells in the intestine. *Am J Physiol* 1998;274:G455-8.
75. Soderstrom K, Bucht A, Halapi E, Lundqvist C, Gronberg A, Nilsson E, et al. High expression of V gamma 8 is a shared feature of human gamma delta T cells in the epithelium of the gut and in the inflamed synovial tissue. *J Immunol* 1994;152:6017-27.

76. Ferreira LM. Gammadelta T cells: innately adaptive immune cells? *Int Rev Immunol* 2013;32:223-48.
77. Deusch K, Luling F, Reich K, Classen M, Wagner H, Pfeffer K. A major fraction of human intraepithelial lymphocytes simultaneously expresses the gamma/delta T cell receptor, the CD8 accessory molecule and preferentially uses the V delta 1 gene segment. *Eur J Immunol* 1991;21:1053-9.
78. Grossi CE, Ciccone E, Migone N, Bottino C, Zarcone D, Mingari MC, et al. Human T cells expressing the gamma/delta T-cell receptor (TcR-1): C gamma 1- and C gamma 2-encoded forms of the receptor correlate with distinctive morphology, cytoskeletal organization, and growth characteristics. *Proc Natl Acad Sci U S A* 1989;86:1619-23.
79. Kabelitz D. Function and specificity of human gamma/delta-positive T cells. *Crit Rev Immunol* 1992;11:281-303.
80. Toulon A, Breton L, Taylor KR, Tenenhaus M, Bhavsar D, Lanigan C, et al. A role for human skin-resident T cells in wound healing. *J Exp Med* 2009;206:743-50.
81. Brenner MB, McLean J, Scheft H, Riberdy J, Ang SL, Seidman JG, et al. Two forms of the T-cell receptor gamma protein found on peripheral blood cytotoxic T lymphocytes. *Nature* 1987;325:689-94.
82. Sturm E, Braakman E, Fisch P, Vreugdenhil RJ, Sondel P, Bolhuis RL. Human V gamma 9-V delta 2 T cell receptor-gamma delta lymphocytes show specificity to Daudi Burkitt's lymphoma cells. *J Immunol* 1990;145:3202-8.
83. Dastot H, Schmid M, Gontier C, Amiot M, Mathieu-Mahul D, Bensussan A, et al. Correlation between T cell receptor gamma delta isotypic forms and cytotoxic activity: analysis with human T cell clones and lines. *Cell Immunol* 1990;125:315-25.
84. Braza MS, Klein B. Anti-tumour immunotherapy with Vgamma9Vdelta2 T lymphocytes: from the bench to the bedside. *Br J Haematol* 2013;160:123-32.
85. Dunne MR, Elliott L, Hussey S, Mahmud N, Kelly J, Doherty DG, et al. Persistent changes in circulating and intestinal gammadelta T cell subsets, invariant natural killer T cells and mucosal-associated invariant T cells in children and adults with coeliac disease. *PLoS One* 2013;8:e76008.
86. Lefrancois L, LeCorre R, Mayo J, Bluestone JA, Goodman T. Extrathymic selection of TCR gamma delta + T cells by class II major histocompatibility complex molecules. *Cell* 1990;63:333-40.

87. Sperling AI, Linsley PS, Barrett TA, Bluestone JA. CD28-mediated costimulation is necessary for the activation of T cell receptor-gamma delta+ T lymphocytes. *J Immunol* 1993;151:6043-50.
88. Horner AA, Jabara H, Ramesh N, Geha RS. gamma/delta T lymphocytes express CD40 ligand and induce isotype switching in B lymphocytes. *J Exp Med* 1995;181:1239-44.
89. Bauer S, Groh V, Wu J, Steinle A, Phillips JH, Lanier LL, et al. Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. *Science* 1999;285:727-9.
90. Kisielow J, Kopf M. The origin and fate of gammadeltaT cell subsets. *Curr Opin Immunol* 2013;25:181-8.
91. Qin G, Liu Y, Zheng J, Xiang Z, Ng IH, Malik Peiris JS, et al. Phenotypic and functional characterization of human gammadelta T-cell subsets in response to influenza A viruses. *J Infect Dis* 2012;205:1646-53.
92. Born WK, Reardon CL, O'Brien RL. The function of gammadelta T cells in innate immunity. *Curr Opin Immunol* 2006;18:31-8.
93. Qin G, Mao H, Zheng J, Sia SF, Liu Y, Chan PL, et al. Phosphoantigen-expanded human gammadelta T cells display potent cytotoxicity against monocyte-derived macrophages infected with human and avian influenza viruses. *J Infect Dis* 2009;200:858-65.
94. Alexander AA, Maniar A, Cummings JS, Hebbeler AM, Schulze DH, Gastman BR, et al. Isopentenyl pyrophosphate-activated CD56+ {gamma}{delta} T lymphocytes display potent antitumor activity toward human squamous cell carcinoma. *Clin Cancer Res* 2008;14:4232-40.
95. Himoudi N, Morgenstern DA, Yan M, Vernay B, Saraiva L, Wu Y, et al. Human gammadelta T lymphocytes are licensed for professional antigen presentation by interaction with opsonized target cells. *J Immunol* 2012;188:1708-16.
96. Tsuji M, Mombaerts P, Lefrancois L, Nussenzweig RS, Zavala F, Tonegawa S. Gamma delta T cells contribute to immunity against the liver stages of malaria in alpha beta T-cell-deficient mice. *Proc Natl Acad Sci U S A* 1994;91:345-9.
97. Sutton CE, Lalor SJ, Sweeney CM, Brereton CF, Lavelle EC, Mills KH. Interleukin-1 and IL-23 induce innate IL-17 production from gammadelta T cells, amplifying Th17 responses and autoimmunity. *Immunity* 2009;31:331-41.
98. Wands JM, Roark CL, Aydintug MK, Jin N, Hahn YS, Cook L, et al. Distribution and leukocyte contacts of gammadelta T cells in the lung. *J Leukoc Biol* 2005;78:1086-96.

99. Maniar A, Zhang X, Lin W, Gastman BR, Pauza CD, Strome SE, et al. Human gammadelta T lymphocytes induce robust NK cell-mediated antitumor cytotoxicity through CD137 engagement. *Blood* 2010;116:1726-33.
100. Jameson J, Havran WL. Skin gammadelta T-cell functions in homeostasis and wound healing. *Immunol Rev* 2007;215:114-22.
101. Ansel KM, Ngo VN, Hyman PL, Luther SA, Forster R, Sedgwick JD, et al. A chemokine-driven positive feedback loop organizes lymphoid follicles. *Nature* 2000;406:309-14.
102. Schaerli P, Willimann K, Lang AB, Lipp M, Loetscher P, Moser B. CXC chemokine receptor 5 expression defines follicular homing T cells with B cell helper function. *J Exp Med* 2000;192:1553-62.
103. Hayday A, Tigelaar R. Immunoregulation in the tissues by gammadelta T cells. *Nat Rev Immunol.* 2003;3:233-42.
104. Huber SA. gammadelta T lymphocytes kill T regulatory cells through CD1d. *Immunology* 2010;131:202-9.
105. Cheng L, Cui Y, Shao H, Han GC, Zhu L, Huang YF, et al. Mouse gamma delta T cells are capable of expressing MHC class II molecules, and of functioning as antigen-presenting cells *J Neuroimmunol.* 2008;203:3-11.
106. Brandes M, Willimann K, Bioley G, Levy N, Eberl M, Luo M, et al. Cross-presenting human gammadelta T cells induce robust CD8+ alphabeta T cell responses. *Proc Natl Acad Sci U S A* 2009;106:2307-12.
107. Poccia F, Agrati C, Martini F, Mejia G, Wallace M, Malkovsky M. Vgamma9Vdelta2 T cell-mediated non-cytolytic antiviral mechanisms and their potential for cell-based therapy. *Immunol Lett* 2005;100:14-20.
108. Selin LK, Santolucito PA, Pinto AK, Szomolanyi-Tsuda E, Welsh RM. Innate immunity to viruses: control of vaccinia virus infection by gamma delta T cells. *J Immunol* 2001;166:6784-94.
109. Qin G, Liu Y, Zheng J, Ng IH, Xiang Z, Lam KT, et al. Type 1 responses of human Vgamma9Vdelta2 T cells to influenza A viruses. *J Virol* 2011;85:10109-16.
110. De Paoli P, Gennari D, Martelli P, Cavarzerani V, Comoretto R, Santini G. Gamma delta T cell receptor-bearing lymphocytes during Epstein-Barr virus infection. *J Infect Dis* 1990;161:1013-6.
111. Wallace M, Bartz SR, Chang WL, Mackenzie DA, Pauza CD, Malkovsky M. Gamma delta T lymphocyte responses to HIV. *Clin Exp Immunol* 1996;103:177-84.

112. Khairallah C, Netzer S, Villacreces A, Juzan M, Rousseau B, Dulanto S, et al. gammadelta T cells confer protection against murine cytomegalovirus (MCMV). *PLoS Pathog* 2015;11:e1004702.
113. Fang H, Welte T, Zheng X, Chang GJ, Holbrook MR, Soong L, et al. gammadelta T cells promote the maturation of dendritic cells during West Nile virus infection. *FEMS Immunol Med Microbiol* 2010;59:71-80.
114. Mishra R, Chen AT, Welsh RM, Szomolanyi-Tsuda E. NK cells and gammadelta T cells mediate resistance to polyomavirus-induced tumors. *PLoS Pathog* 2010;6:e1000924.
115. Par G, Rukavina D, Podack ER, Horanyi M, Szekeres-Bartho J, Hegedus G, et al. Decrease in CD3-negative-CD8dim(+) and Vdelta2/Vgamma9 TcR+ peripheral blood lymphocyte counts, low perforin expression and the impairment of natural killer cell activity is associated with chronic hepatitis C virus infection. *J Hepatol* 2002;37:514-22.
116. Agrati C, D'Offizi G, Narciso P, Abrignani S, Ippolito G, Colizzi V, et al. Vdelta1 T lymphocytes expressing a Th1 phenotype are the major gammadelta T cell subset infiltrating the liver of HCV-infected persons. *Mol Med* 2001;7:11-9.
117. Kabelitz D, Bender A, Schondelmaier S, Schoel B, Kaufmann SH. A large fraction of human peripheral blood gamma/delta + T cells is activated by *Mycobacterium tuberculosis* but not by its 65-kD heat shock protein. *J Exp Med* 1990;171:667-79.
118. D'Souza CD, Cooper AM, Frank AA, Mazzaccaro RJ, Bloom BR, Orme IM. An anti-inflammatory role for gamma delta T lymphocytes in acquired immunity to *Mycobacterium tuberculosis*. *J Immunol* 1997;158:1217-21.
119. Andreu-Ballester JC, Tormo-Calandin C, Garcia-Ballesteros C, Perez-Griera J, Amigo V, Almela-Quilis A, et al. Association of gammadelta T cells with disease severity and mortality in septic patients. *Clin Vaccine Immunol* 2013;20:738-46.
120. Costa G, Loizon S, Guenot M, Mocan I, Halary F, de Saint-Basile G, et al. Control of *Plasmodium falciparum* erythrocytic cycle: gammadelta T cells target the red blood cell-invasive merozoites. *Blood*. 2011;118(26):6952-62.
121. Egan CE, Dalton JE, Andrew EM, Smith JE, Gubbels MJ, Striepen B, et al. A requirement for the Vgamma1+ subset of peripheral gammadelta T cells in the control of the systemic growth of *Toxoplasma gondii* and infection-induced pathology. *J Immunol* 2005;175:8191-9.

122. Sardinha LR, Elias RM, Mosca T, Bastos KR, Marinho CR, D'Imperio Lima MR, et al. Contribution of NK, NK T, gamma delta T, and alpha beta T cells to the gamma interferon response required for liver protection against *Trypanosoma cruzi*. *Infect Immun* 2006;74:2031-42.
123. Eichelberger MC, Suresh P, Rehg JE. Protection from *Cryptosporidium parvum* infection by gammadelta T cells in mice that lack alphabeta T cells. *Comp Med* 2000;50:270-6.
124. Moretto M, Durell B, Schwartzman JD, Khan IA. Gamma delta T cell-deficient mice have a down-regulated CD8+ T cell immune response against *Encephalitozoon cuniculi* infection. *J Immunol* 2001;166:7389-97.
125. Ke Y, Pearce K, Lake JP, Ziegler HK, Kapp JA. Gamma delta T lymphocytes regulate the induction and maintenance of oral tolerance. *J Immunol* 1997;158:3610-8.
126. Fujihashi K, Dohi T, Kweon MN, McGhee JR, Koga T, Cooper MD, et al. gammadelta T cells regulate mucosally induced tolerance in a dose-dependent fashion. *Int Immunol* 1999;11:1907-16.
127. Boismenu R, Havran WL. Modulation of epithelial cell growth by intraepithelial gamma delta T cells. *Science* 1994;266:1253-5.
128. Komano H, Fujiura Y, Kawaguchi M, Matsumoto S, Hashimoto Y, Obana S, et al. Homeostatic regulation of intestinal epithelia by intraepithelial gamma delta T cells. *Proc Natl Acad Sci U S A* 1995;92:6147-51.
129. Peng SL, McNiff JM, Madaio MP, Ma J, Owen MJ, Flavell RA, et al. alpha beta T cell regulation and CD40 ligand dependence in murine systemic autoimmunity. *J Immunol* 1997;158:2464-70.
130. Tsuchiya T, Fukuda S, Hamada H, Nakamura A, Kohama Y, Ishikawa H, et al. Role of gamma delta T cells in the inflammatory response of experimental colitis mice. *J Immunol* 2003;171:5507-13.
131. Szczepanik M, Gryglewski A, Bryniarski K, Stachura J, Ptak W. Experimental inflammatory bowel disease--role of T cells. *J Physiol Pharmacol* 2000;51:333-46.
132. Kuhl AA, Loddenkemper C, Westermann J, Hoffmann JC. Role of gamma delta T cells in inflammatory bowel disease. *Pathobiology* 2002;70:150-5.
133. Boismenu R, Chen Y, Havran WL. The role of intraepithelial gammadelta T cells: a gut-feeling. *Microbes Infect* 1999;1:235-40.

134. Simpson SJ, Hollander GA, Mizoguchi E, Allen D, Bhan AK, Wang B, et al. Expression of pro-inflammatory cytokines by TCR alpha beta+ and TCR gamma delta+ T cells in an experimental model of colitis. *Eur J Immunol* 1997;27:17-25.
135. Kawaguchi-Miyashita M, Shimada S, Kurosu H, Kato-Nagaoka N, Matsuoka Y, Ohwaki M, et al. An accessory role of TCRgammadelta (+) cells in the exacerbation of inflammatory bowel disease in TCRalpha mutant mice. *Eur J Immunol* 2001;31:980-8.
136. Do JS, Visperas A, Dong C, Baldwin WM, 3rd, Min B. Cutting edge: Generation of colitogenic Th17 CD4 T cells is enhanced by IL-17+ gammadelta T cells. *J Immunol* 2011;186:4546-50.
137. Park SG, Mathur R, Long M, Hosh N, Hao L, Hayden MS, et al. T regulatory cells maintain intestinal homeostasis by suppressing gammadelta T cells. *Immunity* 2010;33:791-803.
138. Strickland RG, Korsmeyer S, Soltis RD, Wilson ID, Williams RC, Jr. Peripheral blood T and B cells in chronic inflammatory bowel disease. *Gastroenterology* 1974;67:569-77.
139. Sorensen SF, Hoj L. Lymphocyte subpopulations in Crohn's disease and chronic ulcerative colitis. *Acta Pathol Microbiol Scand C* 1977;85:41-8.
140. Selby WS, Jewell DP. T lymphocyte subsets in inflammatory bowel disease: peripheral blood. *Gut* 1983;24:99-105.
141. Pallone F, Montano S, Fais S, Boirivant M, Signore A, Pozzilli P. Studies of peripheral blood lymphocytes in Crohn's disease. Circulating activated T cells. *Scand J Gastroenterol* 1983;18:1003-8.
142. Andreu-Ballester JC, Amigo-Garcia V, Catalan-Serra I, Gil-Borras R, Ballester F, Almela-Quilis A, et al. Deficit of gammadelta T lymphocytes in the peripheral blood of patients with Crohn's disease. *Dig Dis Sci* 2011;56:2613-22.
143. Merayo-Chalico J, Rajme-Lopez S, Barrera-Vargas A, Alcocer-Varela J, Diaz-Zamudio M, Gomez-Martin D. Lymphopenia and autoimmunity: A double-edged sword. *Hum Immunol* 2016;77:921-9.
144. Atkinson TP. Immune deficiency and autoimmunity. *Curr Opin Rheumatol* 2012;24:515-21.
145. Schulze-Koops H. Lymphopenia and autoimmune diseases. *Arthritis Res Ther* 2004;6:178-80.
146. King C, Ilic A, Koelsch K, Sarvetnick N. Homeostatic expansion of T cells during immune insufficiency generates autoimmunity. *Cell* 2004;117:265-77.

147. Le Saout C, Mennechet S, Taylor N, Hernandez J. Memory-like CD8⁺ and CD4⁺ T cells cooperate to break peripheral tolerance under lymphopenic conditions. *Proc Natl Acad Sci U S A* 2008;105:19414-9.
148. Cheyalier N, Thorburn AN, Macia L, Tan J, Juglair L, Yagita H, et al. Inflammation and Lymphopenia Trigger Autoimmunity by Suppression of IL-2-Controlled Regulatory T Cell and Increase of IL-21-Mediated Effector T Cell Expansion. *Journal of Immunology* 2014;193:4845-58.
149. Maggadottir SM, Sullivan KE. The intersection of immune deficiency and autoimmunity. *Curr Opin Rheumatol* 2014;26:570-8.
150. Senju M, Hulstaert F, Lowder J, Jewell DP. Flow cytometric analysis of peripheral blood lymphocytes in ulcerative colitis and Crohn's disease. *Gut* 1991;32:779-83.
151. Giacomelli R, Parzanese I, Frieri G, Passacantando A, Pizzuto F, Pimpo T, et al. Increase of circulating gamma/delta T lymphocytes in the peripheral blood of patients affected by active inflammatory bowel disease. *Clin Exp Immunol* 1994;98:83-8.
152. Bucht A, Soderstrom K, Esin S, Grunewald J, Hagelberg S, Magnusson I, et al. Analysis of gamma delta V region usage in normal and diseased human intestinal biopsies and peripheral blood by polymerase chain reaction (PCR) and flow cytometry. *Clin Exp Immunol* 1995;99:57-64.
153. Soderstrom K, Bucht A, Halapi E, Gronberg A, Magnusson I, Kiessling R. Increased frequency of abnormal gamma delta T cells in blood of patients with inflammatory bowel diseases. *J Immunol* 1996;156:2331-9.
154. McCarthy NE, Hedin CR, Sanders TJ, Amon P, Hoti I, Ayada I, et al. Azathioprine therapy selectively ablates human Vdelta2(+) T cells in Crohn's disease. *J Clin Invest* 2015;125:3215-25.
155. Kelsen J, Dige A, Schwindt H, D'Amore F, Pedersen FS, Agnholt J, et al. Infliximab induces clonal expansion of gammadelta-T cells in Crohn's disease: a predictor of lymphoma risk? *PLoS One* 2011;6:e17890.
156. Morita CT, Jin C, Sarikonda G, Wang H. Nonpeptide antigens, presentation mechanisms, and immunological memory of human Vgamma2Vdelta2 T cells: discriminating friend from foe through the recognition of prenyl pyrophosphate antigens. *Immunol Rev* 2007;215:59-76.
157. McCarthy NE, Bashir Z, Vossenkamper A, Hedin CR, Giles EM, Bhattacharjee S, et al. Proinflammatory Vdelta2⁺ T cells populate the human intestinal mucosa and enhance IFN-gamma production by colonic alphabeta T cells. *J Immunol* 2013;191:2752-63.

158. Fukushima K, Masuda T, Ohtani H, Sasaki I, Funayama Y, Matsuno S, et al. Immunohistochemical characterization, distribution, and ultrastructure of lymphocytes bearing T-cell receptor gamma/delta in inflammatory bowel disease. *Gastroenterology* 1991;101:670-8.
159. Lee HB, Kim JH, Yim CY, Kim DG, Ahn DS. Differences in immunophenotyping of mucosal lymphocytes between ulcerative colitis and Crohn's disease. *Korean J Intern Med* 1997;12:7-15.
160. Cuvelier CA, De Wever N, Mielants H, De Vos M, Veys EM, Roels H. Expression of T cell receptors alpha beta and gamma delta in the ileal mucosa of patients with Crohn's disease and with spondylarthropathy. *Clin Exp Immunol* 1992;90:275-9.
161. McVay LD, Li B, Biancaniello R, Creighton MA, Bachwich D, Lichtenstein G, et al. Changes in human mucosal gamma delta T cell repertoire and function associated with the disease process in inflammatory bowel disease. *Mol Med* 1997;3:183-203.
162. Mann ER, McCarthy NE, Peake ST, Milestone AN, Al-Hassi HO, Bernardo D, et al. Skin- and gut-homing molecules on human circulating gammadelta T cells and their dysregulation in inflammatory bowel disease. *Clin Exp Immunol* 2012;170:122-30.
163. Beetz S, Marischen L, Kabelitz D, Wesch D. Human gamma delta T cells: candidates for the development of immunotherapeutic strategies. *Immunol Res* 2007;37:97-111.
164. Rincon-Orozco B, Kunzmann V, Wrobel P, Kabelitz D, Steinle A, Herrmann T. Activation of V gamma 9V delta 2 T cells by NKG2D. *J Immunol* 2005;175:2144-51.
165. Dieli F, Vermijlen D, Fulfaro F, Caccamo N, Meraviglia S, Cicero G, et al. Targeting human {gamma}delta T cells with zoledronate and interleukin-2 for immunotherapy of hormone-refractory prostate cancer. *Cancer Res* 2007;67:7450-7.
166. Gertner-Dardenne J, Bonnafous C, Bezombes C, Capietto AH, Scaglione V, Ingoure S, et al. Bromohydrin pyrophosphate enhances antibody-dependent cell-mediated cytotoxicity induced by therapeutic antibodies. *Blood* 2009;113:4875-84.
167. Kobayashi KS, Chamaillard M, Ogura Y, Henegariu O, Inohara N, Nunez G, et al. Nod2-dependent regulation of innate and adaptive immunity in the intestinal tract. *Science* 2005;307:731-4.
168. Fisher JP, Heuvelink J, Yan M, Gustafsson K, Anderson J. gammadelta T cells for cancer immunotherapy: A systematic review of clinical trials. *Oncoimmunology* 2014;3:e27572.

169. Kobayashi H, Tanaka Y, Yagi J, Osaka Y, Nakazawa H, Uchiyama T, et al. Safety profile and anti-tumor effects of adoptive immunotherapy using gamma-delta T cells against advanced renal cell carcinoma: a pilot study. *Cancer Immunol Immunother* 2007;56:469-76.
170. Kobayashi H, Tanaka Y, Yagi J, Minato N, Tanabe K. Phase I/II study of adoptive transfer of gammadelta T cells in combination with zoledronic acid and IL-2 to patients with advanced renal cell carcinoma. *Cancer Immunol Immunother* 2011;60:1075-84.
171. Meraviglia S, Eberl M, Vermijlen D, Todaro M, Buccheri S, Cicero G, et al. In vivo manipulation of Vgamma9Vdelta2 T cells with zoledronate and low-dose interleukin-2 for immunotherapy of advanced breast cancer patients. *Clin Exp Immunol* 2010;161:290-7.
172. Van Acker HH, Anguille S, Van Tendeloo VF, Lion E. Empowering gamma delta T cells with antitumor immunity by dendritic cell-based immunotherapy. *Oncoimmunology* 2015;4:e1021538.
173. Poccia F, Gioia C, Martini F, Sacchi A, Piacentini P, Tempestilli M, et al. Zoledronic acid and interleukin-2 treatment improves immunocompetence in HIV-infected persons by activating Vgamma9Vdelta2 T cells. *AIDS* 2009;23:555-65.
174. Ito Y, Usui T, Kobayashi S, Iguchi-Hashimoto M, Ito H, Yoshitomi H, et al. Gamma/delta T cells are the predominant source of interleukin-17 in affected joints in collagen-induced arthritis, but not in rheumatoid arthritis. *Arthritis Rheum* 2009;60:2294-303.
175. Kohyama M, Nanno M, Kawaguchi-Miyashita M, Shimada S, Watanabe M, Hibi T, et al. Cytolytic and IFN-gamma-producing activities of gamma delta T cells in the mouse intestinal epithelium are T cell receptor-beta-chain dependent. *Proc Natl Acad Sci U S A* 1999;96:7451-5.
176. Pennington DJ, Silva-Santos B, Shires J, Theodoridis E, Pollitt C, Wise EL, et al. The inter-relatedness and interdependence of mouse T cell receptor gammadelta+ and alphabeta+ cells. *Nat Immunol* 2003;4:991-8.
177. Kaser A, Zeissig S, Blumberg RS. Inflammatory bowel disease. *Annu Rev Immunol* 2010;28:573-621.
178. Ponchel F, Cuthbert RJ, Goeb V. IL-7 and lymphopenia. *Clin Chim Acta* 2011;412:7-16.
179. Khan IA, Schwartzman JD, Kasper LH, Moretto M. CD8+ CTLs are essential for protective immunity against *Encephalitozoon cuniculi* infection. *J Immunol* 1999;162 :686-91.
180. Trejdosiewicz LK, Calabrese A, Smart CJ, Oakes DJ, Howdle PD, Crabtree JE, et al. Gamma delta T cell receptor-positive cells of the human gastrointestinal mucosa: occurrence and V region

gene expression in *Helicobacter pylori*-associated gastritis, coeliac disease and inflammatory bowel disease. *Clin Exp Immunol* 1991;84:440-4.

181. Hu C, Qian L, Miao Y, Huang Q, Miao P, Wang P, et al. Antigen-presenting effects of effector memory V γ 9V δ 2 T cells in rheumatoid arthritis. *Cell Mol Immunol* 2012;9:245-54.

182. Kenna TJ, Davidson SI, Duan R, Bradbury LA, McFarlane J, Smith M, et al. Enrichment of circulating interleukin-17-secreting interleukin-23 receptor-positive gamma/delta T cells in patients with active ankylosing spondylitis. *Arthritis Rheum* 2012;64:1420-9.

183. Han G, Wang R, Chen G, Wang J, Xu R, Wang L, et al. Interleukin-17-producing gammadelta+ T cells protect NOD mice from type 1 diabetes through a mechanism involving transforming growth factor-beta. *Immunology* 2010;129:197-206.

184. Yeung MM, Melgar S, Baranov V, Oberg A, Danielsson A, Hammarstrom S, et al. Characterisation of mucosal lymphoid aggregates in ulcerative colitis: immune cell phenotype and TcR-gammadelta expression. *Gut* 2000;47:215-27.

185. Halstensen TS, Scott H, Brandtzaeg P. Intraepithelial T cells of the TcR gamma/delta+ CD8- and V delta 1/J delta 1+ phenotypes are increased in coeliac disease. *Scand J Immunol* 1989;30:665-72.

186. Catalan-Serra I, Garcia-Ballesteros C, Gil-Borras R et al. Gammadelta T cell deficiency in the peripheral blood of patients with Crohn's disease: relationship with clinical and endoscopic activity [abstract]. In: *United European Gastroenterol J*; 2016 Oct 15–19; Vienna, Austria: UEG; 2016. Abstract nr 1351.

187. Uyemura K, Ho CT, Ohmen JD, Rea TH, Modlin RL. Selective expansion of V delta 1 + T cells from leprosy skin lesions. *J Invest Dermatol* 1992;99:848-52.

8.FIGURE LEGENDS

Fig 1. Main differences between $\alpha\beta$ T cells and $\gamma\delta$ T cells.

Fig 2. Immunoregulatory functions of $\gamma\delta$ T cells.

This figure shows a representation of the main intestinal epithelial cells, and the intraepithelial location of $\gamma\delta$ T cells (black square) and their main immunoregulatory functions. $\gamma\delta$ T cells can exert direct cytotoxic action against infected or neoplastic cells (93) (94) and are capable of opsonising and engulfing infected cells (95) . They can stimulate $\alpha\beta$ T cells for the production of Th1 or Th17 cytokines, or suppress their activation by secreting TGF- β or IL-10(70). In addition, $\gamma\delta$ T cells can activate the immune response in a local environment by promoting the maturation of dendritic cells and stimulating macrophages, neutrophils and NK cells (98, 99), as well as migrate to the secondary lymphoid organs and collaborate with B lymphocytes in the production of antibodies (101)(102). Epithelial regeneration and wound healing is actively promoted by $\gamma\delta$ T cells by stimulating the production of hyaluronic acid by epithelial cells (25) and epithelial growth factors (26).

9.TABLES

Table 1. Main antiinflammatory immunomodulator effects of $\gamma\delta$ T cells in preclinical studies.

Antiinflammatory functions of $\gamma\delta$ T cells

- Immunoregulation and protection of the mucosa in infections(103)
- Capacity to repair epithelial tissue via the production of KGF(127)
- Stimulation of IL-10 and TGF- β production(32) (35)
- Decreased expression of MHC II(128)
- Role in oral tolerance following the administration of antigens(125)
- Enhancement of IgA-mediated responses(126)
- Role in suppressing the proinflammatory effects of $\alpha\beta$ T cells(128)

Table 2. Main preclinical $\gamma\delta$ T-cell studies in murine colitis models.

Animal Studies	Publ. year	Mouse model	Protective / Pro-inflammatory	Main results
Szczepanik M. et al. (131)	2000	TNBS	PROTECTIVE	Passive transference of $\alpha\beta$ T cells and not $\gamma\delta$ T cells induce colitis.
Hoffmann J.C. et al. (36)	2001	TNBS	PROTECTIVE	Depletion of $\gamma\delta$ T cells with mAb (and not $\alpha\beta$ T cells) ameliorated the colitis and reduces mortality.
Chen Y. et al. (32)	2002	DSS	PROTECTIVE	$\gamma\delta$ T cells accumulate in the inflamed areas and collaborate in tissue repair through KGF.
Kühl A.A. et al. (132)	2002-2003	TNF Δ ARE/+	PROTECTIVE	Depletion of $\gamma\delta$ T cells aggravates colitis and increases IFN- γ production.
Tsuchiya T. et al. (130)	2003	DSS	PROTECTIVE	$\gamma\delta$ T cells control the migration of neutrophils.
Inagaki-Ohara K. et al. (33)	2004	C δ ^{-/-} / TNBS	PROTECTIVE	$\gamma\delta$ T cells deficient mice develop colitis. Transfer of $\gamma\delta$ T cells ameliorated TNBS induced colitis.
Kühl A.A. et al. (37)	2007	DSS/ TNF Δ ARE/+ / IL-2 ko	PROTECTIVE	Depletion of $\gamma\delta$ T cells aggravates colitis. Increased mortality after early depletion in IL-2 ko mice.
Hoffmann J.C. et al. (35)	2008	TNBS/IL-10 tg	PROTECTIVE	$\gamma\delta$ T cells transfer ameliorates TNBS induced colitis and prolonged survival.

Simpson S.J. et al. (134)	1997	tg ϵ 26	PRO-INFLAMMATORY	$\gamma\delta$ T cells infusion produces colitis.
Kawaguchi-Miyashita M. et al. (135)	2001	TCR α -/-	PRO-INFLAMMATORY	Elimination of $\gamma\delta$ T cells ameliorates colitis.
Do J.S. et al. (136)	2011	TCR $\beta\delta$ (-/-)	PRO-INFLAMMATORY	IL17 + $\gamma\delta$ T cells transfer induces a Th17 differentiation of colitogenic lymphocytes and induces colitis.

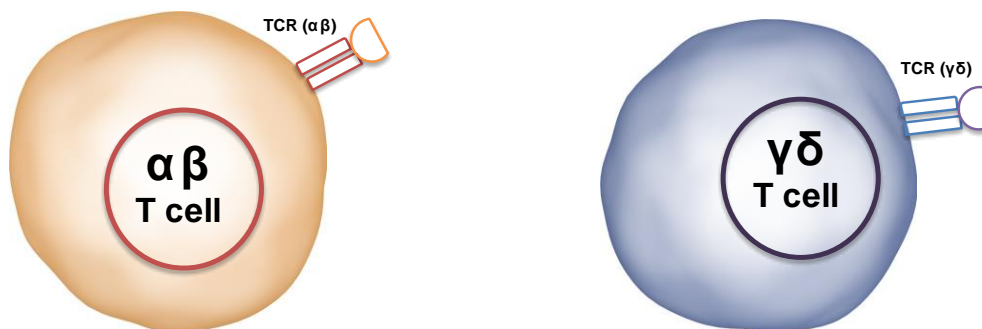
Table 3. Main human $\gamma\delta$ T cell studies in Crohn's disease.

Human Studies	Publ. year	$\gamma\delta$ T in PB/mucosa	Number of patients and disease location	Main results
Trejdosiewicz LK. et al. (180)	1991	Mucosa	6 (colonic)	Increased $\gamma\delta$ T cells percentage in 3 out of 6 CD patients (not statistically significant). Predominance of V δ 1+ subtypes
Fukushima K et al. (158)	1991	Mucosa	10 (8 ileocolonic, 1 ileal, 1 colonic)	Reduced ratio of IEL and LPL $\gamma\delta$ T cells in CD, particularly in the ileum.
Cuvelier CA. et al. (160)	1992	Mucosa	14 (ileal)	No changes in IEL/LPL $\gamma\delta$ T cells proportion in CD.
Giacomelli R. et al. (151)	1994	Blood	9 (7 ileal, 2 ileocolonic)	Increased $\gamma\delta$ T cells numbers in PB only in activ CD.
Bucht A. et al. (152)	1995	Blood/Mucosa	4 (3 colonic, 1 ileocolonic)	Increased proportion of $\gamma\delta$ T cells in PB of CD (increased V δ 1+). Decreased $\gamma\delta$ T cells in healthy mucosa compared with inflamed.
Söderström. et al. (153)	1996	Blood	16 (N/A)	Increased proportion of $\gamma\delta$ T cells in PB in CD.
Lee H.B. et al. (159)	1997	Mucosa	5 (N/A)	Decreased $\gamma\delta$ t cells percentage in CD compared with UC and controls.

McVay LD. et al. (161)	1997	Blood/Mucosa	3 (colonic)	Increased proportion of IEL and LPL $\gamma\delta$ T cells in the inflamed mucosa of CD patients vs non inflamed.
Andreu-Ballester J.C. et al. (142)	2011	Blood	40 (N/A)	Decrease $\gamma\delta$ T cell numbers in PB of CD. Especially CD8+ $\gamma\delta$ subset.
Mann E.R. et al. (162)	2012	Blood/Mucosa	15 (4 colonic, 5 ileal, 4 ileocolonic)	Increased expression of CCR9 in $\gamma\delta$ T cells of CD patients.
McCarthy N.E. et al. (154)	2015	Blood/Mucosa	12 (pediatric, N/A) 12 (N/A)	Decreased V δ 2 in the PB of CD (selective depletion of CD27+V δ 2) vs IBS controls. No differences in V δ 2 numbers in PB in CD vs healthy controls. Increased β 7+V δ 2 "gut tropic" in CD.

10.FIGURES

Figure 1.



- "Classic" T cells
- Adaptive immunity
- Retarded response
- 95% in the peripheral blood
- MHC restricted (CD4+ MHC I / CD8+ MHC II)
- Recognise processed peptides presented by APC
- Majority express CD4 or CD8
- High TCR diversity

- First described 1984 by Saito et al.
- Innate immunity
- Quick response (first line of defense)
- 3-5% in the peripheral blood (50% of the IEL in the gut mucosa)
- Not MHC restricted (direct recognition)
- Recognise unprocessed peptides, viral proteins, lipids etc.
- Majority in PB are double negative (CD4-CD8-)
- Restricted TCR diversity

Abbreviations: TCR: T cell receptor, MHC: major histocompatibility complex, APC: antigen presenting cell, PB: peripheral blood, IEL: intraepithelial lymphocytes.

Figure 2.

